

A Primer of Ecology

NICHOLAS J. GOTELLI
University of Vermont

Sinauer Associates, Inc. • Publishers
Sunderland, Massachusetts



The Cover

Third-instar larvae of the North American ant lion, *Myrmelcon immaculatus*. From *Demons of the Dust* by William Morton Wheeler (W. W. Norton, New York, 1930).

The Frontispiece

A portrayal of the complexity of ecological interactions in nature. Each chapter of the primer highlights a particular interaction from this figure. Original artwork by Shahid Naeem, University of Minnesota.

About the Book

Editor: Andrew D. Sinauer
Project Editor: Carol J. Wigg
Production Manager: Christopher Small
Book and Cover Design: Jefferson Johnson
Copy Editor: Roberta Lewis
Artwork: Precision Graphics
Book Manufacture: Courier Companies, Inc.

A PRIMER OF ECOLOGY

Copyright © 1995 by Sinauer Associates, Inc.
All rights reserved.

This book may not be reproduced in whole or in part without permission.
For information address:

Sinauer Associates, Inc., 23 Plumtree Road, Sunderland, MA 01375-0407 U.S.A.

Fax: 413-549-1118

Internet: publish@sinauer.com

Library of Congress Cataloging-in-Publication Data

Gotelli, Nicholas J., 1959-

A primer of ecology / Nicholas J. Gotelli.

p. cm.

Includes bibliographical references and index.

ISBN 0-87893-270-4 (pbk.)

1. Population biology--Mathematical models. 2. Ecology--mathematical models. I. Title

QH352.G67 1995

94-45884

574.5248'0151--dc20 CIP

Printed in U.S.A.

6 5 4 3 2 1

Dedicated to my parents, Mary and Jim

Table of Contents

Song Sparrows of Mandarte Island	43
Population Dynamics of Subtidal Ascidians	44
Logistic Growth and the Collapse of Fisheries Populations	47
PROBLEMS	50
SOLUTIONS	51

Chapter 3: Age-Structured Population Growth 55

MODEL PRESENTATION AND PREDICTIONS	56
Exponential Growth with Age Structure	56
Notation for Ages and Age Classes	56
The Fertility Schedule $[b(x)]$	57
Fertility Schedules in Nature	59
The Survivorship Schedule $[l(x)]$	59
Survival Probability $[g(x)]$	60
Survivorship Schedules in Nature	61
Calculating Net Reproductive Rate (R_0)	62
Calculating Generation Time (G)	63
Calculating Intrinsic Rate of Increase (r)	64
Describing Population Age Structure	65
Calculating Survival Probabilities for Age Classes (P_i)	66
Calculating Fertilities for Age Classes (F_i)	66
The Leslie Matrix	67
Stable and Stationary Age Distributions	69
MODEL ASSUMPTIONS	72
MODEL VARIATIONS	72
Derivation of the Euler Equation	72
Reproductive Value	73
Life History Strategies	75
Stage- and Size-Structured Population Growth	77
EMPIRICAL EXAMPLES	80
Life Tables for Ground Squirrels	80
Stage Projection Matrices for Teasel	82
PROBLEMS	86
SOLUTIONS	87

Chapter 4: Metapopulation Dynamics 89

MODEL PRESENTATION AND PREDICTIONS	90
Metapopulations and Extinction Risk	91
A Model of Metapopulation Dynamics	92

PREFACE x

ACKNOWLEDGMENTS xv

TO THE STUDENT xvi

Chapter 1: Exponential Population Growth 1

MODEL PRESENTATION AND PREDICTIONS	2
Elements of Population Growth	2
Projecting Population Size	6
Calculating Doubling Time	6
MODEL ASSUMPTIONS	9
MODEL VARIATIONS	11
Continuous versus Discrete Population Growth	11
Environmental Stochasticity	13
Demographic Stochasticity	16
EMPIRICAL EXAMPLES	19
Pheasants of Protection Island	19
Grizzly Bears of Yellowstone National Park	20
PROBLEMS	23
SOLUTIONS	24

Chapter 2: Logistic Population Growth 27

MODEL PRESENTATION AND PREDICTIONS	28
Density Dependence	28
Carrying Capacity	30
MODEL ASSUMPTIONS	32
MODEL VARIATIONS	34
Time Lags	34
Discrete Population Growth	37
Random Variation in Carrying Capacity	40
Periodic Variation in Carrying Capacity	40
EMPIRICAL EXAMPLES	43

MODEL ASSUMPTIONS	95
MODEL VARIATIONS	96
The Island–Mainland Model	96
Internal Colonization	97
The Rescue Effect	98
Other Variations	100
EMPIRICAL EXAMPLES	101
The Checkerspot Butterfly	101
Heathland Carabid Beetles	103
Oklahoma Stream Fishes	103
PROBLEMS	107
SOLUTIONS	108

Chapter 5: Competition

MODEL PRESENTATION AND PREDICTIONS	112
Competitive Interactions	112
The Lotka–Volterra Competition Model	113
Competition Coefficients	114
Equilibrium Solutions	115
The State Space	116
Graphical Solutions to the Lotka–Volterra Competition Model	119
The Principle of Competitive Exclusion	124
MODEL ASSUMPTIONS	126
MODEL VARIATIONS	127
Intraguild Predation	127
EMPIRICAL EXAMPLES	128
Competition between Intertidal Sandflat Worms	128
The Shape of a Gerbil Isocline	132
PROBLEMS	136
SOLUTIONS	137

Chapter 6: Predation

MODEL PRESENTATION AND PREDICTIONS	140
Modeling Prey Population Growth	140
Modeling Predator Population Growth	141
Equilibrium Solutions	142
Graphical Solutions to the Lotka–Volterra Predation Model	143
MODEL ASSUMPTIONS	147

MODEL VARIATIONS	147
Incorporating a Victim Carrying Capacity	148
Modifying the Functional Response	149
The Paradox of Enrichment	154
Incorporating Other Factors in the Victim Isocline	156
Modifying the Predator Isocline	157
EMPIRICAL EXAMPLES	161
Population Cycles of Hare and Lynx	161
Population Cycles of Red Grouse	162
PROBLEMS	167
SOLUTIONS	168

Chapter 7: Island Biogeography

MODEL PRESENTATION AND PREDICTIONS	172
The Species–Area Relationship	172
The Habitat Diversity Hypothesis	174
The Equilibrium Model of Island Biogeography	175
MODEL ASSUMPTIONS	181
MODEL VARIATIONS	182
Nonlinear Immigration and Extinction Curves	182
Area and Distance Effects	183
The Rescue Effect	184
The Target Effect	184
The Passive Sampling Model	185
EMPIRICAL EXAMPLES	187
Insects of Mangrove Islands	187
Breeding Birds of Eastern Wood	188
Breeding Birds of the Pymatuning Lake Islands	190
PROBLEMS	193
SOLUTIONS	194

LITERATURE CITED 196

INDEX 201

Preface

I love to read ecology textbooks. The latest ecology texts are well-written and entertaining to read. They cover all aspects of ecology from population growth to ecosystem ecology and conservation biology. They present students with a balanced mix of theoretical, empirical, and applied topics, supported by a vast bibliography of hundreds of literature citations—everything from the “textbook classics” to the latest cutting-edge research. All this material is packaged in an attractive format, with color photographs, sophisticated graphics, and eye-pleasing type fonts. The downside is encyclopedic length and a hefty price tag for the student.

Despite their massive size, the new texts often fail in helping students with the single most difficult aspect of ecology courses: understanding mathematical models. Many texts exclude or dilute the mathematical and quantitative material, leaving students with a product that has been intellectually gutted. More traditional texts (and instructors) that do cover mathematical models also err by assuming the mathematical details are self-evident, glossing over the derivations, and failing to explicitly and concisely state the assumptions and predictions of the models.

My own pet peeve is the treatment of the exponential model of population growth. The exponential model is the basis for most population and community models, and is often used to introduce students to concepts such as continuous versus discrete population growth, population size (N), growth rate (dN/dt), and per capita growth rate $[(1/N)(dN/dt)]$. Without a firm understanding of these ideas, students cannot grasp more complex models. Yet most textbooks devote no more than a few pages, or even a few paragraphs, to the topic of exponential population growth.

THE ORGANIZATION OF THIS BOOK

This primer grew out of my dissatisfaction with existing textbooks and the fact that I could not relegate mathematical details of the models to “course readings.” In this book, I have tried to present a concise but detailed exposition of the most common mathematical models in population and community ecology. Each chapter follows the same structured format:

Model Presentation and Predictions derives the models from first principles so students can see where the equations come from. Essential equations are

highlighted, but a number of intermediate algebraic “expressions” are also presented so students can understand how we get from point A to point B. With the equations in hand, the predictions from the model are explained. I have relied heavily on graphical approaches, because they are often more enlightening than algebraic solutions of the equations. Although most of the models in this book are continuous differential equations, students do not need to integrate or differentiate equations to follow this material. Instead, I have emphasized the biological interpretation of the variables in the models and how the predictions change when the variables are altered. The material in this section of each chapter is covered in some form in nearly every introductory ecology course.

Model Assumptions lists the mathematical and biological assumptions behind the equations. This material is usually covered in most textbooks, but is often scattered or buried in the text.

Model Variations explains related models that can usually be derived by relaxing one or more of the critical assumptions. In this section, I have introduced topics that are suitable for advanced and graduate-level courses, including models of environmental and demographic stochasticity, stage-structured population growth, nonlinear predator–prey isoclines, intraguild predation, and passive sampling.

Empirical Examples includes two or three field studies that illustrate the utility of the models. The examples are restricted to field studies that actually measure parameters that are relevant to the models, although in many cases I was hard-pressed to find good examples. Often, the studies in which the models fail to predict patterns in nature are more enlightening than the apparent successes.

Problems give students the chance to work with the equations and understand their behavior by plugging in some numbers. The exercises are highly simplified “story problems,” but they teach students how to apply the model concepts to empirical data and give them a better intuitive understanding of the equations. Fully explained solutions follow each problem set. Advanced problems that correspond to the material in “Model Variations” are marked with an asterisk.

Symbols and variable names are often a source of confusion for students. I have tried to use the symbols that are encountered in most ecology textbooks, but have made some changes for clarity and consistency. The primer includes very few literature citations, which are intended only to provide sources for

the equations and examples. There is no glossary, but new terms are introduced in the text in **boldface type** to alert students to novel concepts.

THE CONTENT OF THIS BOOK

Chapters 1–4 cover models for single species, and Chapters 5–7 cover models for two or more species. In Chapter 1, the model of exponential growth is developed carefully from first principles. Advanced topics include environmental and demographic stochasticity. In Chapter 2, the logistic growth model is developed as an extension of the exponential model by incorporating density dependence in birth and death rates. Discrete growth with chaos, and random and periodic variation in carrying capacity are also described. Chapter 3 covers exponential growth for age-structured populations. Advanced topics include the derivation of the Euler equation, reproductive value, and stage-structured matrix models.

Chapter 4 reflects my own interests in metapopulation models. These models relax the unrealistic assumption of no migration of individuals and represent the simplest equations for open populations. There is a close analogy between the births and deaths of individuals in a local population and the colonization and extinction of populations in a metapopulation. There is also an important conceptual link between single-species metapopulation models and the MacArthur–Wilson model of island biogeography, which is developed in Chapter 7. Although metapopulation models are only just beginning to appear in textbooks, they are an important tool for studying population dynamics in a fragmented landscape, and may have applications in conservation biology.

Chapters 5 and 6 present the standard two-species competition and predation models, and include some more complex variations with nonlinear isoclines. Chapter 5 develops a model of intraguild predation, in which species function simultaneously as predators and competitors. Chapter 6 includes a discussion of host–parasite models and briefly addresses the problem of population cycles. Both chapters stress the use of the state-space diagram as an important graphical tool for ecological modeling. Chapter 7 presents the MacArthur–Wilson equilibrium model as one possible explanation for the species–area relationship. Habitat diversity and the passive sampling model are also offered as alternative hypotheses.

PRECEDENTS FOR THIS BOOK

This book was inspired by two earlier ecology texts. The first was *A Primer of Population Biology* by E. O. Wilson and W. H. Bossert. This remarkable book, first published in 1971, has been used by thousands of students. Its concise prose, modest size, and quantitative problems introduced a generation of students to mathematical approaches in ecology and population genetics. The

second was *Theoretical Ecology*, edited by R. M. May. May's overview chapters provided a concise framework for Chapters 1, 2, 5, and 6 of this primer, which cover much of the same material in a greatly expanded form.

At the risk of overstating the obvious, this primer is not a substitute for a full-length ecology text. Because of its brevity, it completely ignores many important topics in ecology that are not amenable to treatment with simple mathematical models. I hope that its concise format and modest price will justify its use as a supplementary text. If this primer helps students to understand the development, application, and limitations of mathematical models in ecology, then I will have been successful.

SOME THOUGHTS FOR THE INSTRUCTOR

I designed this primer with two sorts of courses in mind. First, the primer can serve as a supplementary text for large, introductory undergraduate courses. The material covered in "Model Presentation and Predictions" and "Model Assumptions" assumes that students have had only a single semester of calculus, and have probably forgotten most of what they learned. In my large introductory course at the University of Vermont (> 100 students), I teach all the basic material in Chapters 1, 2, 3, 5, 6, and 7. Although I do not teach the equations from Chapter 4, I do cover basic principles of metapopulations and some empirical examples. The unstarred problem sets in all the chapters are appropriate for an introductory course.

I also use the primer in my community ecology course (< 25 students), which is taught to advanced undergraduates and beginning graduate students. In this course, I treat the introductory material as a concise review, and spend more time developing the material in "Model Variations." This advanced material assumes a minimal grasp of calculus, and an exposure to basic statistical concepts of probability, means, and variances. A knowledge of matrix algebra is helpful, but not essential, for the advanced material in Chapter 3. Both the unstarred and starred problems are appropriate at this level.

My hope is that the primer will be useful to two types of instructors. Those who prefer a quantitative approach, as I do, may use the primer as a template for lectures that build ecological models from first principles. Problem-solving is essential for such a course, and most of the problems at the end of each chapter work well as exam questions.

Other instructors may not wish to devote so much lecture time to models. For these courses, the primer may serve as a tutorial to allow students to learn the details of the models on their own. In this case, instructors might wish to place more emphasis on the model assumptions and empirical examples, and perhaps eliminate the problem sets entirely.

Ecology textbooks continue to increase in size and cost, making it difficult

to justify a supplemental text. However, as good as the standard textbooks are, none of them treats the mathematical models with the care and detail they deserve. I hope *A Primer of Ecology* makes your teaching easier and helps your students to better understand ecological models. For me, this has always been the most challenging and rewarding part of teaching ecology.

February 28th, 1994
 5° 33' 20" N, 87° 02' 35" W
 Cocos Island, Costa Rica

Acknowledgments

First and foremost, I thank my students. Undergraduate and graduate students at the Universities of Oklahoma and Vermont forced me to slow down, speak plainly, and avoid jargon in my lectures. Their questions taught me what needed to be included in a primer of ecology. Andy Sinauer and his staff enthusiastically supported this project, transforming my text and rough drawings into a polished product. I thank Bobbie Lewis for copyediting the manuscript, and Shahid Naceem for his artwork, which graces the frontispiece and chapter headings. Janice Moore and Bob May encouraged me to accept the challenge of writing a primer, and Joe Schall taught me to keep the perspective of the average student while I wrote.

Each chapter benefitted from detailed reviews by a number of colleagues: Bob May, Janice Moore, Bob Ricklefs, and Joe Schall (1), Peter Stiling and Guiyun Yan (2), Hal Caswell (3), Bob Holt (4), Rob Colwell (5), Andy Dobson (6), and Mark Lomolino (7). Stewart Berlocher and Carol Boggs read the entire manuscript, and provided feedback from their undergraduates as well. Bob Ricklefs' critical review of Chapter 1 forced me to sharpen my prose throughout. The treatment of demography in many textbooks is confused and incorrect, and my initial draft of Chapter 3 was no exception. I thank Hal Caswell for reviewing this chapter twice and for setting me straight on a number of major and minor points.

Chapter 5 owes a special debt to Rob Colwell. The organization of this chapter, the restatement of the competitive exclusion principle, and the "milkshake analogy" were all taken from my undergraduate lecture notes from Colwell's community ecology course at the University of California, Berkeley (winter 1980). Andy Dobson encouraged me to include more quantitative material on predator-prey isoclines and provided me with a derivation of the disk equation from his own course lecture notes.

Chris Taylor double-checked all the problem sets and detected several typos and errors that would have frustrated students. Neil Buckley corrected my prose throughout and patiently helped me to read galley proofs. The National Science Foundation and the Fulbright Foundation supported my research while I wrote, and my graduate students cheerfully tolerated my preoccupation. Finally, I thank my wife and best friend, Maryanne Kampmann. She forced me to put down the guitar and get back to work, so that this primer was actually finished on schedule.

To the Student

The most common question beginning ecology students ask me is, "Why do we have to use so much mathematics to study ecology?" Many students enroll in my ecology course expecting to hear about whales, global warming, and the destruction of tropical rain forests. Instead they are confronted with exponential growth, doubling times, and per capita rates of increase. The two lists of topics are not unrelated. But before we can begin to solve complex environmental problems, we have to understand the basics. Just as a mechanical engineer must learn the principles of physics to build a dam, a conservation biologist must learn the principles of ecology to save a species.

The science of ecology is the study of distribution and abundance. In other words, we are interested in predicting where organisms occur (distribution), and the sizes of their populations (abundance). Ecological studies rely on measurements of distribution and abundance in nature, so we need the tools of mathematics and statistics to summarize and interpret these measurements.

But why do we need the mathematical models? One answer is that we need models because nature is so complex. We could spend a lifetime measuring different components of distribution and abundance and still not have a very clear understanding of ecology. The mathematical models act as simplified road maps, giving us some direction and idea of exactly what things we should be trying to measure in nature.

The models also generate testable predictions. By trying to verify or refute these predictions, we will make much faster progress in understanding nature than if we try to go out and measure everything without a plan. The models highlight the distinction between the *patterns* we see in nature and the different *mechanisms* that might cause those patterns.

There are two dangers inherent in the use of mathematical models in ecology. The first danger is that we build models that are too complex. When this happens, the models may contain many variables that we can never measure in nature, and the mathematical solutions may be too complex. Consequently, the most useful ecological models are often the simplest ones, and these have been emphasized throughout this primer.

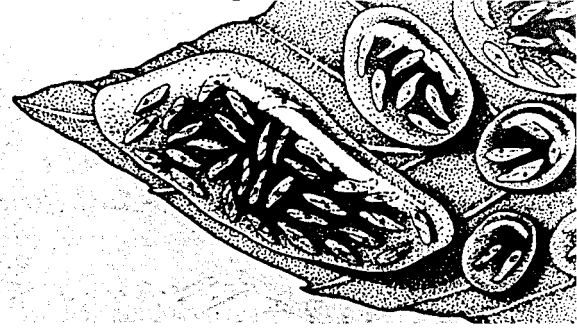
The second danger is that we forget that the models are abstract representations of nature. However logical a model might appear, nothing says that nature must follow its rules. By carefully focusing on the assumptions of the

model, we may be able to pinpoint the places where it departs from reality. As you will see from the examples in this primer, the models often tell us more about nature when their predictions do not match our field observations.

The purpose of this primer is to de-mystify the mathematical models used in ecology. Many of the equations in this primer can also be found in your textbook. However, your textbook may provide little or no explanation for where these equations come from, whereas the primer develops them step by step. I hope this primer will help you to understand the mathematical models and to appreciate their strengths and limitations.

CHAPTER 1

Exponential Population Growth



Model Presentation and Predictions

ELEMENTS OF POPULATION GROWTH

A **population** is a group of plants, animals, or other organisms, all of the same species, that live together and reproduce. Just as an individual grows by gaining weight, a population grows by gaining individuals. What controls population growth? In this chapter, we will build a simple mathematical model that predicts population size. In later chapters, we will flesh out this model by including resource limitation (Chapter 2), age structure (Chapter 3), and migration (Chapter 4). We will also introduce other players: populations of competitors (Chapter 5) and predators (Chapter 6) that can control growth. But for now, we will concentrate on a single population and its growth in a simple environment.

The variable N will be used to indicate the size of the population. Because population size changes with time, we will use the subscript t to indicate the point in time we are talking about. Thus, N_t is the number of individuals in the population at time t . By convention, we use $t = 0$ to indicate the starting point. For example, suppose we census a population of tarantulas and count 500 spiders at the beginning of our study. We revisit the population in one year and count 800 spiders. Thus, $N_0 = 500$ and $N_1 = 800$.

The units of t , in contrast to their numerical values, depend on the organism we are studying. For rapidly growing populations of bacteria or protozoa, t might conveniently be measured in minutes. For long-lived sea turtles or bristlecone pines, t would be measured in years or decades. Whatever units we use, we are interested in predicting the future population size (N_{t+1}) based on its current size (N_t).

The biological details of population growth vary tremendously among different species, and even among different populations within the same species. The factors that cause a tarantula population to increase from 500 to 800 spiders will be very different from the factors that cause an endangered condor population to decrease from 10 to 8 birds. Fortunately, all changes in population size can be classified into just four categories. Populations increase because of births and decrease because of deaths. Population size also changes if individuals move between sites. Populations increase when new individuals arrive (immigration) and decrease when resident individuals depart (emigration).

These four factors operate at different spatial scales. Births and deaths depend on current population size, as we will explain in a moment. To understand births and deaths, we need to study only the target population. By contrast, emigration and immigration depend on the movement of individuals. If

we want to describe these processes, we must keep track of not just one, but several interconnected populations.

Any combination of the four factors will change population size. For our tarantula example, the initial population of 500 spiders might have produced 400 new spiderlings during the year and lost 100 adult spiders to death, with no movement of individuals. Alternatively, there might have been 50 births and 50 deaths, with 300 residents leaving (emigration) and 600 spiders arriving from other populations (immigration). Either scenario leads to an increase of 300 spiders.

These four factors can be incorporated into a mathematical expression for population growth. In this expression, B represents the number of births, D is the number of deaths, I is the number of new immigrants entering the population, and E is the number of emigrants leaving the population between time t and $t + 1$:

$$N_{t+1} = N_t + B - D + I - E \quad \text{Expression 1.1}$$

Expression 1.1 says that population size in the next time period (N_{t+1}) equals the current population size (N_t) plus births (B) and immigrants (I), minus deaths (D) and emigrants (E). We are interested in the change in population size (ΔN), which is simply the difference in population size between last time and this time. We get this by subtracting N_t from both sides of Expression 1.1:

$$N_{t+1} - N_t = N_t - N_t + B - D + I - E \quad \text{Expression 1.2}$$

$$\Delta N = B - D + I - E \quad \text{Expression 1.3}$$

To simplify things, we will assume that our population is **closed**; in other words, there is no movement of individuals between population sites. This assumption is often not true in nature, but it is mathematically convenient and it allows us to focus on the details of local population growth. In Chapter 4, we will examine some models that allow for movement of individuals between patches. If the population is closed, both I and E equal zero, and we do not need to consider them further:

$$\Delta N = B - D \quad \text{Expression 1.4}$$

We will also assume that the population is growing smoothly, so that births and deaths occur continuously. For our tarantula example, recall that the population increased by 300 spiders in one year. If population growth were continuous, we would find that the population had increased by exactly 150 spiders in exactly 6 months. And if we censused the population after 2 days, we should find exactly $(2/365) \cdot (300) = 1.64384$ additional spiders!

Perfectly continuous growth is not possible because individuals do not

reproduce themselves in fractional values. However, population growth of long-lived organisms may be approximately continuous. Mathematically, this allows us to describe population growth with a **continuous differential equation**. Thus, population growth is measured as the change in population size (dN) during a very small interval of time (dt):

$$\frac{dN}{dt} = B - D \quad \text{Expression 1.5}$$

Now we will focus on B and D . Because this is a continuous differential equation, B and D now represent the number of births and deaths during a very short time interval. What factors control births and deaths? The number of births will certainly depend on population size. For example, a population of 1000 warblers will produce many more eggs than a population of only 25 birds. If each individual produces the same number of offspring during a short time interval, the number of births in the population will be directly proportional to population size. Let b (lowercase!) denote the **instantaneous birth rate**. The units of b are number of births per individual per unit time [births/(individual \cdot time)]. Over a short time interval, the number of births in the population is the product of the instantaneous birth rate and the population size:

$$B = bN \quad \text{Expression 1.6}$$

Similarly, we can define an **instantaneous death rate** d , with units being number of deaths per individual per unit time [deaths/(individual \cdot time)]. Of course, an individual either dies or it doesn't, but this rate is measured for a continuously growing population over a short time interval. Again, the product of the instantaneous death rate and the population size gives the number of deaths during the interval:

$$D = dN \quad \text{Expression 1.7}$$

These simple functions will not always apply in the real world. In some cases, the number of births may not depend on the current population size. For example, in some plant populations, seeds remain dormant in the soil for many years in a "seed bank." Consequently, the number of emergent seedlings (births) may reflect the structure of the plant population many years ago. A model for such a population would include a **time lag** because the current growth rate actually depends on population size at a much earlier time.

Expressions 1.6 and 1.7 also imply that b and d are constant. No matter how large the population gets, individuals have the same birth and death rates! But in the real world, birth and death rates may be affected by crowding: the larger the population, the lower the birth rate and the higher the

death rate. We will explore this sort of **density-dependent model** in Chapter 2. For now, we will develop our model assuming a constant birth rate, b , and a constant death rate, d . Substituting Expressions 1.6 and 1.7 into Expression 1.5 and rearranging the terms gives us:

$$\frac{dN}{dt} = (b - d)N \quad \text{Expression 1.8}$$

Let $b - d$ equal the constant r , the **instantaneous rate of increase**. Sometimes r is called the **intrinsic rate of increase**, or the **Malthusian parameter** after the Reverend Thomas Robert Malthus (1766–1834). In his famous "Essay on the Principle of Population" (1798), Malthus argued that food supply could never keep pace with human population growth, and that human suffering and misery were an inevitable consequence.

The value of r determines whether a population increases exponentially ($r > 0$), remains constant in size ($r = 0$), or declines to extinction ($r < 0$). The units of r are individuals per individual per unit time [individuals/(individual \cdot time)]. Thus, r measures the **per capita**, or per individual, rate of population increase over a short time interval. That rate is simply the difference between b and d , the instantaneous birth and death rates. Because r is an instantaneous rate, we can change its units by simple division. For example, because there are 24 hours in a day, an r of 24 individuals/(individual \cdot day) is equivalent to an r of 1 individual/(individual \cdot hour). Substituting r back into Expression 1.8, we arrive at our first model of population growth:

$$\frac{dN}{dt} = rN \quad \text{Equation 1.1}$$

Equation 1.1 is a simple model of exponential population growth. It says that the population growth rate (dN/dt) is proportional to r and that populations only increase when the instantaneous birth rate (b) exceeds the instantaneous death rate (d), so that $r > 0$. If r is positive, population growth continues unchecked and is proportional to N : the larger the population, the faster its rate of increase.

When will our model population not grow? A population will neither increase nor decrease when the population growth rate equals zero ($dN/dt = 0$). For Equation 1.1, there are only two cases when this is true. The first is when $N = 0$. Because of migration, population growth in nature will not necessarily stop when the population reaches zero. But in our simple model immigration is not allowed, so the population will stop growing if it ever hits the "floor" of zero individuals. The population will also stop growing if r should equal zero. In other words, if the birth and death rates are exactly bal-

anced, the population will neither increase nor decrease in size. In all other cases, the population will either increase exponentially ($r > 0$) or decline to extinction ($r < 0$).

PROJECTING POPULATION SIZE

Equation 1.1 is written as a differential equation. It tells us the population growth rate, but not the population size. However, if Equation 1.1 is integrated (following the rules of calculus), the result can be used to project, or predict, population size:

$$N_t = N_0 e^{rt} \quad \text{Equation 1.2}$$

N_0 is the initial population size, N_t is the population size at time t , and e is a constant, the base of the natural logarithm ($e \approx 2.717$). Knowing the starting population size and the intrinsic rate of increase, we can use Equation 1.2 to forecast population size at some later time. Equation 1.2 is similar to the formula used by banks to calculate compound interest on a savings account.

Figure 1.1a illustrates some population trajectories that were calculated from Equation 1.2 for five different values of r . In Figure 1.1b, these same data are shown on a semilogarithmic plot, in which the y axis is the natural logarithm (base e) of population size. This transformation converts an exponential growth curve to a straight line. The slope of this line is r .

These graphs show that when $r > 0$, populations increase exponentially, and that the larger the value of r , the faster the rate of increase. When $r < 0$, populations decline exponentially. Mathematically, such populations never truly reach zero, but when the population reaches a projected size of less than one individual, extinction has occurred (by definition).

CALCULATING DOUBLING TIME

One important feature of a population (or a savings account) that is growing exponentially is a constant **doubling time**. In other words, no matter how large or small the population, it will always double in size after a fixed time period. We can derive an equation for this doubling time, t_{double} , by noting that, if the population has doubled in size, it is twice as large as the initial population size:

$$N_{t_{\text{double}}} = 2N_0 \quad \text{Expression 1.9}$$

Substituting back into Equation 1.2 gives an expression in terms of initial population size:

$$2N_0 = N_0 e^{rt_{\text{double}}} \quad \text{Expression 1.10}$$

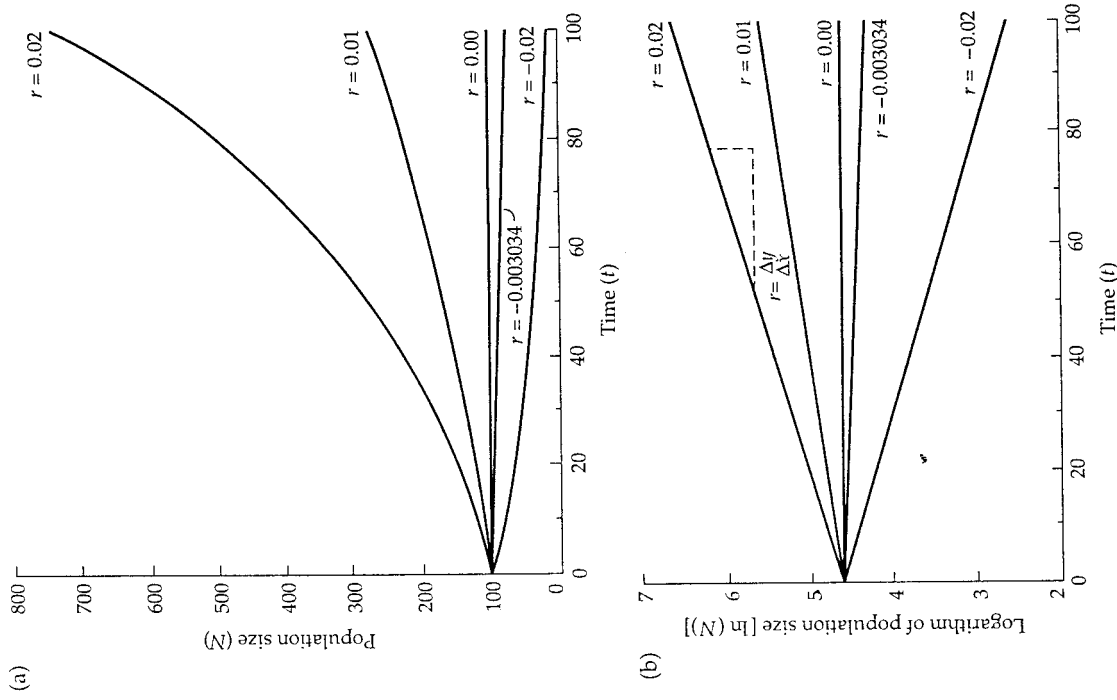


Figure 1.1 (a) Trajectories of exponential population growth, calculated from a starting population size of 100 individuals. The estimated r of -0.003034 [individuals / (individual \cdot year)] corresponds to the projection for the grizzly bear (*Ursus arctos horribilis*) population of Yellowstone National Park (see Figure 1.6). (b) Exponential growth curves plotted on a semilogarithmic graph. The same data are used as in (a), but the y axis (population size) shows the natural logarithm (base e) of population size. In this type of graph, an exponential curve becomes a straight line; the slope of that line is r , the intrinsic rate of increase.

Table 1.1 Estimates of r and doubling times for different organisms.

Species	Common name	r [individuals / (individual • day)]	Doubling time
T phage	Virus	300.0	3.3 minutes
<i>Escherichia coli</i>	Bacterium	58.7	17 minutes
<i>Paramecium caudatum</i>	Protozoan	1.59	10.5 hours
<i>Hydra</i>	Hydra	0.34	2 days
<i>Tribolium castaneum</i>	Flour beetle	0.101	6.9 days
<i>Rattus norvegicus</i>	Brown rat	0.0148	46.8 days
<i>Bos taurus</i>	Domestic cow	0.001	1.9 years
<i>Avicennia marina</i>	Mangrove	0.00055	3.5 years
<i>Nothofagus fusca</i>	Southern beech	0.000075	25.3 years

From Fenchel (1974).

Dividing through by N_0 eliminates it from both sides of the equation:

$$2 = e^{rt_{\text{double}}} \quad \text{Expression 1.11}$$

Taking the natural logarithm of both sides gives:

$$\ln(2) = rt_{\text{double}} \quad \text{Expression 1.12}$$

Expression 1.12 can be rearranged to solve for doubling time:

$$t_{\text{double}} = \frac{\ln(2)}{r} \quad \text{Equation 1.3}$$

Thus the larger r is, the shorter the doubling time. Table 1.1 gives some estimated values of r (with their corresponding doubling times) for different species of plants and animals. Among species, r varies considerably, and much of this variation is related to body size: small-bodied organisms grow faster and have larger rates of population increase than large-bodied organisms. For example, bacteria and protozoa can reproduce by asexual fission every few minutes and have high population growth rates. Larger organisms, such as primates, have delayed reproduction and long generation times, which lead to low values of r . Corresponding doubling times range from minutes for viruses to decades for beech trees.

Note, however, that even “slow-growing” populations eventually will reach astronomical sizes if they increase exponentially. Table 1.2 projects the future population size for a hypothetical herd of 50 Vermont cows [$r = 0.365$

Table 1.2 Exponential growth of a herd of 50 cattle, with $r = 0.365$ cows/(cow • year).

Year	Herd size
0	50.0
1	72.0
2	103.8
3	149.5
4	215.3
5	310.1
10	1923.7
50	4.2×10^9
100	3.6×10^{17}
150	3.0×10^{25}
200	2.5×10^{33}

Population sizes calculated from Equation 1.2.

cows/(cow • year)]. After 150 years of exponential growth, the model predicts a herd of 3×10^{25} cattle, the weight of which would exceed that of the planet earth!

Model Assumptions

What are the assumptions of Equation 1.1? In other words, what is the underlying biology of a population that is growing exponentially? This is a critical question that must be asked for any mathematical model we construct. The predictions of a mathematical model depend on its underlying assumptions. If certain assumptions are violated, or changed, the predictions of the model will also change. Other assumptions may be less critical to the predictions of the model; the model is **robust** to violations of these assumptions. We make the following assumptions for a population growing according to Equation 1.1:

- ✓ **No I or E.** The population is “closed.” changes in population size depend only on local births and deaths. We made this simplifying assumption in Expression 1.4, so that we could model the growth of a single population without having to keep track of organisms moving between populations. In Chapter 4, we will relax this assumption and build some simple models in which there is migration between populations.

✓ **Constant b and d .** If a population is going to grow with constant birth and death rates, an unlimited supply of space, food and other resources must be available. Otherwise, the birth rate will decrease and/or the death rate will increase as resources are depleted. Constant birth and death rates also imply that b and d do not change randomly through time. Later in this chapter, we will incorporate variable birth and death rates in the model to see how the predictions are affected.

✓ **No genetic structure.** Equation 1.1 implies that all the individuals in the population have the same birth and death rates, so there cannot be any underlying genetic variation in the population for these traits. If there is genetic variation, the genetic structure of the population must be constant through time. In this case, r represents an *average* of the instantaneous rate of increase for the different genotypes in the population.

✓ **No age or size structure.** Similarly, there are no differences in b and d among individuals due to their age or body size. Thus, we are modeling a sexless, parthenogenetic population in which individuals are immediately reproductive when they are born. A growing population of bacteria or protozoa most closely approximates this situation. In Chapter 3, we will relax this assumption and examine a model of exponential growth in which individuals have different birth and death rates as they age. If there are differences among ages, the population must have a stable age structure (see Chapter 3); in this case, r is an average calculated across the different age classes.

✓ **Continuous growth with no time lags.** Because our model is written as a simple differential equation, it assumes that individuals are being born and dying continuously, and that the rate of increase changes instantly as a function of current population size. Later in this chapter, we will relax the assumption of continuous growth and examine a model with discrete generations. In Chapter 2, we will explore models with time lags, in which population growth depends not on current population size, but on its size at some time in the past.

The most important assumption on this list is that of constant b and d , which implies unlimited resources for population growth. Only if resources are unlimited will a population continue to increase at an accelerating rate. If the other assumptions are violated, populations may still increase exponentially, although migration and time lags will complicate the picture.

But unlimited resources do not occur in nature, and we know that no real population increases without bound. So, why does the exponential growth model form the cornerstone of population biology? Although no population

can increase forever without limit, all populations have the *potential* for exponential increase. Indeed, this potential for exponential increase in population size is one of the key factors that can be used to distinguish living from non-living objects. The exponential model recognizes the multiplicative nature of population growth and the positive feedback that gives populations the potential to increase at an accelerating rate.

Exponential population growth is also a key feature of Charles Darwin's (1809–1882) theory of natural selection. Darwin read Malthus' writings and recognized that the surplus of offspring resulting from exponential growth would allow natural selection to operate and bring about evolutionary change. Finally, although no population can increase forever, resources may be *temporarily* unlimited so that populations go through phases of exponential increase. Outbreaks of insect pests, invasions of "weedy" plant species, and the plight of overcrowded human populations are compelling evidence of the power of exponential population growth.

Model Variations

CONTINUOUS VERSUS DISCRETE POPULATION GROWTH

We will now explore some variations on our exponential growth model. First, consider the fact that birth and death for many organisms are not continuous processes. Many insects and annual desert plants, for example, reproduce only once, then die; the offspring that survive form the basis for next year's population. If birth and death rates are constant (as in the exponential model), then the population will increase or decrease by the same factor each year. This population has **discrete generations** and is modeled with a **discrete difference equation** rather than a continuous differential equation:

$$N_{t+1} = \lambda N_t \quad \text{Expression 1.13}$$

Here λ is the **finite rate of increase**. λ is always a positive number that measures the proportional change in population size from one year to the next. Thus, λ is the ratio of the population size during the next time period to the population size for the current time period (N_{t+1}/N_t). After two years, the population size will be:

$$N_2 = \lambda N_1 = \lambda(\lambda N_0) = \lambda^2 N_0 \quad \text{Expression 1.14}$$

Or, after t years of growth:

$$N_t = \lambda^t N_0 \quad \text{Equation 1.4}$$

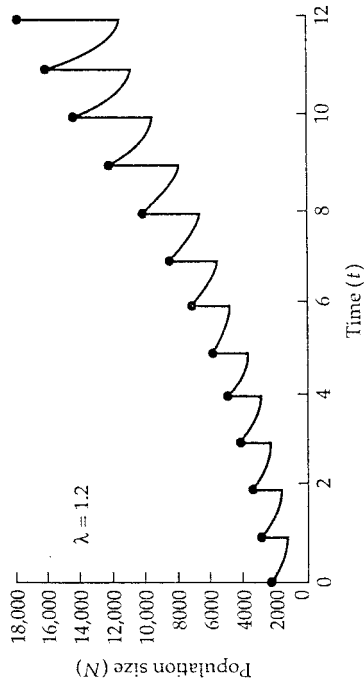


Figure 1.2 Discrete population growth. In this model, births are pulsed at the beginning of the year, and deaths occur continuously.

Equation 1.4 is analogous to Equation 1.2, which we used to project population size in the continuous model. What does population growth look like with the discrete model? The answer depends on the precise timing of birth and death events. Imagine that births are pulsed each spring and that deaths occur continuously throughout the year. The population growth curve will resemble a jagged saw blade, with a sharp vertical increase from births each spring, followed by a gradual decrease from deaths during the rest of the year. In spite of this decrease, the overall curve will rise exponentially, because annual births exceed annual deaths (Figure 1.2). The size of each “tooth” in the growth curve will increase year after year because the same fractional increase will add proportionally more individuals to a large population. For example, if $\lambda = 1.2$, the population increases by 20% each year. If the population size is 100, it will increase by 20 in one year. But when the population size is 1000, it will increase by 200 in one year.

Suppose our population reproduced twice a year, as is the case for some insects. Now we would have a “tooth” on the graph every six months. If the time step between reproductive periods becomes shorter and shorter, the teeth on the graph will be closer and closer together. Finally, if the time step is infinitely small, the curve is no longer jagged but is smooth, and we have arrived again at the continuous model of exponential growth (Equation 1.2). The continuous model essentially “connects the dots” of time in the discrete model. Because the continuous model is equivalent to a discrete difference equation with an infinitely small time step, λ is mathematically related to r by:

$$r = \ln(\lambda) \quad \text{Equation 1.5}$$

where \ln is the natural logarithm (base e). We can express Equation 1.5 in equivalent exponential form as:

$$\lambda = e^r \quad \text{Equation 1.6}$$

This relationship between r and λ also establishes the following numerical equivalents:

$$r > 0 \leftrightarrow \lambda > 1 \quad \text{Expression 1.15}$$

$$r = 0 \leftrightarrow \lambda = 1 \quad \text{Expression 1.16}$$

$$r < 0 \leftrightarrow 0 < \lambda < 1 \quad \text{Expression 1.17}$$

Because λ is a ratio of population sizes, it is a dimensionless number with no units. However, λ is associated with the particular time step of the equation and cannot be changed by a simple scaling. For example, a λ of 1.2 measured with a yearly time step is *not* equivalent to a λ of 0.1 measured with a monthly time step. A λ of 1.2 yields a 20% annual increase, whereas a λ of 0.1 yields a 90% monthly decrease! If you need to change the time step for λ , first convert λ to r using Equation 1.5. Then scale r to the appropriate time units and convert back to λ with Equation 1.6. In this example, $\lambda = 1.2$ is equivalent to $r = 0.18232$ individuals/(individual \cdot year). Dividing by 12 (months) gives $r = 0.01519$ individuals/(individual \cdot month). From Equation 1.6, $\lambda = 1.0153$, with a monthly time step. As a check on this calculation, we can use Equation 1.4 to show that, after 12 months:

$$N_t = (1.0153)^{12} N_0 \quad \text{Expression 1.18}$$

$$N_t = 1.2 N_0 \quad \text{Expression 1.19}$$

This shows that $\lambda = 1.0153$ for a monthly time step is equivalent to $\lambda = 1.2$ for a yearly time step.

In summary, the predictions of the discrete and continuous models of exponential population growth are qualitatively similar to one another. In Chapter 2, we will see that discrete models behave very differently when we incorporate resource limitation.

ENVIRONMENTAL STOCHASTICITY

Equation 1.2 is entirely deterministic. If we know N_0 , r , and t , we can calculate the predicted population size to the last decimal place. If we started over with the same set of conditions, the population would grow to precisely the same size. In such a **deterministic model**, the outcome is determined solely

by the inputs, and nothing is left to chance.

Deterministic models represent an idealized view of a simple, orderly world. But the real world tends to be complex and uncertain. Think of public transportation. Does any commuter ever expect their bus or train to arrive at *precisely* the time indicated in the printed schedule? At least in American cities, buses are delayed, trains break down, and subways travel at irregular speeds, all of which introduce uncertainty (and anxiety) into the daily commute.

Could we incorporate all of the complex sources of variation into a public transportation model? Not very easily. But we could measure, each day, the arrival time of our bus. After many commuting days, we could calculate two numbers that would help us to estimate the uncertainty. The first number is the *average* arrival time of the bus. If we use the variable x to indicate the time the bus arrives, the average is depicted as \bar{x} . Approximately half of all buses will arrive later than \bar{x} and half will arrive earlier. The second number we could calculate is the *variance* in arrival times (σ_x^2). The variance measures the variability or uncertainty associated with the average. If the variance is small, then we know that most days the bus will arrive within, say, two minutes of the average. But if the variance is large, the arrival time of the bus on any given morning could be as much as 20 minutes earlier or 20 minutes later than \bar{x} . Obviously, our "commuting strategy" will be affected by both the mean and the variance of x .

How can we incorporate this type of uncertainty into an exponential population growth model? Suppose that the instantaneous rate of increase is no longer a simple constant, but instead changes unpredictably with time. Uncertainty in r means there are good times and bad times for population growth. During good times, the birth rate is much larger than the death rate, and the population can increase rapidly. During bad times, the difference between birth and death rates is much smaller, or perhaps even negative, so that the population increases slowly, or even decreases, for a short time period. Without specifying all of the biological causes of good and bad years, we can still develop a simple model of population growth in a stochastic environment. Variability associated with good and bad years for population growth is known as **environmental stochasticity**.

Imagine that a population is growing exponentially with an average r (\bar{r}) and a variance in r (σ_r^2). We will use this model to predict the average population size at time t (\bar{N}_t) and the variance in population size ($\sigma_{N_t}^2$). Make sure you understand the difference between these two averages and the two variances: the average and variance in r are used to predict the average and variance in N_t .

The derivation of this model is beyond the scope of this primer, but the results are straightforward. First, the average population size for a population growing with environmental stochasticity is:

$$\bar{N}_t = N_0 e^{rt} \quad \text{Equation 1.7}$$

This is no different from the deterministic model (Equation 1.2) except that we use the average r to predict the average N_t . However, like the "average family" with 2.1 children, \bar{N}_t may not be a very accurate descriptor of any particular population. Figure 1.3 shows a computer simulation of a population growing with environmental stochasticity. Although the population achieves exponential increase in the long run, it fluctuates considerably from one time period to the next. The variance in population size at time t is given by (May 1974a):

$$\sigma_{N_t}^2 = N_0^2 e^{2rt} (e^{\sigma_r^2 t} - 1) \quad \text{Equation 1.8}$$

Other mathematical expressions for this variance are possible, depending on precisely how the model is formulated. Equation 1.8 tells us several things about the variance of the population. First, population variance increases with time. Like stock-market projections or weather forecasts, the further into the future we try to predict population size, the more uncertain our estimate. Consequently, the population growth curve resembles a funnel that flares out with increasing time (Figure 1.3). Second, the variance of N_t is proportional to both the mean and variance of r . Populations that are growing

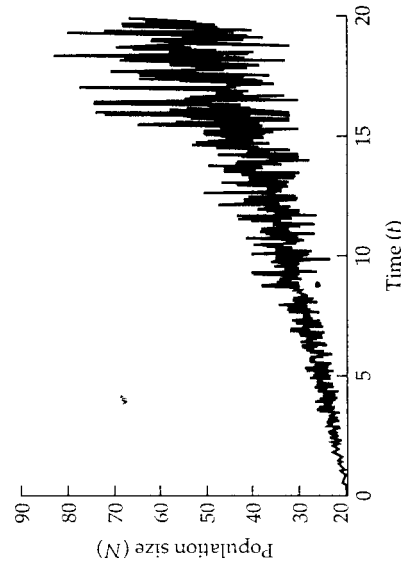


Figure 1.3 Exponential growth with environmental stochasticity. In this model, the instantaneous rate of increase fluctuates randomly through time. Here $N_0 = 20$; $r = 0.05$; $\sigma_r^2 = 0.0001$.

rapidly, or have a variable r , fluctuate more than slow-growing populations or those with a relatively constant r . Finally, if the variance of r is zero, Equation 1.8 collapses to zero—there is no variance in N_t , so we have returned to the deterministic model.

There is a limit to how much the population can vary in size and still persist. If N fluctuates too violently, the population may “crash” to zero. This can happen even if \bar{r} is large enough to ensure rapid growth for the “average” population. Extinction from environmental stochasticity will almost certainly happen if the variance in r is greater than twice the average of r (May 1974a):

$$\sigma_r^2 > 2\bar{r} \quad \text{Equation 1.9}$$

In our deterministic model, the population increased exponentially as long as r was greater than zero. With environmental stochasticity, the average population size also increases exponentially as a function of \bar{r} . However, if the variance in r is too large, there is a measurable risk of population extinction.

DEMOGRAPHIC STOCHASTICITY

Environmental stochasticity is not the only source of variability that can affect populations. Even if r is constant, populations may still fluctuate because of **demographic stochasticity**. Demographic stochasticity arises, in part, because most organisms reproduce themselves as discrete units: an ostrich can lay 2 eggs or 3, but not 2.6! Some clonal plants and corals can reproduce by fragmentation and asexual budding, and in that sense, “pieces” of individuals may contribute to population increase (see Chapter 3). But for most organisms, population growth is an integer process.

If we were to follow a population over a short period of time, we would see that births and deaths are not perfectly continuous, but instead occur sequentially. Suppose that the birth rate is twice as large as the death rate. This means that a birth would be twice as likely to occur in the sequence as a death. In a perfectly deterministic world, the sequence of births and deaths would look like this: ...BBDBBDBBDD... But with demographic stochasticity, we might see: ...BBBBDBBBBB... By chance, we may hit a run of four births in a row before seeing a death in the population. This demographic stochasticity is analogous to genetic drift, in which allele frequencies change randomly in small populations. In a model of demographic stochasticity, the probability of a birth or a death depends on the relative magnitudes of b and d :

$$P(\text{birth}) = \frac{b}{(b+d)} \quad \text{Equation 1.10}$$

$$P(\text{death}) = \frac{d}{(b+d)} \quad \text{Equation 1.11}$$

Suppose that, for a chimpanzee population, $b = 0.55$ births/(individual • year) and that $d = 0.50$ deaths/(individual • year). This yields an r of 0.05 individuals/(individual • year), with a corresponding doubling time of 13.86 years (Equation 1.3). From Equations 1.10 and 1.11, the probability of birth is $[0.55/(0.55 + 0.50)] = 0.524$, and the probability of death is $[0.50/(0.55 + 0.50)] = 0.476$. Note that these probabilities must add to 1.0, because the only “events” that can occur in this population are births or deaths. Because a birth is more likely than a death, the chimpanzee population will, on average, increase. However, population size can no longer be projected precisely; by chance, there could be a run of births or a run of deaths in the population. Figure 1.4 shows a computer simulation of four populations that each began with 20 individuals and grew with stochastic births and deaths. Two of these populations actually declined below N_0 , even though r was greater than zero.

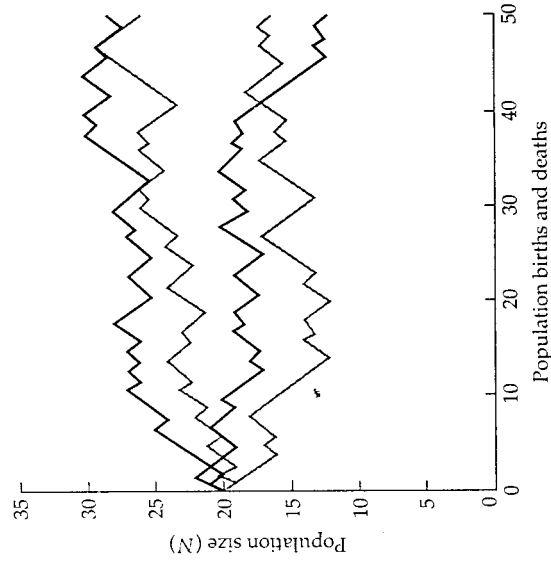


Figure 1.4 Computer simulation of population growth with demographic stochasticity. Each population track starts with an N of 20 individuals. $b = 0.55$ births/(individual • year) and $d = 0.50$ deaths/(individual • year). Although the starting conditions are identical, two of the populations actually dipped below the initial population size by the end of the simulation. Note that the x axis is not absolute time, but the number of sequential population events (births and deaths).

As in our analysis of environmental stochasticity, we are interested in the average population size and its variance. The average population size at time t is again given by:

$$\bar{N}_t = N_0 e^{rt} \quad \text{Equation 1.12}$$

which is the same as in the deterministic model. The equation for variance of population size depends on whether the birth and death rates are equal or not. If b and d are exactly equal, the population will not increase on average, and the variance in population size at time t is (Pielou 1969):

$$\sigma_{N_t}^2 = 2N_0 b t \quad \text{Equation 1.13}$$

If b and d are not equal, use the following:

$$\sigma_{N_t}^2 = \frac{N_0(b+d)e^{rt}(e^{rt}-1)}{r} \quad \text{Equation 1.14}$$

As in the model of environmental stochasticity, the variance in population size increases with time, and there is a risk of extinction even for populations with positive r . Demographic stochasticity is especially important at small population sizes because it doesn't take very many sequential deaths to drive a small population to extinction. Consequently, the probability of extinction depends not only on the relative sizes of b and d , but also on the initial population size. This probability of extinction is:

$$P(\text{extinction}) = \left(\frac{d}{b}\right)^{N_0} \quad \text{Equation 1.15}$$

For the chimpanzee example, if there were 50 chimps initially, the chance of extinction would be $(0.50/0.55)^{50} = 0.009 = 0.9\%$. However, if the initial population size were only 10 chimps, the chance of extinction would be $(0.50/0.55)^{10} = 0.386 = 38.6\%$.

Equations 1.13 and 1.14 also show that demographic stochasticity depends not only on the difference between b and d , but on the absolute sizes of b and d . Populations with high birth and death rates will be more variable than populations with low rates. Thus, a population with $b = 1.45$ and $d = 1.40$ will fluctuate more than a population with $b = 0.55$ and $d = 0.50$. In both popula-

tions, $r = 0.05$, but in the first, there is a much faster turnover of individuals, and thus a much greater chance for a run of several consecutive births or deaths.

To summarize, the average population size in stochastic models of exponential growth is the same as in the deterministic model we originally derived. In a stochastic world, populations can fluctuate because of changes in the environment that affect the intrinsic rate of increase (environmental stochasticity) and because of random birth and death sequences (demographic stochasticity). For both types of variability, a population can fluctuate so much that extinction is likely, even if the average intrinsic rate of increase is positive. Demographic stochasticity is much more important at small population sizes than at large.

Empirical Examples

PHEASANTS OF PROTECTION ISLAND

Humans have introduced many species into new environments, both intentionally and accidentally. Some of these introductions have turned out to be interesting ecological experiments. For example, in 1937, eight pheasants (*Phasianus colchicus torquatus*) were introduced onto Protection Island off the coast of Washington State (Lack 1967). The island had abundant food resources and no foxes or other bird predators. The island was too far from the mainland for pheasants to fly to it, so migration did not influence population size. From 1937 to 1942, the population increased to almost 2000 birds (Figure 1.5a,b). The curve shows a jagged increase that is similar to our discrete model of population growth. This increase reflects the fact that pheasant chicks hatch in the spring, and mortality continues throughout the year.

The initial population of eight birds had increased to 30 by the beginning of 1938. If we assume that resources were not limiting growth at this time, we can estimate λ as $(30/8) = 3.75$, with a corresponding r of $\ln(3.75) = 1.3217$ pheasants/(pheasant \cdot year). We can use this estimate to predict population size from the exponential growth model, and compare it to the actual size of the pheasant population each year. The initial predictions of this model were reasonably accurate, but after 1940, the model overestimated population size. By 1942, the population had grown to 1898 birds, whereas the model prediction was six times larger (11,488 birds). This difference probably reflects depletion of food resources on the island by the increasing pheasant population. Unfortunately, this interesting ecological experiment ended abruptly when the U.S. Army set up a training camp for World War II on the island, and promptly ate the pheasants!

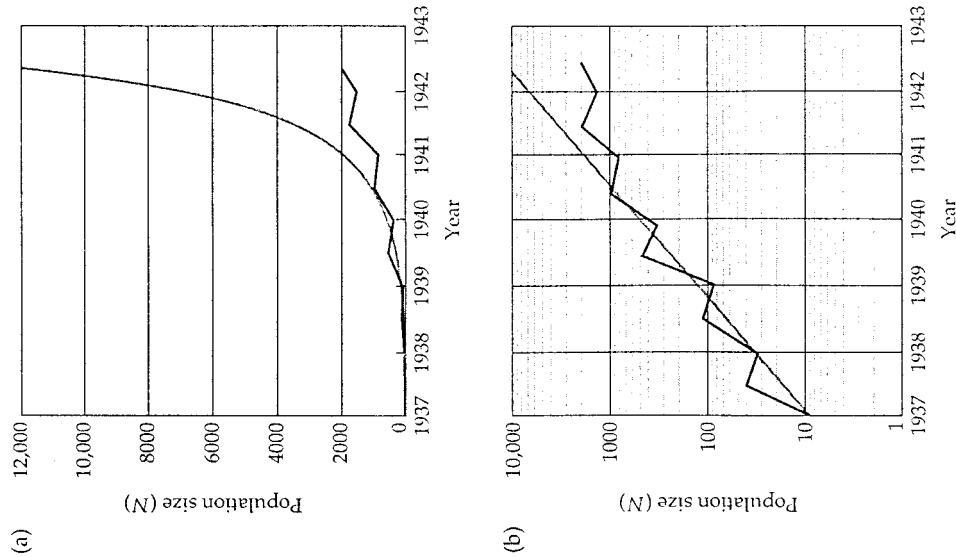


Figure 1.5 Growth of pheasant (*Plasiinthus colchicus torquatus*) population introduced to Protection Island. The thin line shows the hypothetical exponential growth curve, with $r = 1.3217$ individuals / (individual \cdot year); the thick line shows the observed population size. For comparison, population sizes are plotted on a linear scale in (a) and a logarithmic scale in (b). Note that the logarithmic scale is base 10, not base e . (Data from Lack 1967.)

GRIZZLY BEARS OF YELLOWSTONE NATIONAL PARK

The grizzly bear (*Ursus arctos horribilis*) was once widespread throughout most of North America. Today, its range in the lower 48 states consists of only six fragmented populations in the northwest, some of which have fewer than

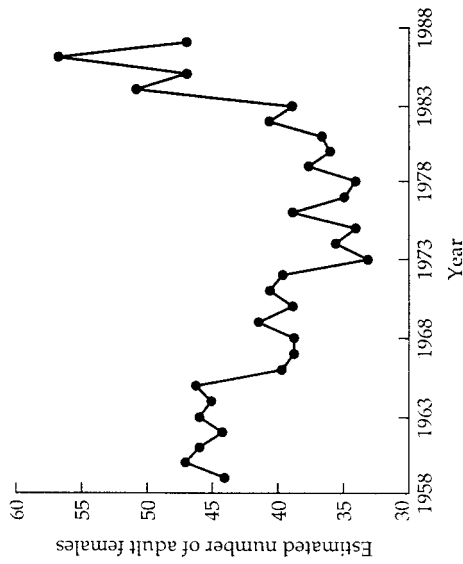


Figure 1.6 Population size of grizzly bears (*Ursus arctos horribilis*) in Yellowstone National Park. These data were used to construct a model of exponential population growth that incorporates environmental stochasticity. The estimate of r from this model was -0.003034 individuals / (individual \cdot year). (From Dennis et al. 1991.)

10 individuals. Yellowstone National Park supports one of the largest remaining populations, which fluctuates markedly from year to year (Figure 1.6).

The grizzly bear population data obviously do not conform to a simple exponential growth model, but they can be described by a more complex exponential model that incorporates environmental stochasticity (Dennis et al. 1991). The estimate of r that emerged from this model is -0.003034 bears / (bear \cdot year), suggesting that the population will decline slowly in the long run. However, the variance for this estimate was relatively large, so we should not be surprised to see periods of population increase. Based on this model, the prognosis for the Yellowstone grizzly bear population is not good. The model forecasts that the population is certain to drop below 10 individuals, at which point extinction is almost guaranteed. However, because r is close to zero and its variance is large, the estimated time to extinction is 200 years. Thus, the model suggests that it is unlikely the grizzly bear population is in immediate danger of extinction, but that the population is likely to reach a dangerously small size in the long run.

This projection assumes that background variability in b and d will continue in the future. Thus, the model does not incorporate catastrophic events, such as the 1988 Yellowstone fire, or future changes in human activity and management strategy, such as the 1970–1971 closure of the park garbage dumps, an important food source for the bears. Because this model is one of

exponential population growth in a stochastic environment, it does not incorporate resource limitation, which might lead to different predictions (see Chapter 2). Finally, the predictions of the model will change as additional data from yearly censuses become available. Increasingly, conservation biologists and park managers are using quantitative population models to estimate the risk of extinction for endangered species. Many of these models are based on the principles of exponential population growth that we have developed in this chapter.

Problems

1. At the time this book was written, the world's human population was expected to double in size in approximately 50 years. Calculate r for the human population. If the population size in 1993 was 5.4 billion, what is the projected population size for the year 2000?
2. You are studying a population of beetles of size 3000. During a one-month period, you record 400 births and 150 deaths in this population. Estimate b , d , and r and project the population size in six months.
3. For five consecutive days, you measure the size of a growing population of flatworms as 100, 158, 315, 398, and 794 individuals. Plot the logarithm (base e) of population size to estimate r .
4. A population of annual grasses increases in size by 12% every year. What is the approximate doubling time?
- *5. You are studying an endangered population of orchids, for which $b = 0.0021$ births/(individual \cdot year) and $d = 0.0020$ deaths/(individual \cdot year). The current population size is 50 plants. A new shopping mall is planned that will eliminate part of the orchid habitat and reduce the population to 30 plants. Estimate the effect of the proposed development on the probability of extinction.

* Advanced problem

Solutions

1. Rearranging Equation 1.3:

$$\begin{aligned} r &= \ln(2) / t = \ln(2) / 50 \text{ years} \\ &= 0.01386 \text{ individuals} / (\text{individual} \cdot \text{year}) \end{aligned}$$

$N_0 = 5.4$ billion and $t = 7$ years. From Equation 1.2 we have:

$$\begin{aligned} N_7 &= 5.4(e^{(0.01386)(7)}) \\ &= 5.95 \text{ billion humans} \end{aligned}$$

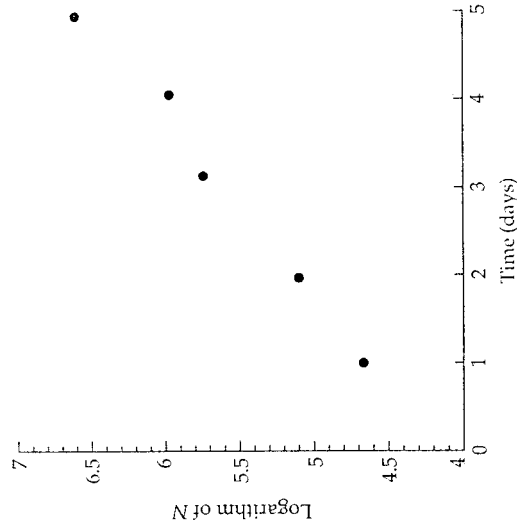
2. Following the approach in Expressions 1.6 and 1.7:

$$\begin{aligned} b &= 400 / 3000 = 0.1333 \text{ births} / (\text{individual} \cdot \text{month}) \\ d &= 150 / 3000 = 0.0500 \text{ deaths} / (\text{individual} \cdot \text{month}) \\ r &= b - d = 0.0833 \text{ individuals} / (\text{individual} \cdot \text{month}) \end{aligned}$$

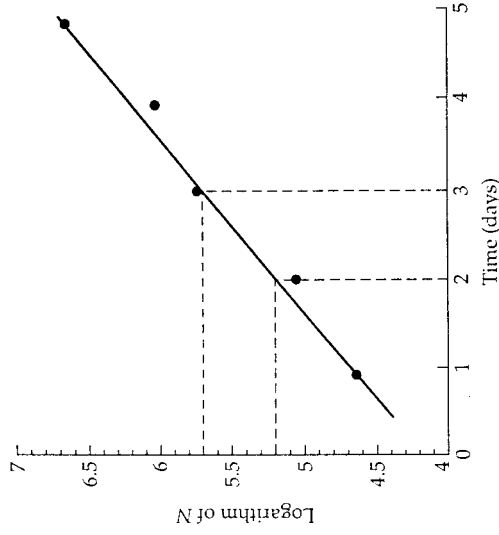
From Equation 1.2, the population size in six months will be:

$$N_6 = 3000(e^{(0.0833)(6)}) = 4945 \text{ beetles}$$

3. First, take the natural logarithms (base e) of the five consecutive population sizes, yielding: 4.605, 5.063, 5.753, 5.986, and 6.677. Next, plot these values as a function of time:



Because resources aren't limited, we can draw a straight line to fit all five of the data points. Although the points don't fall precisely on the line, this line gives a good estimate of population growth:



Finally, we measure the slope of this straight line to estimate r . The slope of a line is simply $(\Delta y) / (\Delta x)$. Using the dashed lines to calculate the slope, we have $(5.7 - 5.2) / (3 - 2) = 0.5$. So, our estimate of r for this flatworm population is:

$$r = 0.5 \text{ individuals} / (\text{individual} \cdot \text{day})$$

4. Because this is an annual species, we need to use the model of discrete population growth. If the population is increasing by 12% each year, $\lambda = 1.12$. From Equation 1.5:

$$r = \ln(\lambda) = \ln(1.12) = 0.113 \text{ individuals} / (\text{individual} \cdot \text{year})$$

Finally, we use Equation 1.3 to calculate the approximate doubling time:

$$\begin{aligned} t_{\text{double}} &= \ln(2) / r = \ln(2) / 0.113 \\ &= 6.1 \text{ years} \end{aligned}$$

5. For small populations growing with demographic stochasticity, the probability of extinction can be calculated from Equation 1.15. In the undisturbed state, the probability of extinction with a population size of 50 plants is:

$$P(\text{extinction}) = \left(\frac{d}{b}\right)N_0 = \left(\frac{0.0020}{0.0021}\right)^{50} = 0.087$$

If the shopping mall is built, we have:

$$P(\text{extinction}) = \left(\frac{d}{b}\right)N_0 = \left(\frac{0.0020}{0.0021}\right)^{30} = 0.231$$

So the proposed development threatens to increase the risk of extinction from about 9% to 23%.

CHAPTER 2

Logistic Population Growth



Model Presentation and Predictions

In Chapter 1, we assumed (unrealistically) that resources for population growth were unlimited. Consequently, the per capita birth and death rates, b and d , remained constant. We did explore some models in which b and d fluctuated through time (environmental stochasticity), but those fluctuations were **density-independent**; in other words, birth and death rates did not depend on the size of the population. In this chapter, we assume that resources for growth and reproduction are limited. As a consequence, birth and death rates depend on population size. To derive this more complex **logistic growth model**, we will start with the familiar growth equation:

$$\frac{dN}{dt} = (b - d)N \quad \text{Expression 2.1}$$

but now we will modify b and d so they are density-dependent and reflect crowding.

DENSITY DEPENDENCE

In the face of increased crowding, we expect the per capita birth rate to *decrease* because less food and fewer resources are available for organisms to use for reproduction. The simplest formula for a decreasing birth rate is a straight line (see Figure 2.1):

$$b = b_0 - aN \quad \text{Expression 2.2}$$

In this expression, N is population size, b is the per capita birth rate, and b_0 and a are constants. From Expression 2.2, the larger N is, the lower the birth rate. On the other hand, if N is close to zero, the birth rate is close to b_0 . The constant b_0 is the birth rate that would be achieved under ideal (uncrowded) conditions, whereas b is the actual birth rate, which is reduced by crowding. The constant a measures the strength of density dependence. The larger a is, the more sharply the birth rate drops with each individual added to the population. If there is no density dependence, then $a = 0$, and the birth rate equals b_0 , regardless of population size. Thus, the exponential growth model is a special case of the logistic model in which there are no crowding effects ($a = 0$).

Using similar reasoning, we can modify the death rate to reflect density dependence. In this case, we expect the death rate to *increase* as the population grows:

$$d = d_0 + cN \quad \text{Expression 2.3}$$

Again, the constant d_0 is the death rate when the population size is close to zero, and the population is growing (almost) exponentially. The constant c

measures the increase in the death rate from density dependence.

Expressions 2.2 and 2.3 are the simplest mathematical descriptions of the effects of crowding on birth and death rates. In real populations, the functions may be more complex. For example, b and d may not decline in a linear fashion; instead, there may be no change in b or d until a critical threshold density is reached. Some animals can reproduce, hunt, care for their offspring, or avoid predators more efficiently in groups than they can by themselves. For these populations, b may actually increase and d decrease as the population grows. This **Allee effect** (Allee et al. 1949) is usually important when the population is small, and may generate a critical minimum population size, below which extinction occurs (see Problem 2.3). But as the population grows, we expect negative density effects to appear as resources are depleted.

Note that *both* birth and death rates are density-dependent in this model. But it might be that only the death rate is affected by population size, and the birth rate remains density-independent, or vice versa. Fortunately, the algebra of this case works out exactly the same (see Problem 2.5). As long as either the birth rate *or* the death rate shows a density-dependent effect, we arrive at the logistic model.

Now we substitute Expressions 2.2 and 2.3 back into 2.1:

$$\frac{dN}{dt} = [(b_0 - aN) - (d_0 + cN)]N \quad \text{Expression 2.4}$$

After rearranging the terms:

$$\frac{dN}{dt} = [(b_0 - d_0) - (a + c)N]N \quad \text{Expression 2.5}$$

Next, we multiply Expression 2.5 by $[(b_0 - d_0)/(b_0 - d_0)]$. This term equals 1.0, so it does not change the results, but allows us to simplify further:

$$\frac{dN}{dt} = \left[\frac{(b_0 - d_0)}{(b_0 - d_0)} \right] [(b_0 - d_0) - (a + c)N]N \quad \text{Expression 2.6}$$

$$\frac{dN}{dt} = [(b_0 - d_0)] \left[\frac{(b_0 - d_0)}{(b_0 - d_0)} - \frac{(a + c)}{(b_0 - d_0)} N \right] N \quad \text{Expression 2.7}$$

Treating $(b_0 - d_0)$ as r , we have:

$$\frac{dN}{dt} = rN \left[1 - \frac{(a + c)}{(b_0 - d_0)} N \right] \quad \text{Expression 2.8}$$

CARRYING CAPACITY

Because a , c , b_0 , and d_0 are all constants in Expression 2.8, we can define a new constant K :

$$K = \frac{(b_0 - d_0)}{(a + c)} \quad \text{Expression 2.9}$$

The constant K is used for more than just mathematical convenience. It has a ready biological interpretation as the **carrying capacity** of the environment. K represents the maximum population size that can be supported; it encompasses many potentially limiting resources, including the availability of space, food, and shelter. In our model, these resources are depleted incrementally as crowding increases. Because K represents maximum sustainable population size, its units are numbers of individuals. Substituting K back into Expression 2.8 gives:

$$\frac{dN}{dt} = rN \left(1 - \frac{N}{K} \right) \quad \text{Equation 2.1}$$

Equation 2.1 is the logistic growth equation, which was introduced to ecology in 1838 by P.-F. Verhulst (1804–1849). It is the simplest equation describing population growth in a resource-limited environment, and it forms the basis for many models in ecology.

The logistic growth equation looks like the equation for exponential growth (rN) multiplied by an additional term in parentheses ($1 - N/K$). The term in parentheses represents the *unused portion of the carrying capacity*. As an analogy, think of the carrying capacity as a square frame that will hold a limited number of flat tiles, which are the individuals. If the population should ever exceed the carrying capacity, there would be more tiles than could fit in the frame. The unused portion of the carrying capacity is the percentage of the area of the frame that is empty (Krebs 1985).

For example, suppose $K = 100$ and $N = 7$. The unused portion of the carrying capacity is $[1 - (7/100)] = 0.93$. The population is relatively uncrowded and is growing at 93% of the growth rate of an exponentially increasing population $[rN(0.93)]$. In contrast, if the population is close to K ($N = 98$), the unused carrying capacity of the environment is small: $[1 - (98/100)] = 0.02$. Consequently, the population grows very slowly, at 2% of the exponential growth rate $[rN(0.02)]$. Finally, if the population should ever exceed carrying capacity ($N > K$), the term in parentheses becomes negative, which means that the growth rate is less than zero, and the population declines towards K . Thus, density-dependent birth and death rates provide an effective brake on exponential population growth.

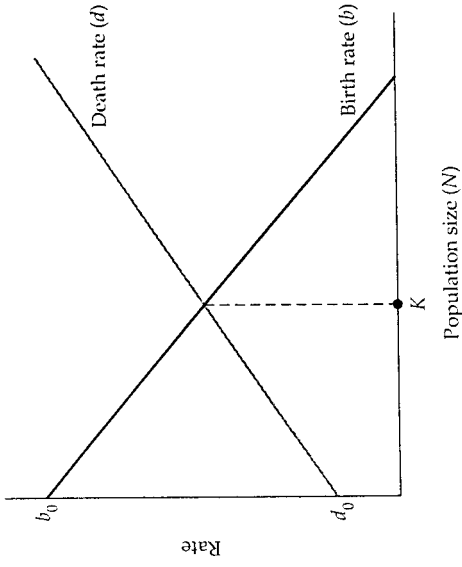


Figure 2.1 Density-dependent birth and death rates in the logistic model. The graph illustrates how the per capita rates of birth and death change as a function of crowding. The population reaches a stable equilibrium ($N = K$) at the intersection of the curves, where birth and death rates are equal.

When will the population stop growing? As in the exponential model, the rate of population growth (dN/dt) is zero when either r or N equals zero. But in the logistic model, the population will also stop growing when $N = K$. This is illustrated in Figure 2.1, which shows the density-dependent birth and death functions in the same graph. The two curves intersect at the point $N = K$ and form a **stable equilibrium**. The equilibrium is stable because no matter what the starting size of the population, it will move towards K . If N is less than K , we are at a point to the left of the intersection of the birth and death curves. In this region of the graph, the birth rate exceeds the death rate, so the population will increase. If we are to the right of the intersection point, the death rate is higher than the birth rate, and the population will decline.

As with the exponential growth model, we can use the rules of calculus to integrate the growth equation and express population size as a function of time:

$$N_t = \frac{K}{1 + [(K - N_0) / N_0]e^{-rt}} \quad \text{Equation 2.2}$$

From Equation 2.2, the graph of N versus time for logistic growth is a characteristic S-shaped curve (Figure 2.2). When the population is small, it

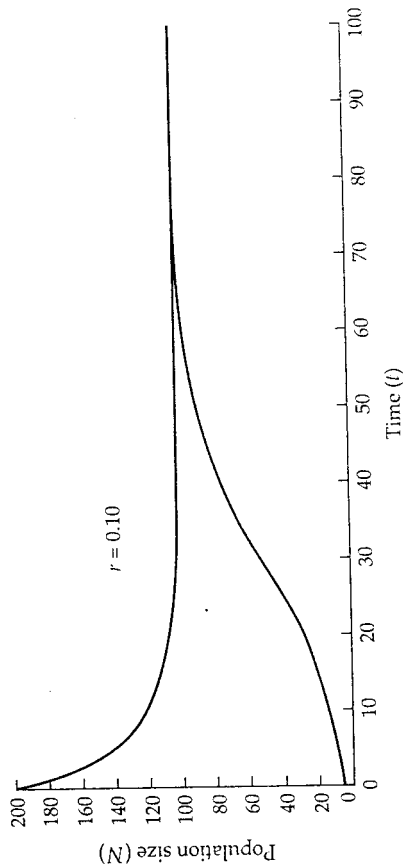


Figure 2.2 Logistic growth curve. The graph of N versus time increases in a characteristic S-shaped fashion when the population begins below carrying capacity. Above carrying capacity, the curve drops rapidly to the equilibrium point. In this example, $K=100$, and the starting population size is 5 or 200.

increases rapidly, at a rate slightly less than that predicted by the exponential model. The population grows at its highest rate when $N = K/2$ (the steepest point on the curve), and then growth decreases as the population approaches K (Figure 2.3a). This is in contrast to the exponential model, in which the population growth rate increases linearly with population size (Figure 2.3b). In the logistic model, if the population should begin above K , Equation 2.1 takes on a negative value, and N will decline towards carrying capacity.

Regardless of the initial number of individuals (N_0), a population growing according to the logistic model will quickly reach a fixed carrying capacity, which is determined solely by K . However, the time it takes to reach that equilibrium is proportional to r ; faster-growing populations reach K more quickly.

Model Assumptions

Because the logistic model is derived from the exponential model, it shares the assumptions of no time lags, migration, genetic variation, or age structure in the population. But resources are limited in the logistic model, so we make two additional assumptions:

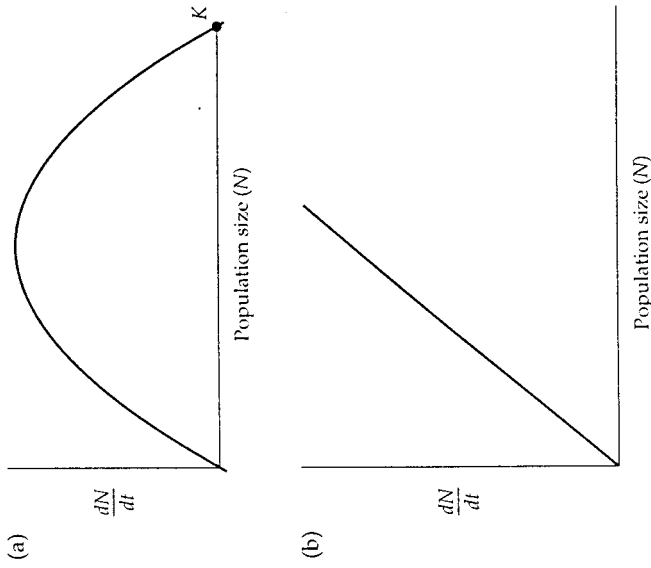


Figure 2.3 Population growth rate (dN/dt) as a function of population size. (a) Logistic growth. (b) Exponential growth.

✓ **Constant carrying capacity.** In order to achieve the S-shaped logistic growth curve, we must assume that K is a constant: resource availability does not vary through time. Later in this chapter, we will relax this assumption.

✓ **Linear density dependence.** The logistic model assumes that each individual added to the population causes an incremental decrease in the per capita rate of population growth. This is illustrated in Figure 2.4a, which shows the per capita growth rate ($1/N)(dN/dt)$ as a function of population size. This per capita rate is at its maximum value of $(b_0 - d_0) = r$ when N is close to zero, then declines linearly to zero when N reaches K . If N exceeds K , the per capita growth rate becomes negative. In contrast, the corresponding graph for the exponential growth model is a horizontal line because the per capita growth rate is independent of population size (Figure 2.4b).

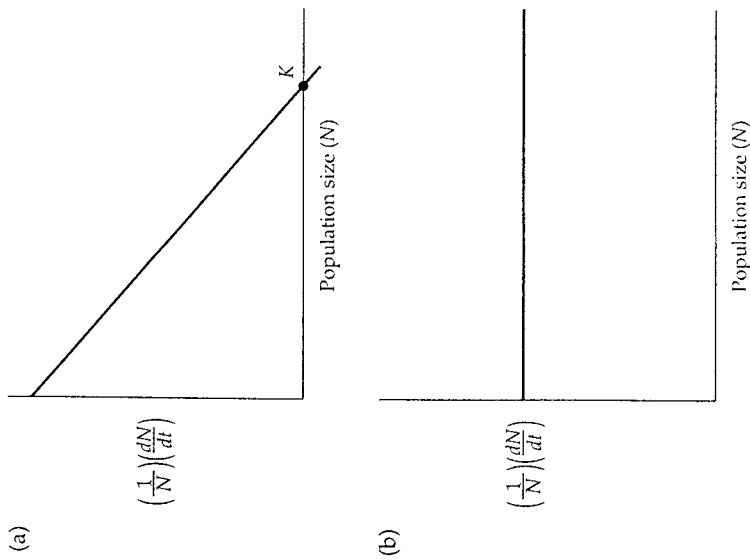


Figure 2.4 Per capita growth rates $(1/N)(dN/dt)$ as a function of population size. (a) Logistic growth. (b) Exponential growth.

Model Variations

TIME LAGS

The logistic growth model assumes that when another individual is added to the population, the per capita growth rate decreases immediately. But in many populations there may be **time lags** in the density-dependent response. For example, if a population of gulls increases in size in the fall, density dependence may not be expressed until the following spring, when females lay eggs. In a tropical rain forest, density-dependent mortality of mahogany trees (*Swietenia mahoganii*) may occur in the seedling stage, but density-dependent reproduction may not occur until 50 years later, when the trees first begin to flower. Individuals do not immediately adjust their growth and reproduction when resources change, and these delays can affect population

dynamics. Seasonal availability of resources, growth responses of prey populations, and age and size structure of consumer populations can introduce important time lags in population growth.

How can time lags be incorporated into our model? Suppose there is a time lag of length τ between the change in population size and its effect on population growth rate. Consequently, the growth rate of the population at time t (dN/dt) is controlled by its size at time $t - \tau$ in the past ($N_{t-\tau}$). Incorporating this time lag into the logistic growth equation gives:

$$\frac{dN}{dt} = rN \left(1 - \frac{N_{t-\tau}}{K} \right)$$

Equation 2.3

The behavior of this **delay differential equation** depends on two factors: (1) the length of the time lag τ , and (2) the "response time" of the population, which is inversely proportional to r (May 1976). Populations with fast growth rates have short response times ($1/r$).

The ratio of the time lag τ to the response time ($1/r$), or $r\tau$, controls population growth. If $r\tau$ is "small" ($0 < r\tau < 0.368$), the population increases smoothly to carrying capacity (Figure 2.5a). If $r\tau$ is "medium" ($0.368 < r\tau < 1.570$), the population first overshoots, then undershoots the carrying capacity; these **damped oscillations** diminish with time until K is reached (Figure 2.5b). The exact numerical values for these trajectories are not important. What is important is to understand how the behavior of the model changes as $r\tau$ is increased.

If $r\tau$ is "large" ($r\tau > 1.570$) the population enters into a **stable limit cycle**, periodically rising and falling about K , but never settling on a single equilibrium point (Figure 2.5c). The carrying capacity is the midpoint between the high and low points in the cycle. The cycle is stable because if the population is perturbed, it will return to these characteristic oscillations. When $r\tau$ is large, the time lag is so much longer than the response time that the population repeatedly overshoots and then undershoots K . The population resembles a heating system with a faulty thermostat that constantly overheats and then overcools, never achieving an equilibrium temperature.

Cyclic populations are characterized by their **amplitude** and **period** (Figure 2.5c). The amplitude is the difference between the maximum and the average population size. It is measured on the y axis of the graph of N vs. t , and its units are number of individuals. The larger the amplitude, the greater the population fluctuations. If the amplitude is too large, the population may hit the "floor" of zero and go extinct. The period is the amount of time it takes for one complete population cycle to occur. It is measured on the x axis, in units of time. The longer the period, the greater the amount of time between

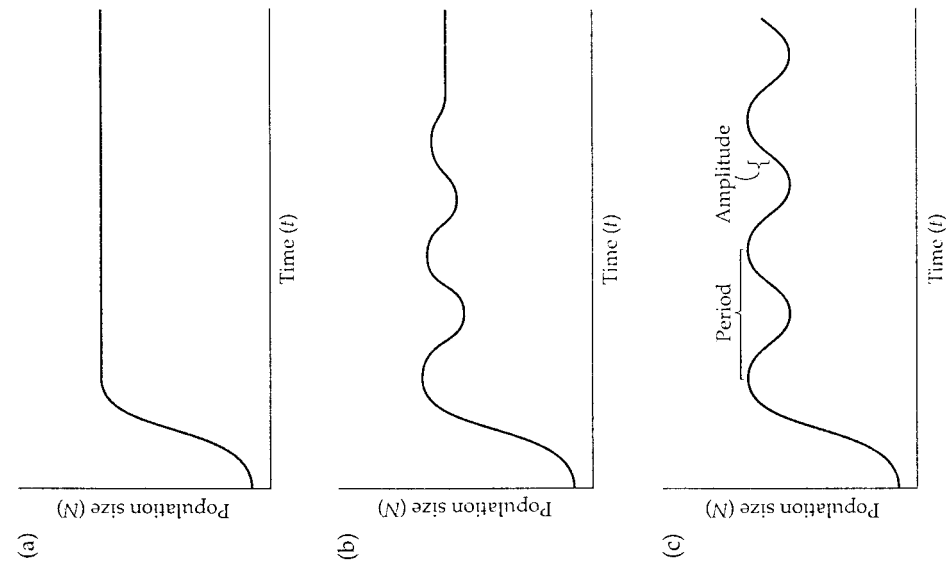


Figure 2.5 Logistic growth curves with a time lag. The behavior of the model depends on $r\tau$, the product of the intrinsic rate of increase and the time lag. (a) “Small” $r\tau$ behaves like the model with no time lag. (b) “Medium” $r\tau$ generates dampened oscillations and convergence on carrying capacity. (c) “Large” $r\tau$ generates a stable limit cycle and does not converge on the carrying capacity.

population peaks.

In a logistic model with a time lag, the amplitude of the cycle increases with increasing values of $r\tau$. This makes intuitive sense—if the population is growing very rapidly, or if the time lag is very long, the population will great-

ly overshoot K before it begins a phase of decline.

The period of the cycle is always about 4τ , regardless of the intrinsic rate of increase. Thus, a population with a time lag of one year can be expected to reach a peak density every four years. Why should the period of the cycle be four times as long as the lag? When the population reaches K , it will continue to increase for a length of time τ before starting to decrease. The distance from K to the population peak is about one-quarter of the cycle, so the length of the entire cycle is approximately 4τ . This result may explain the observation that many populations of mammals in seasonal, high-latitude environments cycle with peaks every three or four years (May 1976; see Chapter 6).

DISCRETE POPULATION GROWTH

We will now explore a model in which population growth is discrete rather than continuous. A discrete version of the logistic equation is:

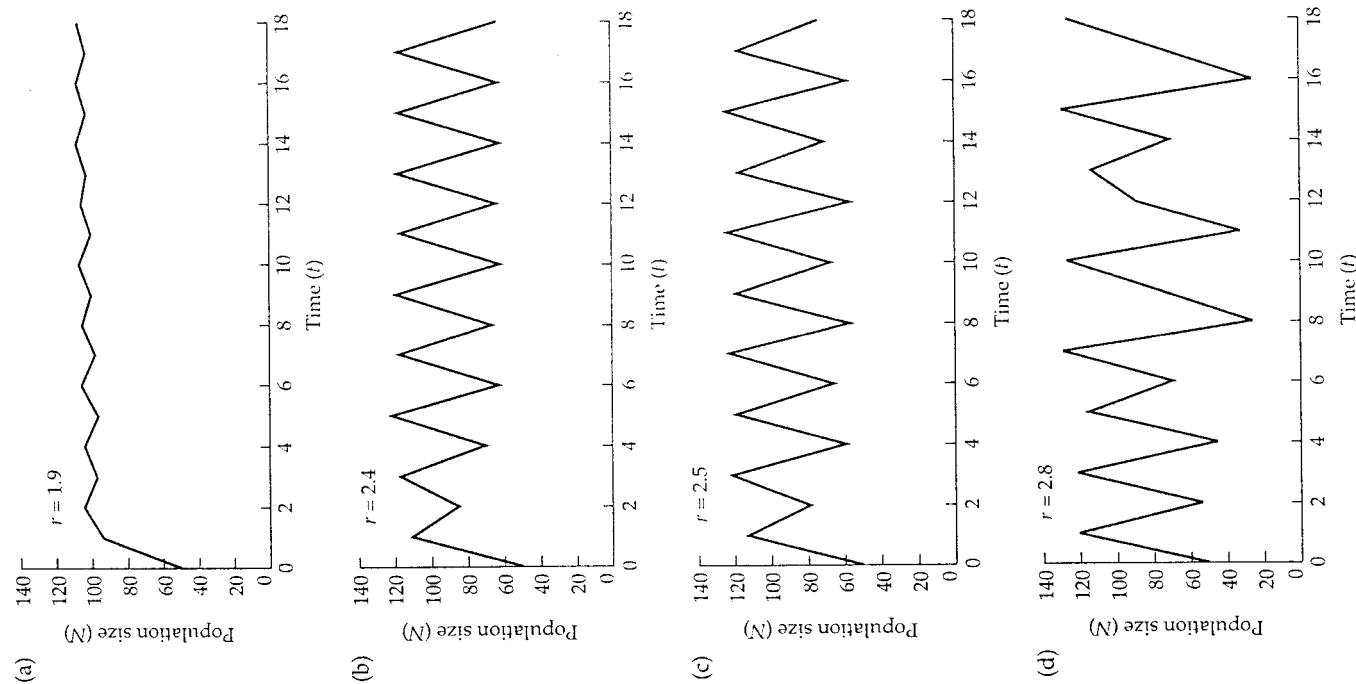
$$N_{t+1} = N_t + rN_t \left(1 - \frac{N_t}{K} \right) \quad \text{Equation 2.4}$$

This discrete growth logistic equation is analogous to the continuous model (Equation 2.1) in the same way that Equation 1.4 was analogous to the original exponential model (Equation 1.2).

A discrete population growth model has a built-in time lag of length 1.0. The population size at one time step in the future (N_{t+1}) depends on the current population size (N_t). In the last section, we saw that the product $r\tau$ controls the dynamics when a time lag is present. For the discrete model, the lag is of length 1.0, so the dynamics depend solely on r .

If r is not large, the behavior of this discrete equation is similar to that of its continuous cousin. At “small” r ($r < 2.000$), the population approaches K with damped oscillations (Figure 2.6a). At “less small” r ($2.000 < r < 2.449$), the population enters into a stable two-point limit cycle. This is similar to the continuous model, except that the population rises and falls to sharp “points,” rather than following a smooth curve. The points in the discrete model correspond to peaks and valleys of the cycle (Figure 2.6b). Between an r of 2.449 and an r of 2.570, the population grows with more complex limit cycles. For example, a four-point limit cycle has two distinct peaks and two distinct valleys before it starts to repeat. The number of points in the limit cycle increases geometrically (2, 4, 8, 16, 32, 64, etc.) as the value of r is increased in this interval (Figure 2.6c).

But if r is larger than 2.570, the limit cycles break down, and the population grows in a complex, nonrepeating pattern known as **chaos** (Figure 2.6d). Mathematical models of chaos are important in many areas of science, from the description of turbulent flow to the prediction of major weather patterns.



◀ **Figure 2.6** The behavior of the discrete logistic growth curve is determined by the size of r . (a) “Small” r generates damped oscillations ($r = 1.9$). (b) “Less small” r generates a stable two-point limit cycle ($r = 2.4$). (c) “Medium” r generates a more complex four-point limit cycle ($r = 2.5$). (d) “Large” r generates a chaotic pattern of fluctuations that appears random ($r = 2.8$).

Population biologists were among the first to appreciate that simple discrete equations may generate complex patterns (May 1974b). What is interesting about chaos is that seemingly random fluctuations in population size can emerge from a model that is entirely deterministic. Indeed, the track of a chaotic population may be so complex that it is difficult to distinguish from the track of a stochastic population.

However, chaos does not mean stochastic, or random, change. The fluctuations in a chaotic population have nothing to do with chance or randomness. Once the parameters of the model are specified (K , r , and N_0), the same erratic population track will be produced each time we run the model. The source of these erratic fluctuations is the density-dependent feedback of the logistic equation, combined with the built-in time lag of the discrete model. If we alter the starting conditions, say, by changing the initial population size (N_0), the populations will diverge more and more as time goes on (Figure 2.7).

In contrast, a truly stochastic population fluctuates because one or more of its parameters (r or K) changes with each time step. In a stochastic model, if we alter the starting population slightly, the population tracks will not diverge. In the next section we explore stochastic models in which the carrying capacity varies with time.

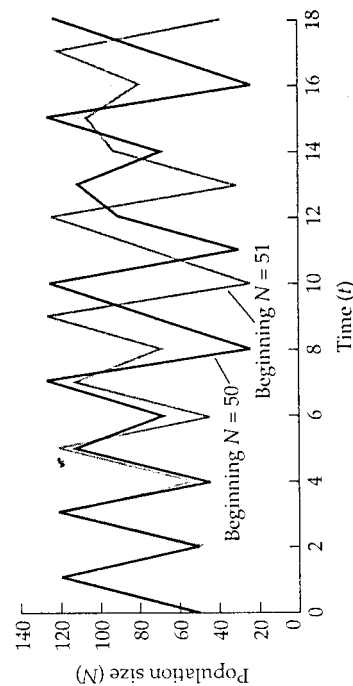


Figure 2.7 Divergence of population tracks with chaos. Both populations followed the same logistic equation, but the starting N for one of the populations was 50 and the other was 51. Note that, as more time passes, the two populations begin to diverge from one another.

RANDOM VARIATION IN CARRYING CAPACITY

In our analysis of environmental stochasticity (Chapter 1), we assumed that resources were unlimited, but that r varied randomly with time. For the logistic model, we will now assume that r is fixed, but that the carrying capacity varies randomly with time. Random variation in K means that the maximum population size that the environment can support changes unpredictably with time. How does this variation in resources affect the behavior of the logistic model? There are several mathematical approaches to the problem (May 1973; Roughgarden 1979), none of which yields a simple answer.

When r varied randomly in our exponential model, we found that the average population size was the same as in the deterministic model ($\bar{N}_t = N_0 e^{rt}$). So, you might reason that the average population size in the logistic model should approximate the average carrying capacity (\bar{K}). But this is not the case. Instead, \bar{N} will always be less than \bar{K} . Why should this be so? When a population is above K , it declines faster than a population that is increasing from a corresponding level below K (see Problem 2.4). This asymmetry is reflected in Figure 2.2, which shows that the population tracks above and below carrying capacity are not mirror images of one another. If the carrying capacity is described by its mean (\bar{K}) and variance (σ_K^2), a rough approximation to the average population size is (May 1974a):

$$\bar{N} \approx \bar{K} - \frac{\sigma_K^2}{2} \tag{Equation 2.5}$$

Thus, the more variable the environment, the smaller the average population size. The pattern of population fluctuations also depends on r (Levins 1969). Populations with large r are very sensitive to changes in K , and they will tend to track these fluctuations quite closely. Consequently, the average population size will be only slightly less than the average carrying capacity. In contrast, populations with small r are relatively sluggish and will not exhibit large increases or decreases (Figure 2.8); \bar{N} will be somewhat smaller than for populations with large r .

PERIODIC VARIATION IN CARRYING CAPACITY

Instead of random fluctuations in carrying capacity, suppose K varies repeatedly, in a cyclic fashion. Cyclic fluctuations in carrying capacity probably characterize many populations in seasonal temperate latitudes, and can be described with a cosine function (May 1976):

$$K_t = k_0 + k_1 [\cos(2\pi t / c)] \tag{Equation 2.6}$$

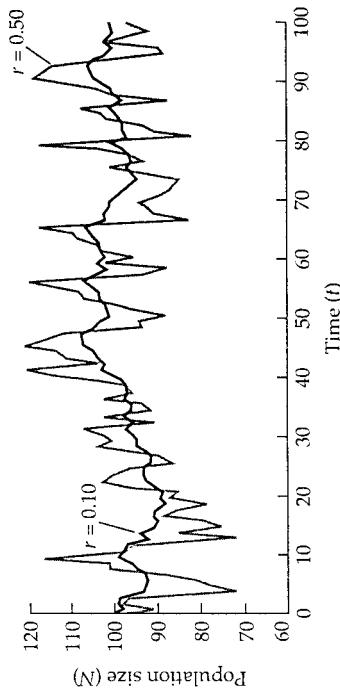


Figure 2.8 Logistic population growth with random variation in carrying capacity. Note that the population with the larger growth rate ($r = 0.50$) tracks the fluctuations in carrying capacity, whereas the population with the small growth rate ($r = 0.10$) is less variable and does not respond as quickly to fluctuations in resources.

Here, K_t is the carrying capacity at time t , k_0 is the mean carrying capacity, k_1 is the amplitude of the cycle, and c is the length of the cycle. As t increases, the cosine term in parentheses varies cyclically from -1 to 1 . Thus, during a single cycle of length c , the carrying capacity of the environment varies from a minimum of $k_0 - k_1$ to a maximum of $k_0 + k_1$.

How does this cyclic variation in carrying capacity affect population growth? The length of the carrying capacity cycle functions as a kind of time lag, so once again, the behavior of the model depends on rc . If rc is small (< 1.0), the population tends to "average" the fluctuations in the environment and persists at roughly:

$$\bar{N} \approx \sqrt{k_0^2 - k_1^2} \tag{Equation 2.7}$$

Thus, if rc is small, \bar{N} is less than \bar{K} , and the reduction is greater when the amplitude of the cycle is large; both patterns are similar to the results for a population in which K varies stochastically. If rc is large (> 1.0), the population tends to track the fluctuations in the environment:

$$N_t \approx k_0 + k_1 \cos(2\pi t / c) \tag{Equation 2.8}$$

although at a value slightly less than the actual carrying capacity (Figure 2.9). In conclusion, both stochastic and periodic variation in carrying capacity

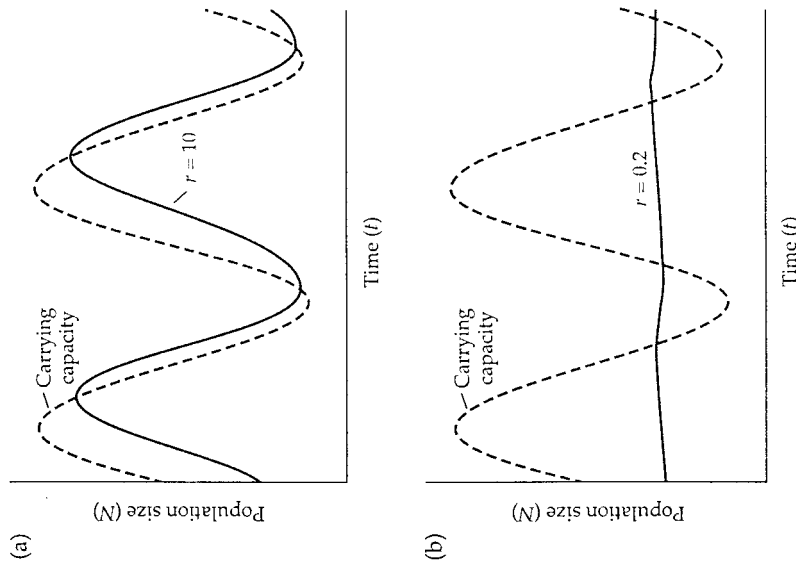


Figure 2.9 Logistic growth with periodic variation in the carrying capacity. The carrying capacity of the environment varies according to a cosine function. As with random variation, the population with the large growth rate ($r = 10$) tends to track the variation (a), and the population with the small growth rate ($r = 0.2$) tends to average it (b). The dashed line indicates K . (From May 1976.)

reduce populations, and the more variable the environment, the lower the average population size. In a variable environment, populations with large r , such as most insects, may be expected to track variation in carrying capacity, whereas populations with small r , such as large mammals, may be expected to average the environmental variation and remain relatively constant.

Empirical Examples

SONG SPARROWS OF MANDARTE ISLAND

Mandarte Island is a rocky, 6-hectare island off the coast of British Columbia. The island is home to a population of song sparrows (*Melospiza melodia*) that has been studied for many decades (Smith et al. 1991). On average, only one new female migrant joins this population each year, so most of the changes in population size are due to local births and deaths. Over the past 30 years, the population has varied between 4 and 72 breeding females and between 9 and 100 breeding males. The sparrow population of Mandarte Island does not conform to a simple logistic growth model; population size is variable and there have been periods of increase followed by rapid declines (Figure 2.10). Some of these, such as the crash in 1988, were caused by an unusually cold winter and an increased death rate. Other declines were not correlated with any obvious change in the environment.

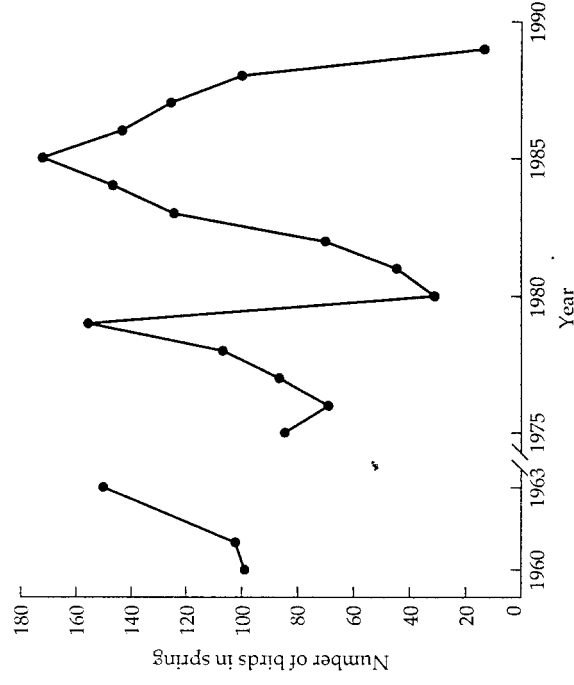


Figure 2.10 Population size of the song sparrow (*Melospiza melodia*) on Mandarte Island. (After Smith et al. 1991.)

Although this population is clearly buffered by density-independent changes, there is good evidence of underlying density dependence. Male song sparrows defend territories that determine their breeding success, but limited food resources and space prevent many males from ever establishing territories. These nonterritorial "floaters" are behaviorally submissive individuals. Their proportion increased in a density-dependent fashion as the population became more crowded (Figure 2.11a). When the resident territory holders were experimentally removed, floater males quickly took over their territories, so the total breeding population size remained relatively constant.

Density dependence is also seen in the number of surviving young produced per female (Figure 2.11b), and in the survival of juveniles (Figure 2.11c), both of which decreased as the population size increased. Experimental studies confirmed that food limitation was the controlling factor: when food levels for sparrows were artificially enhanced, female reproductive output increased fourfold (Arcese and Smith 1988). Thus, both territoriality and food limitation generated density-dependent birth and death rates in song sparrows.

Nevertheless, although density dependence has the potential to control population sizes, the risk of extinction for Mandarte Island sparrows probably comes from unpredictable environmental catastrophes and other density-independent forces. Somewhat paradoxically, it is these density-independent fluctuations that allow us to detect density dependence, because they push the population above or below its equilibrium and reveal the underlying dynamics of birth and death rates.

POPULATION DYNAMICS OF SUBTIDAL ASCIDIANS

Ascidians, or "sea squirts," are filter-feeding invertebrates that live attached to pier pilings and rock walls. These animals are important components of subtidal "fouling" communities throughout the world. Ascidians are actually primitive chordates that disperse with a sexually produced tadpole larva. The perennial ascidian *Ascidia mentula* has been the subject of a long-term study of population dynamics on vertical rock walls off the Swedish west coast (Svane 1984).

Six populations were monitored continually for 12 years with photographs of permanent plots. At sheltered sites within a fjord, density was highest in shallow plots; at exposed stations, density was highest in deep-water plots. At all sites, populations fluctuated considerably (Figure 2.12), in contrast to the predictions of the basic logistic model. Mortality was primarily due to "bulldozing" by sea urchins and temperature fluctuations. These factors seemed to operate in a density-independent fashion, because there was no relationship between mortality rate and population size (Figure 2.13a). In

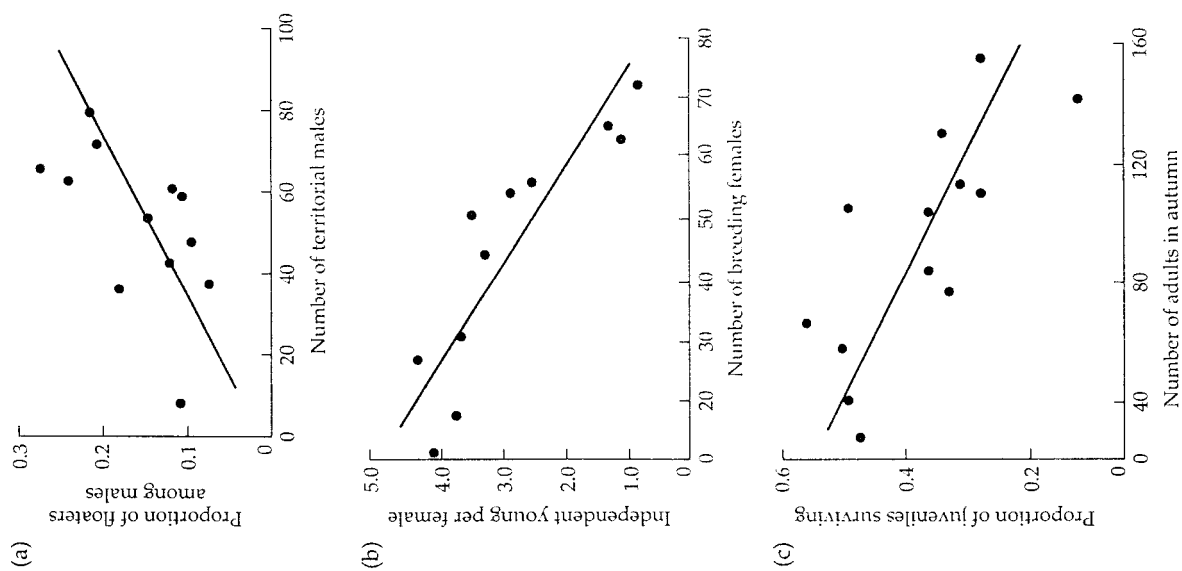


Figure 2.11 Density dependence in the Mandarte Island song sparrow (*Melospiza melodia*) population. As the population becomes more crowded (a) the proportion of nonterritorial "floater" males increases; (b) the number of surviving young produced per female decreases; (c) juvenile survival decreases. (After Arcese and Smith 1988 and Smith et al. 1991.)

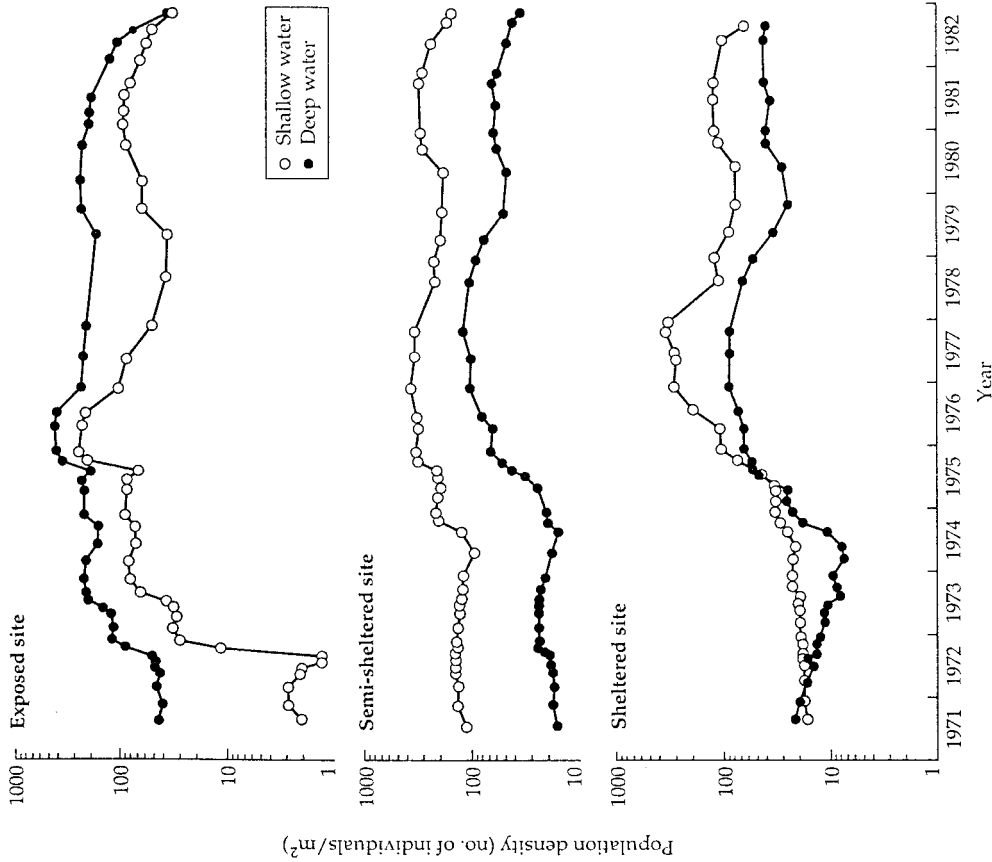


Figure 2.12 Population density of ascidians (*Ascidia mentula*) at six subtidal sites off the coast of Sweden. Population densities are greater in shallow water than in deep, except at the exposed site. Note the use of a logarithmic scale for the *y* axis, which diminishes the appearance of population fluctuations. (After Svane 1984.)

contrast, reproduction (as measured by larval recruitment) was density-dependent and decreased at high densities. At low densities, there was evidence of an Allee effect: recruitment actually increased with population density until a density of approximately 100 animals per square meter was

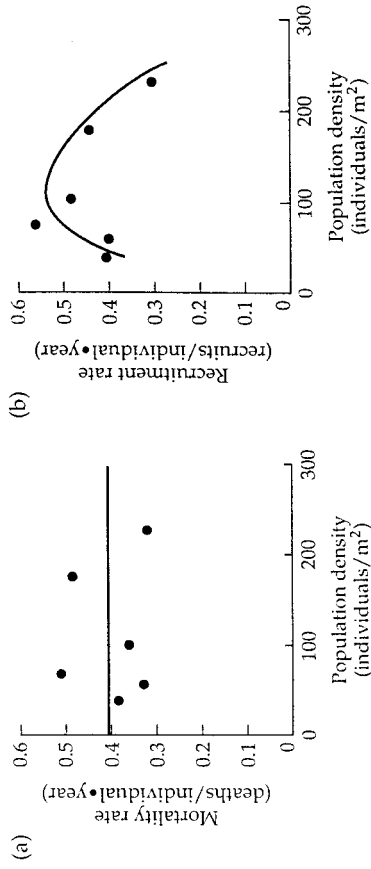


Figure 2.13 (a) Density-independent mortality rates. The mortality rate of ascidians (*Ascidia mentula*) at the six population sites appears to be independent of population size. (b) Density-dependent recruitment rates. The rate of recruitment of new juveniles into ascidian populations is density-dependent and is lower in more dense populations. Note the appearance of a possible Allee effect, as recruitment is also decreased at sites with very low abundance. (After Svane 1984.)

reached (Figure 2.13b). Possible explanations for this Allee effect include the behavioral attraction of larvae to established adults and entrapment of larvae by local water currents.

Like the Mandarte Island sparrows, these ascidians showed some evidence of underlying density dependence, although the population never reached a steady carrying capacity. Both the ascidian and sparrow populations were affected by temperature fluctuations, although these effects seemed more subtle and long-term for the ascidians. Unlike the isolated sparrow population, the ascidian populations were potentially linked by larval dispersal between sites, so that a realistic model of population dynamics might be especially complex (see Chapter 4).

LOGISTIC GROWTH AND THE COLLAPSE OF FISHERIES POPULATIONS

How many tons of fish should be harvested each year to maximize long-term yield? This **optimal yield** problem has been very important to commercial fisheries because of the huge amounts of money involved and because overfishing has been a problem since at least the 1920s, when commercial stocks of many species started to decline. The logistic growth curve provides a simple, though often unpopular, prescription for optimal fishing strategies.

The optimal strategy is the one that maximizes the population growth rate, because this rate determines how quickly fish can be removed from the pop-

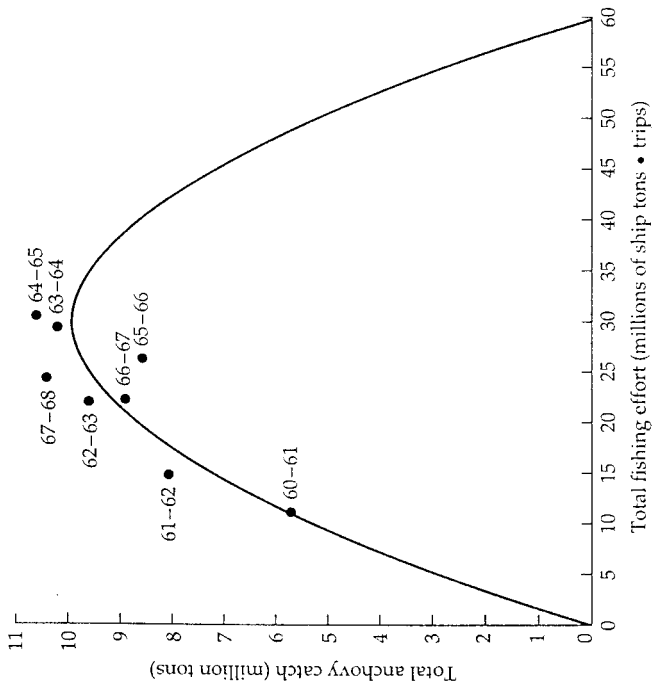


Figure 2.14 Relationship between fishing effort and total catch for the Peruvian anchovy (*Engraulis ringens*) fishery. Each point represents the fishing catch and effort for a particular year. The data include fishing effort by humans and fish catches by seabird populations. The parabola is drawn by fitting the logistic model to data from Boerema and Gulland (1973). (After Krebs 1985.)

ulation while still maintaining a constant stock for future production. If a population is growing according to the logistic equation (or some other model that incorporates a carrying capacity), maximum population growth rate occurs if the population is held at $K/2$, half the carrying capacity (Figure 2.3a). Two other strategies are guaranteed to produce low yields. The first is to be extremely conservative and remove very few animals at each harvest. This keeps the standing stock large, but the yield is low because the population is close to carrying capacity and grows slowly. The other strategy is to harvest the population down to a very small size. This also produces low yield because there are so few individuals left to reproduce.

Unfortunately, this latter strategy of overdepletion has been followed by all the world's fisheries. Figure 2.14 shows the yearly catch of Peruvian anchovy (*Engraulis ringens*) fitted to the predictions of a simple logistic model. The

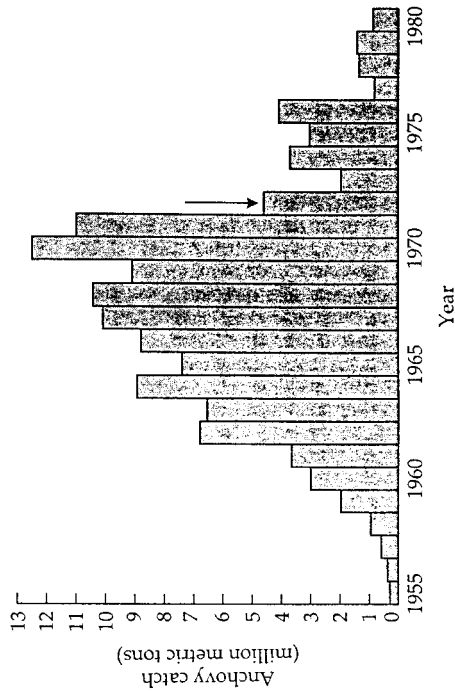


Figure 2.15 Total catch for the Peruvian anchovy (*Engraulis ringens*) fishery from 1955 to 1981. This was the largest fishery in the world until its collapse in 1972. (After Krebs 1985; unpublished data from M. H. Glanz.)

model predicts a maximum sustained yield of approximately 10 to 11 million metric tons per year. The annual catch was close to this sustained maximum from 1964 to 1971. In 1972, the Peruvian anchovy fishery collapsed, in part due to overfishing, and in part due to an El Niño event, in which a warm tropical water mass moved off the coast of Peru and greatly reduced productivity. Although fishing was reduced to allow stocks to recover, anchovy populations have never reached their former abundance and fishing yields remain low (Figure 2.15). Increasingly sophisticated technology and large factory-ships have depleted world stocks of many fish populations to the point where the industry itself is doomed to economic collapse. In 1989, for example, the cost of operating the world's 3 million fishing vessels was estimated at \$92 billion, whereas the total catch was worth only \$72 billion (Pitt 1993). The disappearance of human societies that depend on fishing is also inevitable.

The situation can only be remedied by worldwide restrictions on fishing and short-term reductions in catch. Unfortunately, this will not be easy because each individual fishing vessel tries to maximize its short-term yield by intensive fishing. Migratory fish populations do not obey political boundaries, making international policies difficult to enforce. The problem of short-term versus long-term profits in the exploitation of natural resources is known as "the tragedy of the commons" (Hardin 1968).

Problems

1. Suppose a population of butterflies is growing according to the logistic equation. If the carrying capacity is 500 butterflies and $r = 0.1$ individuals/(individual \cdot month), what is the maximum possible growth rate for the population?
2. A fisheries biologist is maximizing her fishing yield by maintaining a population of lake trout at exactly 500 individuals. Predict the population growth rate if the population is stocked with an *additional* 600 fish. Assume that r for the trout is 0.005 individuals/(individual \cdot day).
3. You are studying a density-dependent turtle population that has the following relationships for the birth rate (b) and the death rate (d) as a function of population size (N):

$$b = 0.10 + 0.03N - 0.0005N^2$$

$$d = 0.20 + 0.01N$$

Plot these functions in the same graph and discuss the population dynamics of the turtle. How does this model differ from the simple logistic model with linear birth and death functions?

- *4. Prove that the decline of a population above its carrying capacity is always faster than the corresponding increase below carrying capacity. (The starting population above carrying capacity should be represented as $K + x$.)
- *5. In our derivation of the logistic equation, we assumed that both the birth and the death rates were density-dependent. Prove that the logistic model holds for a population in which the birth rate is density-dependent and the death rate is density-independent. Use the same approach as in Expressions 2.1 to 2.9.
- *6. Tropical populations of many organisms experience seasonal variation in rainfall and food supply, even though temperatures are fairly constant year-round. Suppose that a water-filled tropical tree hole has a carrying capacity of 500 mosquito larvae. The water level in the hole declines gradually through the dry season, so the carrying capacity varies seasonally between 250 and 750 larvae. If the population is slow-growing, what is the long-term average population size, and what sort of temporal fluctuations in population size would you expect to see? Assume that $rc \ll 1.0$.

* Advanced problem

Solutions

1. To solve this problem, we first need to determine N , the population size. From Figure 2.3a, we know that the maximum possible growth rate for a population growing according to the logistic model occurs when $N = K/2$, so $N = 250$ butterflies. Plugging these values into Equation 2.1 gives:

$$\frac{dN}{dt} = rN \left(1 - \frac{N}{K} \right)$$

$$\frac{dN}{dt} = 0.1(250) \left[1 - (250/500) \right]$$

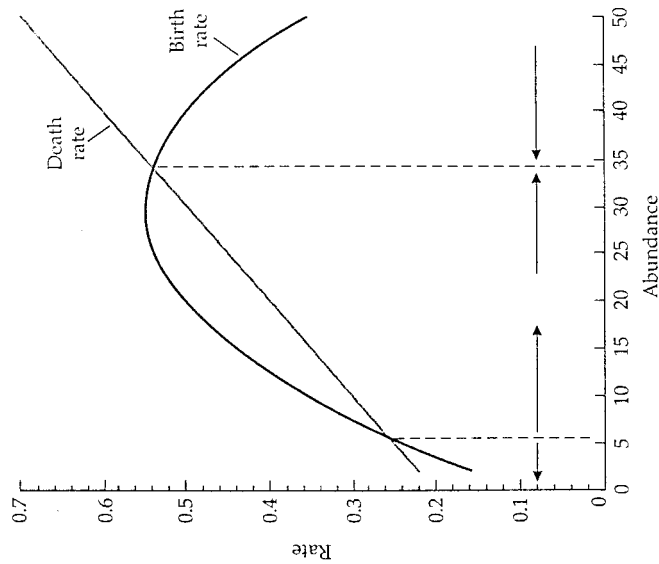
$$\frac{dN}{dt} = 12.5 \text{ individuals / month}$$

2. For a population growing according to the logistic equation, we know that the maximum population growth rate occurs at $K/2$, so K must be 1000 fish in this case. If the population is stocked with an additional 600 fish, the total size will be 1100. From Equation 2.1, the growth rate will be:

$$\frac{dN}{dt} = (0.005)(1100) \left[1 - (1100/1000) \right]$$

$$\frac{dN}{dt} = -0.55 \text{ fish / day}$$

- The growth rate is negative because the additional stock pushes the population above its carrying capacity.
3. The equation for the death rate is linear; as in the simple logistic, the greater the number of turtles in the population, the greater the death rate. However, the equation for the birth rate is quadratic; it includes an N^2 term. This quadratic equation generates an Allee effect for reproduction: the birth rate first increases and then decreases with population size. Substitute different values of N into the birth and death functions to construct the graph seen on page 52.
- Notice that the birth and death curves intersect in two different places. These points represent two different equilibrium population densities. One point of intersection is at a population size of approximately 34 turtles. If we are to the right of this equilibrium, the death rate exceeds the birth rate and the population declines, as shown by the arrow pointing to the left. If the population is less than 34, the birth rate exceeds the death rate and the population increases, as shown by the right-pointing arrow. Thus, this larger equilibrium point is stable.



The smaller equilibrium occurs at a population size of approximately 6 turtles. If the population is greater than 6, the birth rate exceeds the death rate, and the population will continue to increase until the equilibrium of 34 is reached. But if the population is less than 6, the death rate exceeds the birth rate, and the population declines to zero. Thus, this second equilibrium is unstable. By incorporating an Allee effect into the birth rate, we generate a minimum population size (6) that is necessary for the population to persist. This result is in contrast to the simple logistic model, in which the population always increased as long as it was below carrying capacity.

4. We wish to compare the growth rates of two populations, one of which is x individuals above carrying capacity, and one of which is x individuals below carrying capacity. For the first population, let $N = K + x$. Substituting into Equation 2.1 gives:

$$\frac{dN}{dt} = r(K+x) \left(1 - \frac{K+x}{K}\right)$$

For the population that is below carrying capacity, $N = K - x$, so its growth is represented by:

$$\frac{dN}{dt} = r(K-x) \left(1 - \frac{K-x}{K}\right)$$

In order to determine which growth rate is larger, we compare the size of both equations by factoring out equivalent terms:

$$r(K+x) \left(1 - \frac{K+x}{K}\right) \stackrel{?}{\leftarrow} r(K-x) \left(1 - \frac{K-x}{K}\right)$$

Dividing through by r and substituting K/K for 1 gives:

$$(K+x) \left(\frac{K}{K} - \frac{K+x}{K}\right) \stackrel{?}{\leftarrow} (K-x) \left(\frac{K}{K} - \frac{K-x}{K}\right)$$

After subtracting and dividing through by K , this simplifies to:

$$(K+x)(-x) \stackrel{?}{\leftarrow} (K-x)(x)$$

Notice that the expression on the left is negative, because this is the growth rate when the population is above K . Because we are interested in the magnitude of growth, we take the absolute value of both sides of the inequality. Then, dividing through by x yields:

$$(K+x) > (K-x)$$

This result proves that the decline of a population above its carrying capacity is always faster than the increase from below carrying capacity. For this reason, the average population size will always be less than the average carrying capacity in a variable environment.

5. If the birth rate is density-dependent and the death rate is density-independent then:

$$b = b_0 - aN$$

$$d = d_0$$

Substituting these two terms back into Equation 2.1 gives:

$$\frac{dN}{dt} = (b_0 - aN - d_0)N$$

$$\frac{dN}{dt} = [(b_0 - d_0) - aN]N$$

Treating $(b_0 - d_0)$ as r , we have:

$$\frac{dN}{dt} = rN \left(1 - \frac{aN}{r}\right)$$

Because a and r are both constants, we can define K as r/a , which again leads to the logistic equation:

$$\frac{dN}{dt} = rN \left(1 - \frac{N}{K} \right)$$

You can think of this as a special case of Expression 2.5, in which the constant c equals zero because the death rate is density-independent.

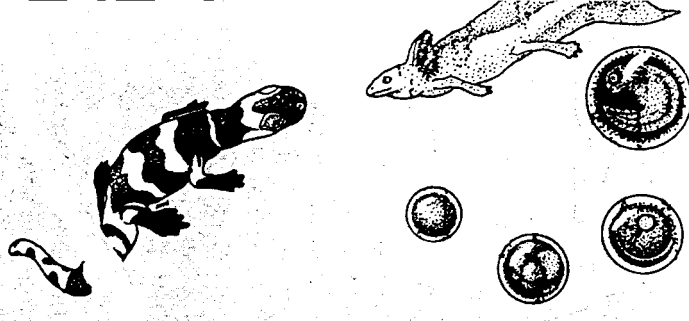
6. Because this is a slow-growing population with seasonal fluctuations in carrying capacity, we can use Equation 2.7, with a mean carrying capacity of 500 larvae, and an amplitude of 250 larvae. With these values, the average population size is predicted to be:

$$\sqrt{(500)^2 - (250)^2} = 433 \text{ larvae}$$

Because the population growth rate is slow, we expect this population to respond sluggishly to the seasonal changes in carrying capacity and not show much variation in population size.

CHAPTER 3

Age-Structured Population Growth



Model Presentation and Predictions

EXPONENTIAL GROWTH WITH AGE STRUCTURE

In Chapter 1, we represented birth and death rates as single constants (b and d), which allowed us to easily calculate r for a population with exponential growth. The resulting model was appropriate for “simple” organisms such as single-celled bacteria or protozoa. But for most plants and animals, birth and death rates depend on the *age* of an individual.

For example, a newborn elephant cannot reproduce immediately, but must grow for a decade or more before it is reproductively mature. Death rates also vary with age. Seeds, larvae, and hatchlings usually have higher mortality rates than older age classes. Death rates also tend to be high for the very oldest individuals in a population, which may be more vulnerable to predators, parasites, and disease.

The age structure of a population has the potential to affect population growth. For example, if a population consisted only of tadpoles, it would not begin to grow until the tadpoles had metamorphosed into frogs and reached sexual maturity. In contrast, if a population of monkeys consisted only of old, postreproductive individuals, it would decline to extinction.

In this chapter, we will learn how to calculate r for a population in which birth and death rates depend on the age of an organism. Next, we will illustrate the short-term changes in age structure of a population that occur before it settles into a pattern of steady exponential growth. We will briefly consider the problem of life history strategies—why natural selection tends to favor certain birth and death schedules. Finally, we will develop a model of population growth for organisms with complex life histories, such as corals and perennial plants, that do not exhibit simple age structure.

Many students find the analysis of life tables to be one of the most confusing topics in ecology. Admittedly, the calculations in this chapter are tedious—we have to keep track of the birth rate, death rate, and number of individuals in each age class of the population. Be careful with your subscripts, but try not to get bogged down in notation. Keep in mind that we are still using a simple model of exponential growth in an environment with unlimited resources. In that sense, the concepts presented in this chapter are no different than those in Chapter 1.

NOTATION FOR AGES AND AGE CLASSES

To begin our analysis, we need some notation to keep track of the different ages and age classes in a population. Technically, we are modeling a population with continuous births and deaths. However, because we are classifying individuals into discrete age classes, our calculations will represent approximations to continuous growth. There is more than one way to approximate

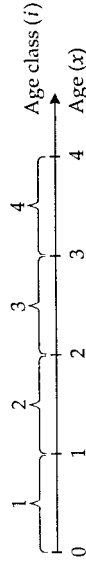


Figure 3.1 The relationship between ages (x) and age classes (i) in population growth models. (From Caswell 1989.)

these continuous functions, and the equations we use depend on the timing of the population censuses and the seasonal pattern of births and deaths.

We use the variable x in parentheses to refer to the **age** of an individual. For our discussion, the units of x will be years. However, any convenient time interval can be used, and the choice will usually be based on the life span of the organism and the type of census data that are available. By convention, we classify a newborn as age 0 (*not* age 1). An individual is age 0 at birth, age 0.5 at 6 months, and age 1 at its first birthday, which is the start of the second year. We use the constant k to refer to the final age in the life table, that is the age by which all individuals have died. Thus, x is a number whose value ranges from 0 to k . The number of ages in the life table depends on the length of the census interval and the life span of the organism.

Equivalently, we can designate the age of an individual by its **age class**. An individual in age class x is between the ages of $x - 1$ and x . For example, an individual in the third age class is between the ages of 2 and 3. Similarly, a newborn is of age 0, but is in the first age class. If the ages in the population range from 0 to k , the age classes range from 1 to k . The age-class counter is always one greater than the age counter (Figure 3.1). To keep the distinction clear, variables that indicate age will appear in parentheses, whereas variables that indicate age class will be designated by a subscript. For example:

$$f(5) = f_6 \quad \text{Expression 3.1}$$

Expression 3.1 says that the value of f for individuals of age 5 is equivalent to the value of f for individuals of age class 6.

We can analyze our demographic model using the notation of either ages or age classes. We will follow the textbook tradition of using the age notation to describe the life-table analysis. However, we will switch to the age class notation to describe population growth and the analysis of complex life cycles.

THE FERTILITY SCHEDULE $|f(x)|$

The fertility schedule consists of the average number of offspring born to an individual female of a particular age. The fertility schedule is a column of values represented as $b(x)$ or $m(x)$, abbreviations for birth or maternity. For

example, if $b(6) = 3$, a female of age 6 will give birth to an average of 3 offspring. Thus, the $b(x)$ schedule gives per capita fertility rates for females. Technically, we should be modeling the numbers of both males and females, because the population sex ratio will affect the growth rate. However, as long as the sex ratio is approximately 50/50, we can reasonably model population growth by ignoring the males.

The entries in the fertility schedule are non-negative real numbers. An entry of zero in the fertility schedule means that individuals of a particular age do not reproduce. The fertility schedule gives the *average* reproduction for a female of a particular age, so these numbers do not have to be integers, and may be less than 1.0 for ages with very little reproduction.

Table 3.1 gives a hypothetical life table for an organism that lives to the end of its fourth year. The ages are 0 through 4, and the age classes are 1

through 4. We will use the data in Table 3.1 to illustrate all the calculations necessary for a typical life-table analysis. If you look at the $b(x)$ column, you see that newborns do not reproduce. One-year-olds produce an average of 2 offspring, two-year-olds produce 3 offspring, and three-year-olds produce 1 offspring.

FERTILITY SCHEDULES IN NATURE

In nature, what sorts of fertility schedules do we find? Animal ecologists distinguish between **semelparous** and **iteroparous** reproduction. Plant ecologists use the equivalent terms **monocarpic** and **polycarpic**. In semelparous (monocarpic), or “big bang” reproduction, an organism reproduces only once in its lifetime. Examples are oceanic salmon and many flowering desert plants. The fertility schedule for a semelparous organism would have zeroes for all ages except for the single reproductive age. In iteroparous (polycarpic) reproduction, the individual reproduces repeatedly during its lifetime. Examples include long-lived organisms such as sea turtles and oak trees. Fertility schedules for iteroparous organisms have non-zero entries for two or more ages.

Plant ecologists use two similar terms, **annual** and **perennial**, to refer to plants that complete their life cycle in a single season, and those that live for more than one season. Although there are many exceptions, most annual species are semelparous, and most perennial species are iteroparous. We will postpone our discussion of the evolutionary significance of these reproductive strategies. For now, we will simply use the fixed birth schedule for a population to help us calculate the intrinsic rate of increase.

THE SURVIVORSHIP SCHEDULE $l(x)$

Fertility is only half the story. The population growth rate depends equally on the rates of mortality for different ages. Individuals of a particular age might produce dozens of offspring, but if very few individuals survive to that age, the effect on population growth rate will be minor.

How can we measure the survivorship schedule of a population? Imagine that we have a **cohort** of individuals that were all born at the same time. We follow this cohort from birth until all the individuals have died. We keep track of the number of individuals that have survived to the start of each new year. These data can be represented as a column of numbers, $S(x)$. Table 3.1 gives some cohort data for our hypothetical life table. We begin with a cohort of 500 individuals at birth, and by the beginning of the fifth year, all of them have died.

The raw data in the $S(x)$ column must now be converted to the **survivorship schedule**, designated as $l(x)$, where l stands for life table. The quantity $l(x)$ is defined as the proportion of the original cohort that survives to the start

Table 3.1 Standard life-table calculations.^a

x	$S(x)$	$b(x)$	$l(x) = \frac{S(x)}{S(0)}$	$\frac{g(x)}{l(x+1)l(x)}$	$l(x)b(x)x$	Initial estimate $e^{-rx}(x)b(x)$	Corrected estimate $e^{-rx}l(x)b(x)$	
0	500	0	1.0	0.80	0.0	0.000	0.000	
1	400	2	0.8	0.50	1.6	0.780	0.736	
2	200	3	0.4	0.25	1.2	0.285	0.254	
3	50	1	0.1	0.00	0.1	0.012	0.010	
4	0	0	0.0		0.0	0.000	0.000	
				$R_0 = \frac{\sum l(x)b(x)x}{\sum l(x)b(x)}$	$= 2.9$ offspring	$\Sigma = 4.3$	$\Sigma = 1.077$	$\Sigma = 1.000$

$G = \frac{\sum l(x)b(x)x}{\sum l(x)b(x)}$	$= 1.483$ years
r (estimated) $= \ln(R_0)/G$	$= 0.718$ individuals/ (individual \cdot year)
Correction added to estimated r	$= 0.058$
r (Euler)	$= 0.776$ individuals/ (individual \cdot year)

^a The x , $S(x)$, and $b(x)$ columns are supplied. All others are calculated from these.

of age x . Equivalently, we can think of $l(x)$ in terms of the survivorship of an individual. $l(x)$ is the *probability* that an individual survives from birth to the beginning of age x . To calculate $l(x)$, divide the number of survivors of age x [$S(x)$] by the size of the original cohort [$S(0)$]:

$$l(x) = \frac{S(x)}{S(0)} \quad \text{Equation 3.1}$$

The first entry in the $l(x)$ column is $l(0)$. It represents the survivorship of the cohort to birth. By definition, all individuals in the cohort have "survived" to the start, so the value of $l(0)$ is always 1.0 [$l(0) = S(0)/S(0) = 1.0$]. The last entry in the $l(x)$ column is $l(k)$. It represents the age that none of the original cohort reaches: $l(k)$ always equals 0.0 [$l(k) = 0.0/S(0) = 0.0$]. Between these endpoints, $l(x)$ shrinks in size as individuals in the cohort age and die. Thus, the $l(x)$ column is a set of consecutively decreasing real numbers between 1.0 and 0.0.

For the data in Table 3.1, the original cohort was 500 individuals, so we will divide each observation by this value to calculate $l(x)$. Notice that 80% of the original cohort survived to age 1 [$l(1) = 0.80$], but only 10% of the cohort made it to the start of age 3 [$l(3) = 0.10$]. This remaining 10% died between the start of age 3 and the start of age 4, so $l(4) = 0.0$; none of the original cohort is left.

When you calculate $l(x)$ from a survivorship schedule, take care to divide all the entries by the original cohort size [$S(0)$]. Do not make the common mistake of dividing $S(x)$ by other values in the life table. In the next section, we will calculate age-specific survival probabilities, which do use consecutive values of $S(x)$. But for the calculation of $l(x)$, always divide the observed values by $S(0)$.

SURVIVAL PROBABILITY $g(x)$

The survivorship schedule $l(x)$ gives the probability of survival from birth to age x . To compare the survival of different ages directly, we must determine the probability of survival from age x to age $x + 1$, *given* that an individual has already survived to age x . The probability that an individual of age x survives to age $x + 1$ is:

$$g(x) = \frac{l(x+1)}{l(x)} \quad \text{Equation 3.2}$$

From Table 3.1, for example, the probability that a newborn survives its first year and reaches age 1 is $g(0) = 0.8/1.0 = 0.8$. Thus, there is an 80% chance

that a newborn will still be alive at age 1. If we are thinking in terms of a cohort analysis, 80% of all newborns will be alive at age 1. In contrast, the probability of survival between ages 1 and 2 [$g(1)$] is $(0.4/0.8) = 0.5$. Although the $l(x)$ schedule never increases with age, the $g(x)$ schedule may either increase or decrease. The way in which survival probabilities change with age is an important component of the life history of an organism, as described in the next section.

SURVIVORSHIP SCHEDULES IN NATURE

What are the different types of survivorship curves observed in nature? There are three basic patterns. These can be seen by plotting the logarithm of $l(x)$ on the y axis and age (x) on the x axis. The points on this graph are connected to form a survivorship curve. The slope of this curve at any point is $\ln[g(x)]$. Therefore, if the survivorship curve forms a straight line, the probability of survival is constant over those ages.

Figure 3.2 illustrates the three types of curves. A "Type I" survivorship curve has high survivorship during young and intermediate ages, then a steep drop-off in survivorship as individuals approach the maximum life span. Examples include humans and other mammals that invest a good deal

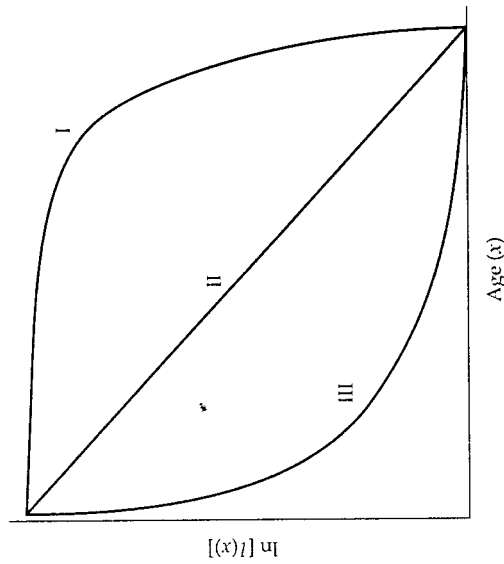


Figure 3.2 Type I, II, and III survivorship curves. Note the logarithmic transformation of the y axis.

of parental care in their offspring, ensuring high survivorship of young age classes.

The opposite, and more common, pattern is a "Type III" survivorship curve. In this case, survivorship is very poor for the young age classes, but much higher for older individuals. Examples include many insects, marine invertebrates, and flowering plants. These organisms may produce hundreds or thousands of eggs, larvae, or seeds, most of which die. However, the handful of individuals that do pass through this vulnerable stage have relatively high survivorship in later years.

Finally, the "Type II" curve is intermediate between these two. Because it is a straight line on a logarithmic graph, the Type II survivorship schedule is one in which the mortality rate is constant throughout life. Few organisms have a true Type II survivorship curve, because it is unusual for the probability of death to remain constant as an organism ages. Some birds have a Type II curve for much of their lives, but often with a steeper mortality curve during the more vulnerable egg and chick stages.

The $l(x)$ and $b(x)$ schedules are the basis for all our life-table calculations. Keep in mind that these schedules are independent pieces of data about death and birth. The $l(x)$ schedule is calculated by following the survivorship of a cohort of organisms. It tells us only the chances of individuals surviving to a particular age, and contains no information about their reproduction. In contrast, the $b(x)$ schedule reveals only the per capita birth rates of females of different ages, and does not say anything about how many females actually survive to those ages. If we know the $l(x)$ and $b(x)$ schedules, we can calculate the intrinsic rate of increase, as illustrated in the next section. When you work with the $l(x)$ and $b(x)$ schedules, be careful with your notation. Remember that the $l(x)$ column gives the survivorship *up to* the start of age x , whereas the $b(x)$ schedule gives the per capita birth rates of females of age x .

CALCULATING NET REPRODUCTIVE RATE (R_0)

To estimate r from the $l(x)$ and $b(x)$ schedules, we first have to compute two other numbers, the net reproductive rate (R_0) and the generation time (G). These numbers are part of the recipe for estimating r , but they tell us important things about an age-structured population in their own right. The net reproductive rate, R_0 , is defined as the mean number of offspring produced per female over her lifetime. To compute R_0 , multiply each value of $l(x)$ by the corresponding value of $b(x)$ and sum these products across all ages:

$$R_0 = \sum_{x=0}^k l(x)b(x) \tag{Equation 3.3}$$

The units of R_0 are numbers of offspring. The net reproductive rate represents the reproductive potential of a female during her entire lifetime, adjusted for the mortality schedule. Suppose that there was no mortality in the population until females reached their maximum age. This would mean that $l(x) = 1.0$ for all ages except the last. In this case, Equation 3.3 would simply add up the lifetime production of offspring—the gross reproductive rate. But in most populations, mortality in each age class reduces the potential contribution of offspring to the next generation. Thus, the net reproductive rate is the offspring production discounted by mortality. For the birth and death schedules in Table 3.1, $R_0 = 2.9$ offspring.

If R_0 is greater than 1.0, there is a net surplus of offspring produced each generation, and the population increases exponentially. If R_0 is less than 1.0, the mortality is so great that the population cannot replace itself, and it declines to extinction. Finally, if $R_0 = 1.0$, the offspring production exactly balances the mortality each generation, and the population size does not change.

This description of R_0 is very similar to the description of λ , the finite rate of increase in the exponential growth model (see Chapter 1). In fact, you might be tempted to conclude that $r = \ln(R_0)$, because $r = \ln(\lambda)$ for populations with no age structure (Equation 1.5). However, λ measures the rate of increase as a function of *absolute time*, whereas R_0 measures increase as a function of *generation time*. Therefore, if we want to calculate r , we must scale R_0 to account for generation time.

CALCULATING GENERATION TIME (G)

Generation time is a somewhat elusive concept for populations with continuous growth. Imagine that we followed a cohort from birth and kept track of all the offspring it produced. One definition of the generation time is the average age of the parents of all the offspring produced by a single cohort (Caughley 1977). This is calculated as:

$$G = \frac{\sum_{x=0}^k l(x)b(x)x}{\sum_{x=0}^k l(x)b(x)} \tag{Equation 3.4}$$

The units of $l(x)$ and $b(x)$ cancel in the numerator and denominator, leaving us with an answer in units of time (x). Note that the numerator will always be larger than the denominator in Equation 3.4. Consequently, the generation time will always be greater than 1.0 for populations with age structure. For the data in Table 3.1, $G = 1.483$ years.

CALCULATING INTRINSIC RATE OF INCREASE (r)

We can use the equation for exponential growth to solve for r in terms of R_0 and G (Mertz 1970). Imagine a population is growing exponentially for a time G :

$$N_G = N_0 e^{rG} \quad \text{Expression 3.2}$$

Dividing both sides by N_0 gives:

$$\frac{N_G}{N_0} = e^{rG} \quad \text{Expression 3.3}$$

The ratio on the left side of the expression is an approximation to the net reproductive rate, R_0 :

$$R_0 \approx e^{rG} \quad \text{Expression 3.4}$$

Taking the natural logarithm of both sides gives:

$$\ln(R_0) \approx rG \quad \text{Expression 3.5}$$

Rearranging Expression 3.5 gives us an approximation for r :

$$r \approx \frac{\ln(R_0)}{G} \quad \text{Equation 3.5}$$

Thus, the rate of population increase is slower for organisms with long generation times. Continuing with the data in Table 3.1, the estimate of r is 0.718 individuals/(individual \cdot year).

Equation 3.5 is only an approximation, although it is usually within 10% of the true value (Stearns 1992). To obtain an exact solution for r , you must solve the following equation:

$$1 = \sum_{x=0}^k e^{-rx} l(x)b(x) \quad \text{Equation 3.6}$$

Equation 3.6 is adapted from the Euler equation (pronounced "oiler"), named after the Swiss mathematician L. Euler (1707–1783), who developed it in his analyses of human demography. Later in this chapter, we will illustrate the derivation of the Euler equation. For now, we will simply use Equation 3.6 as a formula for determining the precise value of r .

Because we know the $l(x)$ and $b(x)$ schedules, the only unknown quantity in Equation 3.6 is r . Unfortunately, there is no way to solve this equation except by plugging in different values of r and adjusting your estimate upwards or downwards. A good starting place is the estimate of r from Equation 3.5. For

the data in Table 3.1, substituting $r = 0.718$ into Equation 3.6 gives a sum of 1.077, whereas the correct value of r will generate a sum of exactly 1.0. This calculation indicates that our original estimate of r was too small. Because we are summing with the negative exponent of r , a larger value of r will generate a smaller sum. If we experiment with different values, we find that an r of 0.776 is a close solution to the Euler equation.

DESCRIBING POPULATION AGE STRUCTURE

Once we have calculated r from the birth and death schedules, we can forecast the total population size by using any of the equations for exponential growth from Chapter 1. But we are also interested in knowing the number of individuals in each age class of the population. This means we will shift our notation from ages to age classes.

We will use $n_i(t)$ to indicate the number of individuals at time t in age class i . For example, if $n_1(3) = 50$, there are 50 individuals in the first age class at the third time step. Because there are k age classes in the population, the age structure at time t consists of a vector of abundances. We indicate this vector with a boldfaced, lowercase \mathbf{n} :

$$\mathbf{n}(t) = \begin{pmatrix} n_1(t) \\ n_2(t) \\ \vdots \\ n_k(t) \end{pmatrix} \quad \text{Expression 3.6}$$

For example, the vector for the population in Table 3.1 after five years might be:

$$\mathbf{n}(5) = \begin{pmatrix} 600 \\ 270 \\ 100 \\ 50 \end{pmatrix} \quad \text{Expression 3.7}$$

Thus, there are 600 individuals in the first age class, but only 50 individuals in the terminal age class (age class 4). Using information in the mortality and fertility schedules, we can predict how the age structure of a population changes from one time period $[\mathbf{n}(t)]$ to the next $[\mathbf{n}(t+1)]$.

Describing the population in terms of its age structure requires us to shift from using ages to using age classes. First, we need to obtain survivorship probabilities P_i for each age class. These probabilities represent the chance that an individual in age class i survives to age class $i+1$. Next, we need to calculate fertilities F_i for each age class. These fertilities represent the average number of offspring produced by an individual in age class i . Clearly, the sur-

vivorship probabilities and fertilities for individuals of different age classes are related to the $l(x)$ and $b(x)$ schedules for individuals of different ages.

However, the conversion of these values is tricky; it depends on the timing of births and deaths within an age class, and the timing of the population census (Caswell 1989). In this primer, we will assume a simple **birth-pulse model**, in which individuals give birth to all their offspring on the day they enter a new age class. We will further assume a **postbreeding census**, in which individuals are counted each year just after they breed.

These assumptions make the calculation of P_i and F_i relatively simple. A more realistic **birth-flow model**, in which individuals reproduce continuously in an age class, would require more complex calculations. Keep in mind that the estimates of population growth will depend on how the age-class model is set up. The estimates of population growth also may not match the exact calculations from the Euler equation. Once we have the survivorship and fertility values for each age class, we will use them to calculate the changes in population structure with time.

CALCULATING SURVIVAL PROBABILITIES FOR AGE CLASSES (P_i)

For the birth-pulse model with a postbreeding census, the probability that an individual in age class i survives to age class $i + 1$ is:

$$P_i = \frac{l(i)}{l(i-1)} \quad \text{Equation 3.7}$$

This equation is similar to the calculation of the age-specific survival probability $g(x)$ (Equation 3.2), although note the shift in notation as we go to a model of age classes. With Equation 3.7, it is easy to calculate the change in the number of individuals in a particular age class from one time period to the next:

$$n_{i+1}(t+1) = P_i n_i(t) \quad \text{Equation 3.8}$$

Equation 3.8 says that the number of individuals in a particular age class next time step $[n_{i+1}(t+1)]$ is the number of individuals currently in the *previous* age class $[n_i(t)]$ multiplied by the survival probability for that age class (P_i). So, the survival probability controls the rate at which individuals "graduate" to each successive age class.

CALCULATING FERTILITIES FOR AGE CLASSES (F_i)

Equation 3.8 works for all age classes except the first. The number of individuals in the first age class depends on the reproduction of all the age classes.

We define the fertility of age class i as:

$$F_i = b(i)P_i \quad \text{Equation 3.9}$$

Equation 3.9 says that the fertility of a particular age class is the number of offspring produced, discounted by the survival probability for that age class. The discount is necessary because the parents must survive through the age class in order to reproduce and have their offspring counted.

Once F_i is known for each age class, we multiply these fertilities by the number of individuals in each age class. This product is then summed over all age classes to calculate the number of new offspring:

$$n_1(t+1) = \sum_{i=1}^k F_i n_i(t) \quad \text{Equation 3.10}$$

Having derived fertility and survivorship coefficients for each age class from the $l(x)$ and $b(x)$ schedules, we can now calculate the number of individuals in each age class for a single time step. For a population with four age classes, we would have:

$$\begin{aligned} n_1(t+1) &= F_1 n_1(t) + F_2 n_2(t) + F_3 n_3(t) + F_4 n_4(t) \\ n_2(t+1) &= P_1 n_1(t) \\ n_3(t+1) &= P_2 n_2(t) \\ n_4(t+1) &= P_3 n_3(t) \end{aligned} \quad \text{Expression 3.8}$$

In the next section we will express these changes in matrix form.

THE LESLIE MATRIX

We can represent the growth of an age-structured population in matrix form. The **Leslie matrix**, named after the population biologist P. H. Leslie, describes the changes in population size due to mortality and reproduction (Leslie 1945). If there are k age classes, the Leslie matrix is a $k \times k$ square matrix. It always has the following form:

$$A = \begin{bmatrix} F_1 & F_2 & F_3 & F_4 \\ P_1 & 0 & 0 & 0 \\ 0 & P_2 & 0 & 0 \\ 0 & 0 & P_3 & 0 \end{bmatrix} \quad \text{Expression 3.9}$$

Each column of the Leslie matrix is the age at time t and each row is the age at time $t + 1$. Each entry in the matrix represents a transition, or change in the number of individuals from one age class to another. In the Leslie matrix, the

fertilities are always in the first row; they represent contributions to newborns from reproduction of each age class. The survival probabilities are always in the subdiagonal. They represent transitions from one age class to the next. All other entries in the Leslie matrix are 0 because no other transitions are possible. Individuals cannot remain in the same age class from one year to the next, so the diagonals must equal zero. Similarly, individuals cannot skip or repeat age classes, so other entries in the matrix are zero.

The reason for using the matrix format is that we can now describe population growth as a simple matrix multiplication:

$$\mathbf{n}(t+1) = \mathbf{A}\mathbf{n}(t) \quad \text{Equation 3.11}$$

In other words, the population vector in the next time step $[\mathbf{n}(t+1)]$ equals the Leslie matrix (\mathbf{A}) multiplied by the current population vector $[\mathbf{n}(t)]$. The rules of matrix algebra are used to calculate the changes in abundance in each age class, and these are equivalent to the calculations in Expression 3.8. If you have had matrix algebra, λ is the dominant eigenvalue of the Leslie matrix. Now that we have converted our age-based life-table data to an age-class Leslie matrix, we are ready to see how age structure changes during population growth.

Table 3.2 Calculation of age-specific survival probabilities and fertilities for the Leslie matrix. Data from Table 3.1.

x	i	$l(x)$	$b(x)$	$P_i = l(i)/l(i-1)$	$F_i = b(i)P_i$
0		1.0	0		
1	1	0.8	2	0.80	1.60
2	2	0.4	3	0.50	1.50
3	3	0.1	1	0.25	0.25
4	4	0	0	0.00	0.00

The resulting Leslie matrix is:

$$\mathbf{A} = \begin{bmatrix} 1.6 & 1.5 & 0.25 & 0 \\ 0.8 & 0 & 0 & 0 \\ 0 & 0.5 & 0 & 0 \\ 0 & 0 & 0.25 & 0 \end{bmatrix}$$

TABLE AND STATIONARY AGE DISTRIBUTIONS

Table 3.2 converts the life-table data of Table 3.1 to a Leslie matrix. We use this Leslie matrix to compare the growth of two hypothetical populations. One population has 50 individuals in each age class, and the second population has 200 newborns, but no other age classes present. Figure 3.3 shows the

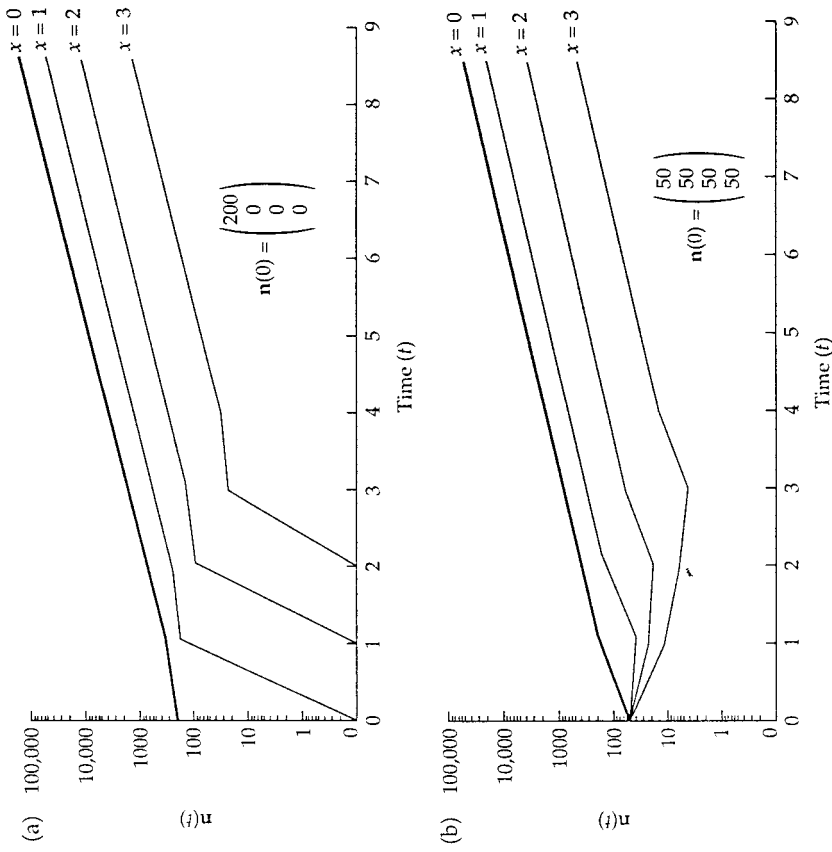


Figure 3.3 Stable age distributions, showing the effects of initial age structure on population growth. Each line represents a different age class, growing according to the birth and death schedules of Table 3.1. In (a), the initial age distribution was 200 newborns. In (b), the initial age distribution was 50 individuals in each age class. After some initial fluctuations, both populations settle into identical stable age distributions. On the logarithmic scale, the straight line for each age class indicates exponential increase.

number of individuals in each age class as a function of time. You can see that the graphs for the two populations initially appear quite different from one another as the relative numbers in the different age classes change in the early phases of population growth. In particular, you can see that the population with 200 newborns is dominated by this single age class, which passes as a cohort through the older age classes. However, after about 6 time steps, both populations have converged on the same age structure—they both have the same relative numbers in each age, with newborns being most common, and the oldest individuals being most rare. These relative proportions are maintained as the numbers in all ages increase exponentially.

These graphs illustrate an important property of age-structured populations. For most life tables, if a population is growing with constant birth and death rates, it will quickly converge on a **stable age distribution**, regardless of its initial age structure. In the stable age distribution, the *relative* numbers of individuals in each age class remain constant. Remember that the *absolute* numbers will increase exponentially, as evidenced by the linear population growth curves on the logarithmic scale of Figure 3.3. A special kind of stable age distribution is the **stationary age distribution**. In a stationary age distribution, $r = 0$, so both the relative and the absolute numbers in each age class remain constant.

What are the relative proportions in the different ages once the stable age distribution has been achieved? The proportion of the population represented by each age is just the number in that age divided by the total population size. This ratio is (Mertz 1970):

$$c(x) = \frac{e^{-rx}l(x)}{\sum_{x=0}^k e^{-rx}l(x)} \quad \text{Equation 3.12}$$

Once r has been calculated from the $l(x)$ and $b(x)$ schedules, Equation 3.12 can be used to determine the stable age distribution. The calculations are illustrated in Table 3.3. In a stable age distribution, newborns are the most common age, and the oldest age is least common. In most cases, the larger r is, the greater the proportion of the total population represented by newborns and young individuals. For the matrix algebra solution, the stable age distribution is the right-hand eigenvector of the Leslie matrix.

The Leslie matrix calculations of population growth can also be used as an independent check on the calculation of r . Table 3.4 illustrates some of the raw data of age structure and population size from Figure 3.3a. For any two consecutive time steps in the model, the ratio of the current population size to the previous population size is a measure of λ , the finite rate of increase. The final column of Table 3.4 gives the natural logarithm of this ratio, which is r .

Table 3.3 Calculation of stable age and reproductive value distributions.^a

x	Stable age distribution			Reproductive value distribution			
	$l(x)$	$b(x)$	$l(x)e^{-rx}$	$c(x)$	$e^{-rx}l(x)$	$e^{-rx}l(x)b(x)$	$v(x)$
0	1.0	0	1.000	0.684	1.000	1.000	1.000
1	0.8	2	0.368	0.252	2.716	0.736	2.716
2	0.4	3	0.085	0.058	11.802	0.254	3.116
3	0.1	1	0.010	0.007	102.574	0.010	1.026
			$\Sigma = 1.463$				

^aThese calculations use $r = 0.776$, from the solution to the Euler equation in Table 3.1.

By 6 or 7 time steps in the model, the stable age distribution has been achieved, and the estimate of r is 0.776, which matches the calculation from the Euler equation in Table 3.1.

Table 3.4 Estimating r from the Leslie matrix calculations.^a

Time step (t)	$n_1(t)$	$n_2(t)$	$n_3(t)$	$n_4(t)$	$n_5(t)$	$n_6(t)$	$n_{\text{total}}(t)$	$\lambda = \frac{n_{\text{total}}(t)}{n_{\text{total}}(t-1)}$	$r = \ln(\lambda)$
0	200	0	0	0	0	0	200		
1	320	160	0	0	0	0	480	2.4	0.875
2	752	256	80	0	0	0	1088	2.267	0.818
6	16,549	6091	1402	161	0	0	24,203	2.173	0.776
7	35,965	13,239	3045	351	0	0	52,600	2.173	0.776
8	78,165	28,772	6620	761	0	0	114,318	2.173	0.776

^aThe data are from different time steps in Figure 3.3a. Fractions for the age-class values have been rounded to the nearest whole number.

Model Assumptions

In spite of the lengthy calculations, the model presented here shares the basic assumptions of the simple exponential growth model we derived in Chapter 1. In other words, we assume a closed population, no genetic structure, and no time lags. In the simple exponential model, we assumed that b and d were constant—they did not vary with time or with population density. In the age-structured model, we assume that the $l(x)$ and the $b(x)$ schedules are constant. As before, if each age class has a constant birth and death rate no matter how large the population, resources must be unlimited.

Incidentally, if we use the value of r from the Euler equation to forecast population growth, we must further assume that the population has achieved a stable age distribution. One final point is that we have described the $l(x)$ schedule from a cohort analysis, in which the fate of a cohort is followed through time. This **horizontal**, or **cohort life table** is the simplest method of obtaining the $l(x)$ schedule, but it assumes that death rates are constant during the time the cohort is followed. A more reliable method is to measure short-term death rates directly for each age class. Finally, it is possible to take a cross-section of the population at one time and estimate death rates from the relative sizes of consecutive age classes. This **vertical**, or **static life table** is much less reliable and assumes the population has reached a stable age distribution. However, birth and death rates can be very difficult to measure in the field, and we often have to rely on a number of methods to piece together the data needed for a life-table analysis.

Model Variations

DERIVATION OF THE EULER EQUATION

The Euler equation forms the basis for age-structured demography, so it is important to understand how this equation is derived. The key to the Euler equation is recognizing the relationship between the number of births now and the number of births at some point in the past (Roughgarden 1979). The number of births in the population now, $B(t)$, is simply the sum of the number of births from parents of all different ages:

$$B(t) = \sum_{x=0}^k (\text{births from parents of age } x) \quad \text{Expression 3.10}$$

If we allow the age intervals to become infinitely small, we can express this as an integral equation:

$$B(t) = \int_0^k (\text{births from parents of age } x) dx \quad \text{Expression 3.11}$$

The number of births from parents of age x is the product of the number of

individuals born at time $t - x$, their offspring production $[b(x)]$ and their probability of surviving to age x $[l(x)]$:

$$B(t) = \int_0^k B(t-x)l(x)b(x)dx \quad \text{Expression 3.12}$$

Remember that the number of births comes from a population that is increasing exponentially. Using C as an arbitrary starting population size, we have:

$$B(t) = Ce^{rt} \quad \text{Expression 3.13}$$

Substituting this back into Expression 3.12 yields:

$$Ce^{rt} = \int_0^k Ce^{r(t-x)}l(x)b(x)dx \quad \text{Expression 3.14}$$

Finally, if we divide both sides of Expression 3.14 by Ce^{rt} , we have the Euler equation:

$$1 = \int_0^k e^{-rx}l(x)b(x)dx \quad \text{Equation 3.13}$$

As we noted earlier, the equivalent equation in discrete time is:

$$1 = \sum_{x=0}^k e^{-rx}l(x)b(x) \quad \text{Equation 3.14}$$

REPRODUCTIVE VALUE

Using the Euler equation, we can calculate another useful statistic from the life table—the **reproductive value** of each age (Fisher 1930). The reproductive value is the relative number of offspring that remain to be born to individuals of a given age. You might think that a newborn individual would have the highest reproductive value because it has not yet produced any offspring. However, its reproductive value is discounted by the fact that it might not achieve its maximum potential lifespan and produce all of its potential offspring. Let $v(x)$ equal the reproductive value for an individual of age x . We can define reproductive value as the following ratio in a stable age distribution (Wilson and Bossert 1971):

$$v(x) = \frac{\text{number of offspring produced by individuals of age } x \text{ or older}}{\text{number of individuals of age } x} \quad \text{Expression 3.15}$$

We can use the Euler equation to quantify the terms in the numerator and the

denominator. For the numerator, we add the terms in the Euler equation from the current age forward:

$$\text{Offspring production} = \int_x^k e^{-ry} l(y) b(y) dy \quad \text{Expression 3.16}$$

For the denominator, the number of individuals in age x is the number born at time x in the past, multiplied by the probability of surviving to age x . Thus:

$$\text{Number in age } x = e^{-rx} l(x) \quad \text{Expression 3.17}$$

Substituting Expressions 3.16 and 3.17 into 3.15 gives:

$$v(x) = \frac{\int_x^k e^{-ry} l(y) b(y) dy}{e^{-rx} l(x)} \quad \text{Expression 3.18}$$

Rearranging the right-hand side yields a formula for reproductive value:

$$v(x) = \frac{e^{rx}}{l(x)} \int_x^k e^{-ry} l(y) b(y) dy \quad \text{Equation 3.15}$$

The discrete-time version of Equation 3.15 allows us to use the $l(x)$ and $b(x)$ schedules to calculate the reproductive value for individuals of age x :

$$v(x) = \frac{e^{rx}}{l(x)} \sum_{y=x}^k e^{-ry} l(y) b(y) \quad \text{Equation 3.16}$$

For the matrix algebra solution, the left-hand eigenvector of the Leslie matrix is the vector of reproductive values. From Equation 3.15, the reproductive value of newborns always equals 1.0. Thus, reproductive value is measured relative to that of the first age. For example, if $v(3) = 2.0$, an individual of age 3 will produce roughly twice as many offspring during the remainder of its lifetime as will a newborn. Reproductive value reflects the survivorship of an individual to its current age, its survivorship and reproduction in future ages, and the magnitude of r . Reproductive value usually peaks at or near the age of first reproduction, then drops off rapidly with later ages. For the data in Table 3.1, reproductive value is maximal for individuals of age 2 (Table 3.3).

Reproductive value tells us which ages in the population are most "valuable" for future population growth. In Chapter 2, we noted that maximum yield for a harvested population occurred when the population was harvested to maximize population growth rate. For the simple logistic model, the best

strategy turned out to be maintaining the population at $K/2$. For an age-structured population, maximizing population growth rate would mean harvesting individuals with relatively low reproductive value—usually newborns and very old individuals, depending on the age structure of the population.

Reproductive value is also relevant to problems of population management and conservation biology. If we are going to transplant captive-bred individuals to a new population in order to increase the population growth rate, we should wait until those individuals reach the age with the highest reproductive value. Finally, natural selection will operate most heavily on ages with high reproductive value. For example, an allele that expresses deleterious effects in reproductive age classes will be eliminated by selection much more quickly than an allele that expresses the effects in older age classes, with lower reproductive value. **Senescence** may represent the accumulation of deleterious effects in old individuals. Selection pressure is weaker on older individuals (Rose 1984), in part because of their lower reproductive value (Fisher 1930).

LIFE HISTORY STRATEGIES

Life-table data are essential for ecological predictions of population growth rates and age structure. From an evolutionary perspective, we can ask why we see certain life history patterns. In other words, why has natural selection favored certain $l(x)$ and $b(x)$ schedules? Selection will favor any life history schedule that maximizes an individual's contribution of offspring to the next generation. Thus, the "perfect" life history schedule would be one with maximum survivorship and maximum fertility in all age classes!

However, two forces prevent the evolution of this optimal life history. First, we expect a number of **tradeoffs** to occur among life history traits. Organisms that invest heavily in reproduction have less energy to devote towards growth, maintenance, and resource acquisition. This may lead to tradeoffs between reproduction and survivorship. An organism may produce many small offspring that survive poorly or a few large offspring that survive well. Hence, there may be tradeoffs between offspring number and offspring survivorship.

Life history strategies will also be shaped by **constraints**—physiological or evolutionary limitations that prevent the evolution of certain life history traits. For example, organisms with large body size must take longer to grow and reach maturity, so the age at first reproduction may be constrained by body size. If an organism bears live offspring, body size will also constrain the number of offspring produced. The life history traits of an organism may reflect a long evolutionary heritage, and may not represent the best "solution" to the problem of maximizing fitness in the organism's current environment.

One popular body of theory envisions that relative population density serves as an important selective force on life history traits (MacArthur and Wilson 1967; Pianka 1970). The theory of r - K selection takes its name from the two constants of the logistic growth equation. Imagine a population that is maintained at low population density, so that resources for growth are not limited. Under these circumstances, the best reproductive strategy is simply to maximize offspring production. So, the traits expected under r -selection are early, semelparous reproduction, large r , many offspring with poor survivorship, a Type III survivorship curve, and small adult body size.

By contrast, in K -selection, an organism is growing in an environment that is chronically crowded. An r -strategy will not work in this case because the offspring will face limited resources and be relatively poor competitors. Instead, the best strategy is one that leads to fewer, high-quality offspring that are superior competitors. With resource limitation, K -selection should favor late, iteroparous reproduction, small r , few offspring with good survivorship, a Type I survivorship curve, and large body size. Classic examples of species thought to have evolved under the different regimes include mosquitoes and weeds (r -selected), and humans and whales (K -selected).

In spite of its popularity in textbooks, the theory of r - and K -selection is beset by a number of problems. One fundamental problem is that the "predictions" of r - K selection theory were never derived from a population model with age structure. Another difficulty is that population density is not the only force driving the evolution of life history traits. For example, the theory predicts that iteroparity evolves when organisms face resource competition and must devote more of their energy to growth and maintenance than to reproduction. But iteroparity could also evolve as a "bet-hedging" strategy if the survival of offspring is uncertain from one time period to the next (Murphy 1968). It may be advantageous to spread reproduction over many time periods if there is a risk of losing all your offspring if they are born at the wrong time.

Moreover, not all organisms have life history traits that neatly fit the predictions of the model. For example, many forest trees are long-lived and iteroparous (K -selection), but they have a Type III survivorship curve (r -selection). Finally, the r - K selection theory has not been confirmed experimentally. Laboratory populations of fruit flies (Taylor and Condra 1980) and protozoa (Luckinbill 1979) did not always evolve r -selected traits when they were maintained in uncrowded conditions or K -selected traits when they were maintained in crowded conditions. In spite of its simplistic appeal, the theory of r - and K -selection is of limited value in understanding life history strategies. Although we can recognize the operation of tradeoffs and constraints, we are still a long way from a general theory of life history evolution.

STAGE- AND SIZE-STRUCTURED POPULATION GROWTH

An implicit assumption in our development of the life table model is that the age of an organism is the "correct" variable to use in defining the life history. But for many life histories, age is not the critical variable. For example, many insects pass through egg, larval, pupal, and adult stages. Survival may be influenced more by an insect's stage than its age. That is, survival of a beetle may not depend on whether the beetle is three or six months old, but on whether it is in the larval or adult stage. Of course, age and stage are not independent of one another, because an organism's life history stage will depend, in part, on how old it is. But the transitions between stages are often flexible and depend on biotic factors, such as food supply and population density, and abiotic factors, such as temperature and photoperiod.

Even for organisms that do not have distinct life history stages, survival and reproduction may depend more on the size or an organism than on its age. Many organisms have indeterminate growth—a small fish may be either a fast-growing juvenile or a stunted adult. If the risk of mortality is from predation by other fishes, only the individual's size, rather than its age, may be relevant. Finally, "modular" organisms such as plants and corals may be organized as colonies or semi-independent units (plant shoots) that are capable of reproduction. In these cases, the life history may be extremely complex, as coral colonies can fragment or fuse, and plants can reproduce through vegetative propagation. In all these examples, the age of the organism is less important than its size or stage in determining its survivorship and reproduction.

Fortunately, the Leslie matrix can be modified to account for these kinds of life histories (Lefkovich 1965). The key change is that the entries in the population matrix no longer represent the age of an organism, but rather its stage (or size). We still incorporate a time step that represents the transition from one stage to the next. For example, here is a transition matrix for a simplified insect life cycle with three stages—egg, larva, and adult:

$$\begin{array}{c} \text{egg} \quad \text{larva} \quad \text{adult} \\ \begin{bmatrix} 0 & 0 & F_{ae} \\ P_{ll} & P_{ll} & 0 \\ 0 & P_{la} & P_{aa} \end{bmatrix} \end{array} \quad \text{Expression 3.19}$$

Remember that each column represents the stage at time t and each row represents the stage at time $t + 1$. The entries in the first row represent fertilities. The entries in the other rows represent transition probabilities between stages. In contrast to the Leslie matrix, we now have positive entries on the

diagonal. This means that larvae and adults can stay in a particular stage at a given time, whereas eggs will either die or advance to the larval stage. Only the adult can reproduce, so there is a single fertility entry (F_{ac}) for this stage. Here is a transition matrix for a long-lived forest tree that is classified into five size classes:

$$\begin{matrix}
 & \text{size 1} & \text{size 2} & \text{size 3} & \text{size 4} & \text{size 5} \\
 \text{size 1} & P_{11} & F_{21} & F_{31} & F_{41} & F_{51} \\
 \text{size 2} & P_{12} & P_{22} & 0 & 0 & 0 \\
 \text{size 3} & 0 & P_{23} & P_{33} & 0 & 0 \\
 \text{size 4} & 0 & 0 & P_{34} & P_{44} & 0 \\
 \text{size 5} & 0 & 0 & 0 & P_{45} & P_{55}
 \end{matrix}$$

Expression 3.20

Again, there is the possibility that an individual will remain in the same size class (diagonal elements) or grow to the next consecutive size class (subdiagonal elements). All size classes except the first reproduce, giving positive fertility values in the first row of the matrix.

As a final, and more complex, example, consider a population of reef-building corals with three size classes (small, medium, and large):

$$\begin{matrix}
 & \text{small} & \text{medium} & \text{large} \\
 \text{small} & P_{ss} + F_{ss} & P_{ms} + F_{ms} & P_{ls} + F_{ls} \\
 \text{medium} & P_{sm} & P_{mm} & P_{lm} \\
 \text{large} & P_{sl} & P_{ml} & P_{ll}
 \end{matrix}$$

Expression 3.21

As before, the diagonal elements represent the probability that a colony remains in the same size class, and the subdiagonal elements represent the probability that a colony grows to the next size class. However, there is now the possibility that large colonies can fragment into medium (P_{lm}) or small (P_{ls}) colonies, and that medium colonies can fragment into small colonies (P_{ms}). Small colonies can also fuse with one another, thus "skipping" a stage and going directly from small to large (P_{sl}). Finally, look at the first row of the matrix and notice that the entries are sums of fecundities and stage transitions. This relationship occurs because the production of small colonies has components of sexual reproduction (F) and asexual fragmentation and persistence (P).

As illustrated in Figure 3.4, these complex life cycles can also be represented in loop diagrams. Each circle in the loop represents a different life history stage, and each arrow represents a transition from one stage to the next. Stages not connected by arrows have a zero for the corresponding entry in the transition matrix.

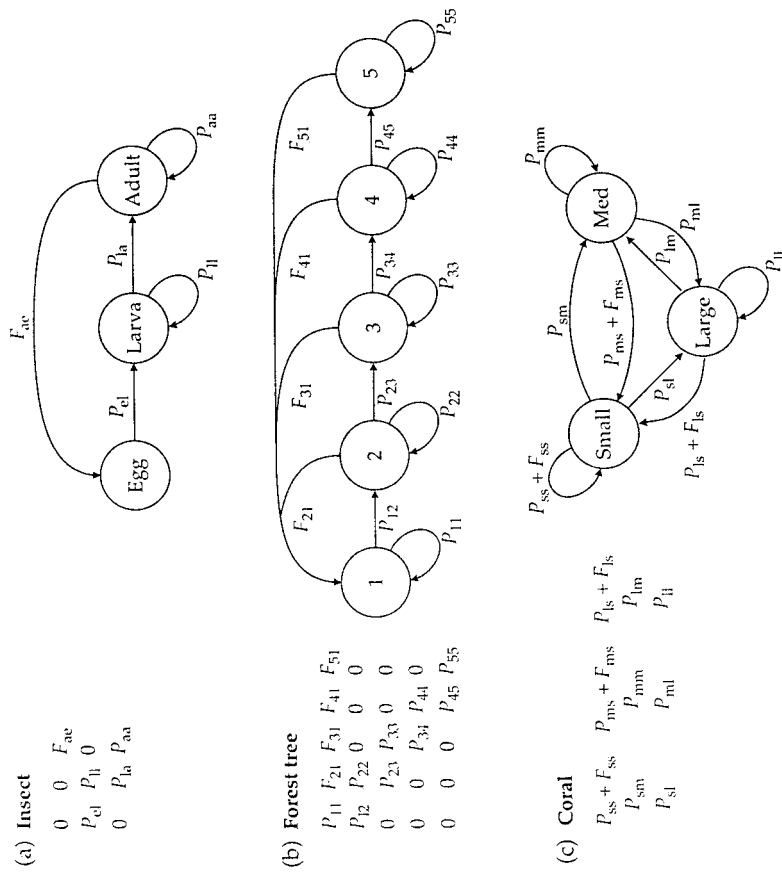


Figure 3.4 Stage-transition matrices and loop diagrams for different life histories. (a) Simplified insect life history. (b) Long-lived forest tree life history. (c) Coral life history, with sexual and asexual reproduction.

In spite of the complexities of these life cycles, the mechanics of the matrix multiplication are exactly the same as for the simple Leslie matrix. As long as the transition elements are constant, the population will eventually exhibit exponential growth and a stable stage distribution. However, we can no longer use the Euler equation for these life histories, and must obtain the matrix solutions for r and the stable stage distribution. For any transition matrix, λ is the dominant eigenvalue. The right-hand eigenvector is the stable stage distribution, and the left-hand eigenvector is the reproductive value distribution (Caswell 1989). The matrix approach allows us to use the same analytical framework to study complex life histories that do not fit a simple age classification.

Empirical Examples

LIFE TABLES FOR GROUND SQUIRRELS

A long-term demographic study of the Uinta ground squirrel (*Spermophilus armatus*) demonstrates the importance of life-table analysis in understanding population growth (Slade and Balph 1974). At a field station in northern Utah, squirrels emerged from hibernation each year between late March and mid-April, depending on the weather. Females bred shortly after they emerged and established territories. The first young were born in early May, and juveniles left their natal burrows about three weeks later. During June and July, all age classes and sexes in the population were active. Adults began hibernating in July, and by September all squirrels had disappeared underground.

Researchers trapped and tagged all individuals in the 8.9-hectare study area and monitored their activity from observation towers. The research was conducted over a seven-year period and divided into two phases. During the first phase (1964–1968), the population was left undisturbed, except for the monitoring. Population size fluctuated from 178 to 255, with a mean of 205.

Table 3.5 Life tables for Uinta ground squirrels (*Spermophilus armatus*) before and after density reduction.

x (years)	Pre-reduction life table		Post-reduction life table	
	$l(x)$	$b(x)$	$l(x)$	$b(x)$
0.00	1.000	0.00	1.000	0.00
0.25	0.662	0.00	0.783	0.00
0.75	0.332	1.29	0.398	1.71
1.25	0.251	0.00	0.288	0.00
1.75	0.142	2.08	0.211	2.24
2.25	0.104	0.00	0.167	0.00
2.75	0.061	2.08	0.115	2.24
3.75	0.026	2.08	0.060	2.24
4.75	0.011	2.08	0.034	2.24
5.75	0.000	0.00	0.019	2.24
6.75	—	—	0.010	2.24
7.75	—	—	0.000	0.00

Data from Slade and Balph (1974).

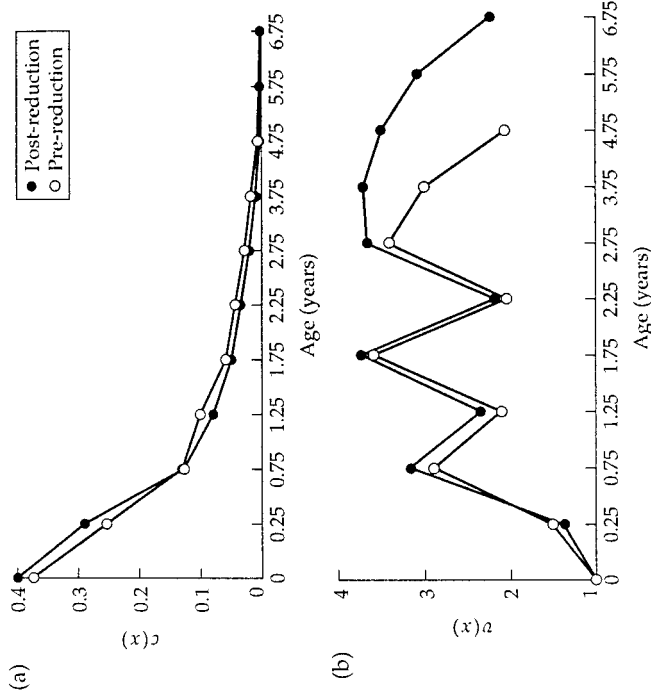


Figure 3.5 (a) Stable age distribution and (b) reproductive value distribution for Uinta ground squirrels (*Spermophilus armatus*) before and after density reduction. Data from Table 3.5.

During the second phase (1968–1971), researchers reduced the squirrel population to about 100 individuals. Life-table analysis (Table 3.5) revealed the dramatic effects of density reduction on growth rate and age structure.

Before the population was reduced, age-specific birth and death rates were approximately balanced, generating a slightly negative growth rate [$r = -0.046$ individuals/(individual \cdot year)]. The maximum life span was approximately five years, although this varied somewhat between different habitats. In the stable age distribution, 37% of the population was juveniles (Figure 3.5a), and reproductive value peaked for individuals near the end of their second year (Figure 3.5b).

After density was reduced, reproduction exceeded mortality, and there was a substantial rate of population increase [$r = 0.306$ individuals/(individual \cdot year)]. The maximum life span increased to seven years, and the stable age distribution shifted slightly towards older ages (Figure 3.5a). The reproductive value showed a broader peak for three- and four-year olds (Figure 3.5b), reflecting the increased reproduction and survival of older ages.

The density reductions revealed that crowding had many effects beyond a slowing of population growth rate. Survivorship, reproduction, life span, and age structure were all sensitive to population density. The manipulations also point to a key weakness of our exponential growth model: age-specific birth and death rates change with population size!

Density dependence can be incorporated into either the mortality or the fecundity schedules for one or more age classes. Even if it limits the increase of only a single age class, density dependence can be an effective brake on total population growth, and can lead to complex population dynamics. In the remainder of this primer, we will return to simple models of populations that do not incorporate age structure. However, the biological details of migration (Chapter 4), competition (Chapter 5), predation (Chapter 6), and colonization (Chapter 7) almost certainly reflect the age and size structure within a population.

STAGE PROJECTION MATRICES FOR TEASEL

Teasel (*Dipsacus sylvestris*) is a European perennial "weed" that is common in abandoned fields and meadows of the eastern United States. The plant has a complex life cycle that can be described with a stage-based matrix model. Most seeds fall within two meters of the adult plant, and the seeds may lie dormant for one or two years. Seeds that successfully germinate form a large-leaved rosette. The rosette phase is variable and may last for more than five years. The rosette requires cold-hardening (vernalization) before it will form a flowering stalk the following summer. Teasel flowers and sets seed only once, and then the plant dies.

Teasel was studied in eight abandoned fields in Michigan, which were sown with teasel seed at the start of the study (Werner 1977; Werner and Caswell 1977). To construct the stage-based transition matrix, individual plants were monitored in marked plots for several consecutive years. The life cycle of teasel can be divided into six stages (Caswell 1989):

1. Dormant first-year seeds
2. Dormant second-year seeds
3. Small rosettes (<2.5 cm diameter)
4. Medium rosettes (2.5–18.9 cm diameter)
5. Large rosettes (≥ 19.0 cm diameter)
6. Flowering plants

Figure 3.6 gives the loop diagram and corresponding stage matrix for this life cycle as measured on one of the eight experimental plots. From the positive entries on the diagonals and subdiagonals, the rosettes can remain in their own size class, grow to a larger rosette size, or flower. The single entry in the

	Seed (1)	Seed (2)	Ros (s)	Ros (m)	Ros (l)	Flowering plant
Seed (1)	0	0	0	0	0	322.380
Seed (2)	0.966	0	0	0	0	0
Ros (s)	0.013	0.010	0.125	0	0	3.448
Ros (m)	0.007	0	0.125	0.238	0	30.170
Ros (l)	0.008	0	0	0.245	0.167	0.862
Flowering plant	0	0	0	0.023	0.750	0

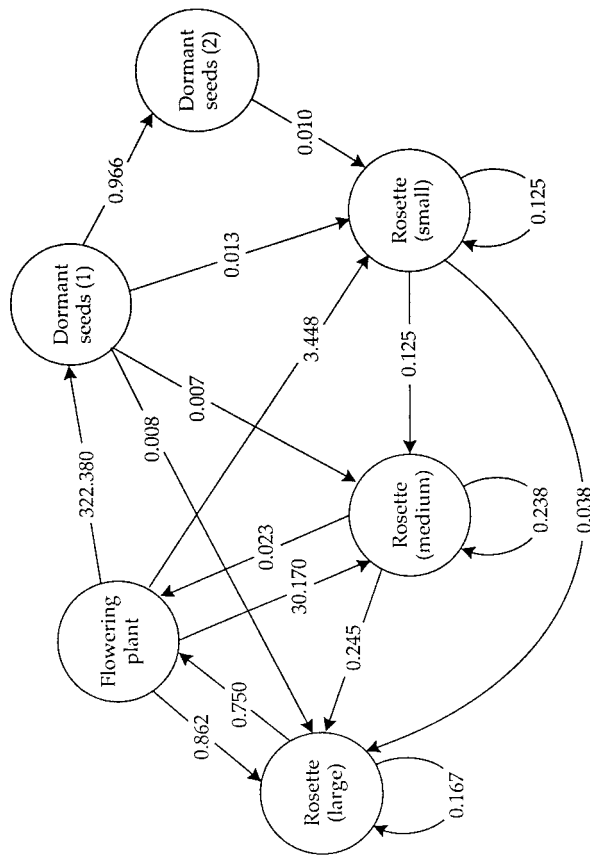


Figure 3.6 Transition matrix and loop diagram for teasel (*Dipsacus sylvestris*). Transitions are shown for dormant first-year and second-year seeds [seed (1) and seed (2)], small, medium, and large rosettes [ros (s), ros (m), ros (l)], and flowering plants. (Data from Caswell 1989.)

first row of the matrix reflects the fact that only the flowering plants can produce seed. Also, notice that the diagonal element for flowering plants (P_{66}) is zero, indicating that they do not survive after they flower. The population growth rate for this matrix is $\lambda = 1.3242$. This corresponds to an r of 0.8434 individuals / (individual \cdot year), with a projected doubling time of less than 10 months.

In contrast to a simple age-classified model, the stable stage distribution does not always decrease with later stages. In the stable stage distribution for teasel, there were more medium than small rosettes (Figure 3.7a). Reproduc-

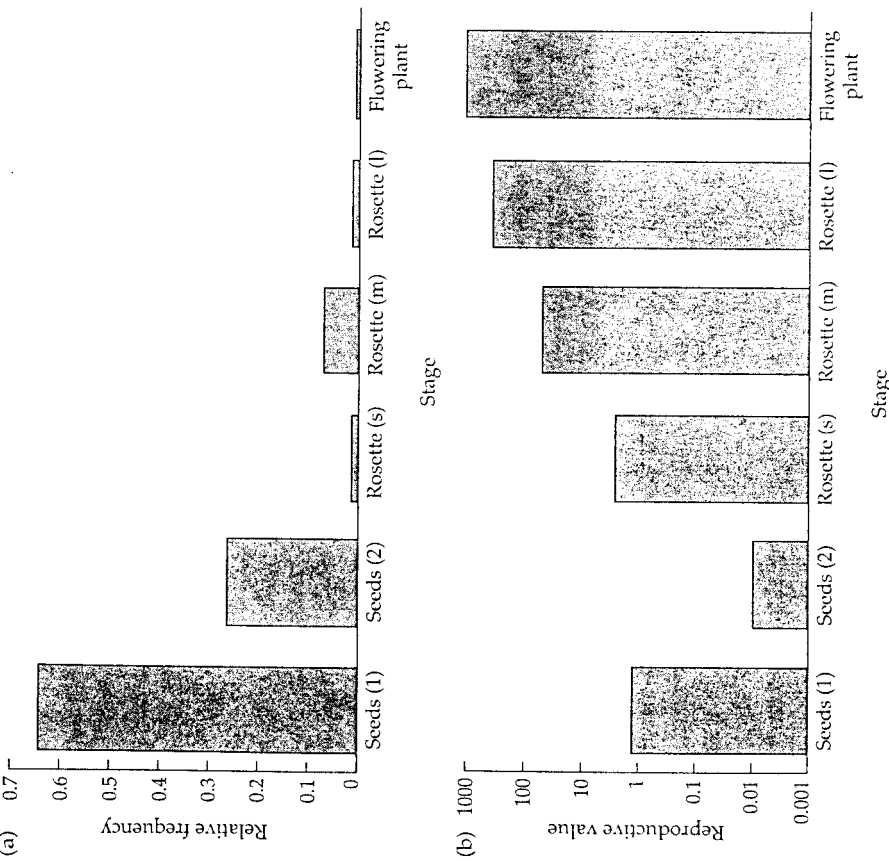


Figure 3.7 (a) Stable-stage distribution and (b) reproductive-value distribution for teasel (*Dipsacus sylvestris*). Note logarithmic scale. Derived from data in Figure 3.6.

tive values varied over six orders of magnitude, from a minimum for second-year dormant seeds to a maximum for flowering plants (Figure 3.7b).

These same data were also analyzed as an age-classified model, treating rosettes of 1–4 years as separate age classes (Werner and Caswell 1977). However, the stage-based model predicted the year of first flowering more accurately than did the age-based model. The results suggested that the size of a rosette, rather than its age, is the more important determinant of growth and survivorship for teasel.

The model results for teasel varied greatly between different fields, and population growth rates (r) ranged from -0.46 to 0.96 individuals/(individual \cdot year). Fields with the lowest r had high levels of grass litter, which suppressed teasel seed germination. Population growth rate was also low in fields with high densities of herbaceous plants, which reduced survivorship of teasel rosettes through competition and shading. Finally, r was correlated with annual primary productivity of a field. Population growth rates were highest in the least productive fields, perhaps reflecting competition with other plants. The very high rates of increase measured for some teasel populations are unlikely to be sustained in the long run. As in the ground squirrel example, a density-dependent model may be more appropriate for forecasting population size.

Problems

- Plot the logarithm (base 10) of squirrel survivorship for the pre- and post-reduction populations (Table 3.5). What is the general shape of these curves (Type I, II, or III), and how does density reduction affect survivorship?
- Here is a set of hypothetical life-table data for a population of snails:

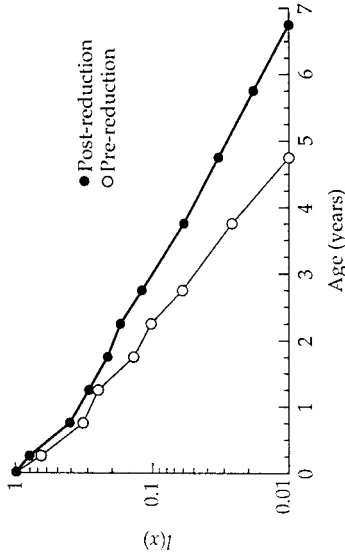
Age in years (x)	S(x)	b(x)
0	500	0.0
1	400	2.5
2	40	3.0
3	0	0.0

- Complete the life-table analysis by calculating $l(x)$, $g(x)$, R_0 , G , and the estimate of r . Calculate the exact value of r with the Euler equation.
 - Determine the stable age and reproductive value distributions for this life table.
- *3. Suppose the snail population in Problem 3.2 consisted of 50 newborns, 100 one-year-olds, and 20 two-year-olds. Construct the Leslie matrix for this life table, and project population growth for the next two consecutive years.

* Advanced problem

Solutions

- Here are the $l(x)$ schedules from Table 3.5, with the y axis plotted on a base 10 logarithmic scale:



The curves are roughly linear, suggesting a Type II survivorship. Reducing density enhanced the survivorship of all ages.

2a. Life-table data are calculated as follows:

x	S(x)	b(x)	$l(x) = \frac{S(x)}{S(0)}$	$g(x) = \frac{l(x+1)}{l(x)}$	$l(x)b(x)$	$l(x)b(x)x$	Initial estimate $e^{-r}l(x)b(x)$	Corrected estimate $e^{-r}l(x)b(x)$
0	500	0.0	1.00	0.8	0.00	0.00	0.000	0.000
1	400	2.5	0.80	0.1	2.00	2.00	0.965	0.946
2	40	3.0	0.08	0.0	0.24	0.48	0.056	0.054
3	0	0.0	0.00		0.00	0.00	0.000	0.000
				$R_0 = \frac{\sum l(x)b(x)}{\sum l(x)b(x)}$	$= 2.24$	$\Sigma = 2.48$	$\Sigma = 1.021$	$\Sigma = 1.000$
				offspring				

$G = \frac{\sum l(x)b(x)x}{\sum l(x)b(x)}$	$= 1.107$ years
r (estimated) $= \ln(R_0)/G$	$= 0.779$ individuals / (individual · year)
Correction added to estimated r	$= 0.020$
r (Euler)	$= 0.749$ individuals / (individual · year)

2b. Stable age and reproductive value distributions are calculated as follows, using $r = 0.749$:

Stable age distribution			Reproductive value distribution					
x	$l(x)$	$b(x)$	$l(x)e^{-rx}$	$c(x)$	$e^{-rx}l(x)$	$e^{-rx}l(y)b(y)$	$\sum e^{-ry}l(y)b(y)$	$v(x)$
0	1.00	0.0	1.000	0.716	1.000	0.000	1.000	1.000
1	0.80	2.5	0.378	0.271	2.644	0.946	1.000	2.644
2	0.08	3.0	0.018	0.013	55.909	0.054	0.054	3.019
			$\Sigma = 1.396$					

3. Age-specific survival probabilities and fertilities are calculated as follows:

x	i	$l(x)$	$b(x)$	$P_i = l(i)/l(i-1)$	$F_i = b(i)P_i$
0		1.00	0.0		
1	1	0.80	2.5	0.80	2.00
2	2	0.08	3.0	0.10	0.30
3	3	0.00	0.0	0.00	0.00

The resulting Leslie matrix is:

$$A = \begin{bmatrix} 2.0 & 0.3 & 0 \\ 0.8 & 0 & 0 \\ 0 & 0.1 & 0 \end{bmatrix} \text{ and the initial population vector is: } \mathbf{n}(0) = \begin{bmatrix} 50 \\ 100 \\ 20 \end{bmatrix}$$

Using Equations 3.8 and 3.10, we have:

$$n_1(1) = (2)(50) + (0.30)(100) + (0)(20) = 130$$

$$n_2(1) = (0.8)(50) = 40$$

$$n_3(1) = (0.1)(100) = 10$$

$$\mathbf{n}(1) = \begin{bmatrix} 130 \\ 40 \\ 10 \end{bmatrix}$$

Repeating the calculation using this new vector gives:

$$n_1(2) = (2)(130) + (0.30)(40) + (0)(10) = 272$$

$$n_2(2) = (0.8)(130) = 104$$

$$n_3(2) = (0.1)(40) = 4$$

$$\mathbf{n}(2) = \begin{bmatrix} 272 \\ 104 \\ 4 \end{bmatrix}$$

CHAPTER 4

Metapopulations



Model Presentation and Predictions

In Chapters 1–3, we explored several models of population growth. These models differed in their major assumptions: unlimited resources (Chapter 1) versus a finite carrying capacity (Chapter 2), and a homogenous population (Chapter 1) versus an age-structured one (Chapter 3). All of these models described a closed population. In other words, the population changed size because of births and deaths that occurred locally. We explicitly assumed that individuals did not move between populations.

This assumption of a closed population was mathematically convenient, but not biologically realistic. For migratory animals, such as North American songbirds that overwinter in the tropics, or oceanic salmon that spawn in freshwater streams, the seasonal movement of individuals is the dominant cause of population change. Many nonmigratory species also move between populations. In particular, organisms with complex life histories often have seeds or larvae that are adapted for movement to new populations. The ascidians described in Chapter 2 are a good example. The adults are filter-feeding invertebrates that attach permanently to rock walls, but the “tadpole” larvae are free-swimming and drift in the current for several days before settlement and metamorphosis. Consequently, the “births” in a local ascidian population consist of juveniles that originated from many different sites.

The movement of individuals between populations may be density-dependent. In territorial species, such as black-throated blue warblers (*Dendroica caerulescens*), not all individuals are able to establish territories, and those that do not may migrate in search of less crowded populations. Mathematical models that ignore the biology of animal and plant movement may not give an accurate description of population dynamics.

In this chapter, we will develop a class of simple models that takes into account the fact that individuals do move among sites and that such movement is potentially important to the persistence and survival of populations. This chapter explores the concept of a **metapopulation**. The metapopulation can be thought of as a “population of populations” (Levins 1970)—a group of several local populations that are linked by immigration and emigration.

In order to study metapopulations, we need to make two important shifts in our frame of reference. The first shift concerns how we measure populations. In Chapters 1–3, our models predicted the size of a population—the number of individuals at equilibrium. Our metapopulation models will not predict the size of a population, but only its *persistence*. Thus, the range of numbers representing population size will be collapsed to only two possible values: 0 (local extinction) or 1 (local persistence). We will no longer distinguish between large and small populations, or between populations that

cycle, fluctuate, or remain constant. Instead, the only distinction is between populations that persist and those that go extinct.

The second shift concerns the spatial scale at which we study populations. In Chapters 1–3 we emphasized equilibrium solutions for population size, implicitly focusing on local populations that persisted through time. In contrast, the metapopulation perspective is that local populations frequently go extinct, and that the appropriate spatial scale for recognizing an equilibrium is the regional or landscape level, which encompasses many connected sites. At this scale, we will no longer focus on the persistence of any particular population. Instead, we will build a model that describes the fraction of all population sites that are occupied. Thus, we will ignore the fate of individual populations and model the extent to which populations fill the landscape. This large-scale view will allow us to use simple mathematics and avoid the complexities of trying to explicitly model local population size and individual migration. As an analogy, if we were modeling the dynamics of a busy parking lot, we would try to predict how many parking spaces were filled, not which particular spaces were occupied.

METAPOPOPULATIONS AND EXTINCTION RISK

The metapopulation perspective allows us to make a distinction between **local extinction**, in which a single population disappears, and **regional extinction**, in which all populations in the system die out. Even if populations are not connected by migration, the risk of regional extinction is usually much lower than the risk of local extinction.

To explore this concept quantitatively, we can define p_c as the **probability of local extinction**—that is, the probability that the population in an occupied patch goes extinct. This probability is a number that ranges between 0 and 1. If $p_c = 0$, persistence is certain, whereas if $p_c = 1$, extinction is certain. All populations go extinct in the long run, so probabilities of extinction must be measured relative to a particular time scale. For metapopulation dynamics, the appropriate time scale is often years or decades.

Suppose that $p_c = 0.7$, for probabilities measured on a yearly time scale. This means there is a 70% chance (100×0.7) that a population will go extinct during a single year, and a 30% chance that it will persist ($1 - p_c = 0.3$). What are the chances that the population will persist for two years? The probability of persistence for two years is the probability of no extinction in the first year ($1 - p_c$) multiplied by the probability of no extinction in the second year ($1 - p_c$). Thus:

$$P_2 = (1 - p_c)(1 - p_c) = (1 - p_c)^2 \quad \text{Expression 4.1}$$

In general, the probability that a population will persist for n years (P_n) is the

probability of no extinction for n years in a row:

$$P_n = (1 - p_e)^n \quad \text{Equation 4.1}$$

For example, if $p_e = 0.7$, and $n = 5$, $P_n = (1 - 0.7)^5 = 0.00243$. So, if there is a 70% chance that a population goes extinct in one year, the chance of persistence for five years in a row is only 0.2%.

Now suppose that instead of a single population, we have two identical populations, each with a p_e of 0.7. For now, we assume that these populations are independent of one another—the chance of extinction in one patch is not affected by the presence or absence of populations in other patches. For this pair of populations, what is the probability of regional persistence, that is, what are the chances that *at least* one population persists for one year? The probability of regional persistence for one year (P_x) is 1 minus the probability that both patches go extinct during the year:

$$P_2 = 1 - (p_e)(p_e) = 1 - (p_e)^2 \quad \text{Expression 4.2}$$

More generally, the probability of regional persistence in a set of x patches is the probability that all x patches do not go simultaneously extinct:

$$P_x = 1 - (p_e)^x \quad \text{Equation 4.2}$$

Thus, if we had 10 patches, each with $p_e = 0.7$, the probability of regional persistence is $P_{10} = 1 - (0.7)^{10} = 0.97$. In other words, with 10 patches, there is a 97% chance that at least one population will persist, even though it is likely that any particular population will go extinct ($p_e = 0.7$)! Figure 4.1 shows that P_x increases rapidly as more patches are added, although there is an overall decrease as p_e is increased.

Equation 4.2 illustrates an important principle: multiple patches “spread the risk” of extinction. Even if individual populations are doomed to extinction, a set of populations can persist for a surprisingly long time. In the next section, we will build metapopulation models in which these local populations are linked to one another, so that probabilities of local extinction and local colonization depend on patch occupancy.

A MODEL OF METAPOPOPULATION DYNAMICS

Imagine a set of homogeneous patches, each of which can be occupied by a single population. Let f equal the fraction of these population sites that are actually occupied. Thus, f is a number constrained between 0 and 1. If $f = 1$, all sites are occupied by populations, and the landscape is saturated. If $f = 0$, all sites are unoccupied, and the metapopulation is regionally extinct.

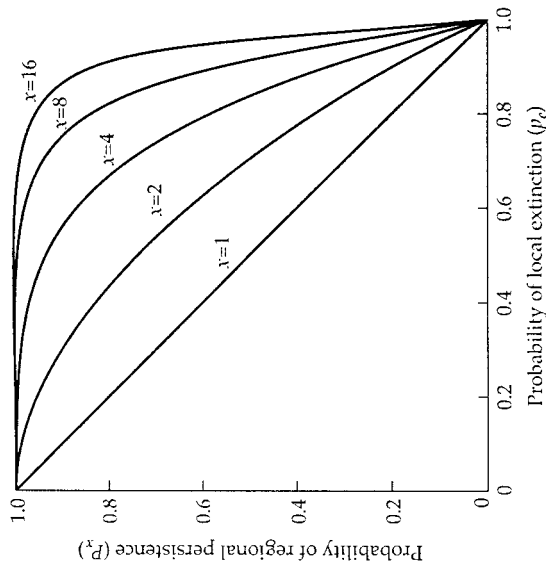


Figure 4.1 The relationship between the probability of regional persistence (P_x), the probability of local extinction (p_e), and the number of populations (x). Note that as the number of populations is increased, the probability of regional persistence is substantially higher, for a given probability of local extinction.

How does f change with time? f can increase if empty sites are successfully colonized. Let $I =$ the **immigration rate**: the proportion of sites successfully colonized per unit time. Technically, we should refer to this as the colonization, not the immigration, rate, but we use this terminology for consistency with the MacArthur–Wilson model, which is developed in Chapter 7. f can also decrease if occupied sites undergo extinction. Let $E =$ the **extinction rate**, that is, the proportion of sites that go extinct per unit time. The change in f is determined by the balance of gains from colonization and losses from extinction:

$$\frac{df}{dt} = I - E \quad \text{Expression 4.3}$$

There is a close analogy between Expression 4.3 and our initial derivation of the exponential growth model in Chapter 1 (Expression 1.5). In the exponential growth model, there was continuous turnover of individuals from births and deaths. Population size (N) reached an equilibrium only if the birth rate precisely equaled the death rate. Similarly, at the metapopulation level, there

is continuous turnover of individual *populations* through colonization and extinction. The fraction of population sites (f) reaches an equilibrium when the immigration rate precisely equals the extinction rate. We will see this same derivation once more in Chapter 7, when we model the number of species in a community.

We now wish to describe the immigration and extinction functions in more detail. The immigration rate depends on two factors. First is the **probability of local colonization**, p_i . If each site is colonized independently, this probability will depend only on the physical and biological conditions within a patch. Many factors can affect p_i , including patch area, the availability of critical habitats and food resources, and the absence or scarcity of predators, pathogens, and competitors.

The probability of local colonization can also be affected by factors that are external to the site. Specifically, if the sites are linked by migration, the probability of colonization may depend on the presence of populations in other sites. In other words, when many sites are occupied (large f), there are many individuals migrating, so the probability of colonization is higher than when few sites are occupied (small f). Therefore, p_i will depend on f . In the following sections, we will develop models in which p_i is either dependent or independent of f .

The immigration rate depends not only on p_i , but also on the availability of unoccupied sites, which is measured by $(1 - f)$. The more sites available for colonization, the faster the overall immigration rate. Thus, the immigration rate is the product of the probability of local colonization (p_i) and the fraction of unoccupied sites $(1 - f)$:

$$I = p_i(1 - f) \quad \text{Expression 4.4}$$

The immigration rate will equal zero in two cases: first, if the probability of local colonization is zero ($p_i = 0$); and second, if all the sites in the metapopulation are occupied ($f = 1$).

If we follow a similar line of reasoning, the extinction rate, E , is the product of the probability of local extinction (p_e) and the fraction of sites occupied (f):

$$E = p_e f \quad \text{Expression 4.5}$$

The extinction rate equals zero if the probability of extinction is zero ($p_e = 0$), or if none of the sites in the metapopulation is occupied ($f = 0$). Substituting Expressions 4.4 and 4.5 back into 4.3 gives us a general metapopulation model:

$$\frac{df}{dt} = p_i(1 - f) - p_e f \quad \text{Equation 4.3}$$

Because this is a continuous differential equation, p_i and p_e are technically not probabilities, but fractional rates. However, p_i and p_e behave as probabilities when they are multiplied by a finite time interval. Over such a time interval, we would need to add a correction term to Equation 4.3 to account for the chance that an occupied patch could undergo an extinction and a recolonization (Ray et al. 1991). However, the correction term is small, and it is simpler to use the continuous differential equation and to interpret p_i and p_e as immigration and extinction probabilities.

Equation 4.3 is a simple model of metapopulation dynamics that will serve as a template for developing alternative models. By changing some of our assumptions about colonization and extinction processes, we can generate new metapopulation models that make different predictions about the fraction of sites occupied at equilibrium (\hat{f}). Before exploring these variations, we will first examine the general assumptions of this model.

Model Assumptions

Equation 4.3 makes the following assumptions:

- ✓ **Homogenous patches.** The population sites must not differ in their size, isolation, habitat quality, resource levels, or other factors that would affect the probability of local colonization and local extinction.
- ✓ **No spatial structure.** The model assumes that probabilities of colonization and extinction may be affected by the overall fraction of occupied sites (f), but not their spatial arrangement. In a more realistic metapopulation model, the probability of colonization for a particular site would depend on the occupancy of close neighboring patches, rather than on the overall f . This sort of "neighborhood" model can be studied by computer simulation or by using equations of diffusion, in which the spread of populations through empty sites is analogous to the dispersion of an ink droplet through a beaker of water.
- ✓ **No time lags.** Because we are describing metapopulation dynamics with a continuous differential equation, we assume that the metapopulation "growth rate" (df/dt) responds instantly to changes in f , p_i , or p_e .
- ✓ **Constant p_e and p_i .** The probabilities p_e and p_i do not change from one time period to the next. Although we cannot say precisely which populations will go extinct and which will be colonized, the probabilities of these events do not change.
- ✓ **Regional occurrence (f) affects local colonization (p_i) and extinction (p_e).** Except for the basic island-mainland model (see below), metapopula-

tion models assume that migration is substantial enough to affect local population dynamics and influence probabilities of colonization and/or extinction. Consequently, p_i and/or p_e are functions of f .

✓ **Large number of patches.** The fraction of occupied sites in our model can become infinitely small, and the metapopulation will still persist. Thus, we are not assuming any demographic stochasticity (see Chapter 1) of the metapopulation due to small patch numbers.

Model Variations

THE ISLAND–MAINLAND MODEL

The simplest model for our metapopulation is that both the p_i and p_e are constants. If p_e is a constant, the probability of extinction is the same for each population and does not depend on how many population sites are occupied. This assumption is analogous to a density-independent death rate in a population growth model, because the death rate does not depend on population size (see Chapter 2). Similarly, the probability of colonization may be fixed. Constant p_i implies a **propagule rain**—a continuous source of migrants that could potentially colonize an empty site (Figure 4.2a). If there is a large, stable “mainland” population, it may generate a propagule rain for a set of “islands” in the metapopulation. A propagule rain may also characterize some plant populations that may be colonized by a seed bank of long-lived buried seeds. The equilibrium value of f for this **island–mainland model** can be found by setting Equation 4.3 equal to zero and solving for f :

$$0 = p_i - p_i f - p_e f \quad \text{Expression 4.6}$$

$$p_i f + p_e f = p_i \quad \text{Expression 4.7}$$

Dividing both sides of Expression 4.7 by $(p_i + p_e)$ gives \hat{f} , the equilibrium for f :

$$\hat{f} = \frac{p_i}{p_i + p_e} \quad \text{Equation 4.4}$$

In the island–mainland model, the fraction of sites occupied at equilibrium is a balance between extinction and immigration probabilities. Notice that even if the probability of extinction (p_e) is very large and the probability of colonization (p_i) is very small, at least some of the sites in the metapopulation will be occupied ($\hat{f} > 0$), because the metapopulation is continually replenished by the external propagule rain.

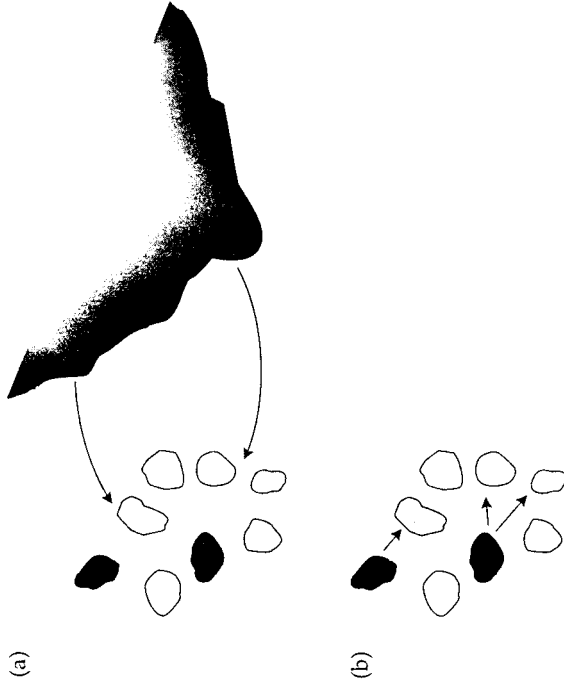


Figure 4.2 (a) Colonization in the island–mainland model. Colonists for a set of islands always come from a large mainland area. Open islands represent empty sites and filled islands represent sites that contain populations. (b) Colonization for the internal model. Colonists do not originate from a permanent external source, but instead originate only from currently occupied islands.

INTERNAL COLONIZATION

Now we will relax the assumption of the propagule rain and instead imagine that the only source of propagules for the metapopulation is the set of occupied population sites (Figure 4.2b). In other words:

$$p_i = if \quad \text{Expression 4.8}$$

The constant i is a measure of how much the probability of colonization of empty sites increases with each additional patch that is occupied. In this model, each population contributes individuals to a pool of propagules, which then have the potential to colonize unoccupied sites. Note that if all of the populations go extinct ($f = 0$), the probability of colonization goes to zero because there is no other source of colonists. This condition is in contrast to the island–mainland model, in which colonists were always present because of the external mainland population.

Assuming that the extinctions are still independent and substituting Expression 4.8 back into the general model (Equation 4.3) gives (Levins 1970):

$$\frac{df}{dt} = if(1-f) - p_c f$$

Equation 4.5

Again, we set this equation equal to zero and solve for the equilibrium f :

$$p_c f = if(1-f)$$

Expression 4.9

$$p_c = i - if$$

Expression 4.10

$$if = i - p_c$$

Expression 4.11

Dividing both sides by i yields:

$$\hat{f} = 1 - \frac{p_c}{i}$$

Equation 4.6

In contrast to the predictions of the island-mainland model, persistence of the metapopulation ($\hat{f} > 0$) is no longer guaranteed. Instead, the metapopulation will persist only if the strength of the internal colonization effect (i) is greater than the probability of local extinction (p_c). If this condition is not met, the metapopulation will go extinct ($\hat{f} \leq 0$). Extinction can happen because the metapopulation is no longer receiving the benefit of external colonization.

THE RESCUE EFFECT

Our first two metapopulation models (island-mainland and internal colonization) both assumed that the probability of extinction was independent of the fraction of sites occupied. Now we should consider the possibility that extinction might be affected by f . How might this happen? As before, we assume that each occupied site produces an excess number of propagules that leave the site and arrive at other populations. If the propagules arrive at an empty site, they represent potential colonists. If conditions are good, these propagules may be able to establish a breeding population in the site. But migrants may also arrive at occupied sites and increase the size of established populations. This increase in N is a **rescue effect** that may prevent the local population from going extinct due to demographic and environmental stochasticity (see Chapter 1). The rescue effect is defined as the reduction in the probability of extinction that occurs when more population sites are occupied, and hence more individuals are available to boost local population sizes.

The tradeoff of propagules leaving a site and those entering from other sites cannot be strictly linear. Otherwise, it would not be possible to achieve a

rescue effect—the reduction in p_c due to immigration would be canceled by the increase in p_c due to emigration. However, the loss of some individuals as migrants may have a negligible effect on p_c . In fact, if migration is density-dependent, individuals that do not migrate to other population sites might reproduce or survive poorly in their sites of origin. An explicit model of the rescue effect would need to include parameters for N , p_c , and migration. But we can capture the essence of the rescue effect in our simple metapopulation model by assuming that:

$$p_c = e(1-f)$$

Expression 4.12

Expression 4.12 says that the probability of local extinction decreases as more population sites are occupied. e is a measure of the strength of the rescue effect, because it controls how much p_c decreases with the addition of another occupied site. Notice that if all population sites are occupied ($f = 1$), the probability of local extinction is zero. This is unrealistic, because even in a saturated landscape there should be some intrinsic background extinction risk. But we would have to introduce another parameter into the model to account for background extinction, so instead we will use Expression 4.12 to keep things simple. Assuming an external propagule rain and a rescue effect, we substitute Expression 4.12 into the general model (Equation 4.3):

$$\frac{df}{dt} = p_i(1-f) - ef(1-f)$$

Equation 4.7

As before, we set Equation 4.7 equal to zero and then solve for the equilibrium value of f :

$$ef(1-f) = p_i(1-f)$$

Expression 4.13

$$ef = p_i$$

Expression 4.14

Dividing both sides by e gives:

$$f = \frac{p_i}{e}$$

Equation 4.8

As in our original island-mainland model, persistence of the metapopulation is assured when there is both a propagule rain and a rescue effect. In fact, if the extinction parameter (e) is less than the probability of colonization (p_i), the metapopulation will be saturated at equilibrium, and all population sites will be occupied ($f = 1$).

OTHER VARIATIONS

One final variation based on our simple metapopulation model would be to combine internal colonization with the rescue effect. In this case, the metapopulation is entirely closed to outside influences; both colonization and extinction probabilities are a function of the fraction of sites occupied. The equation for this model comes from substituting Expression 4.8 (internal colonization) and Expression 4.12 (rescue effect) into Equation 4.3 (the general model):

$$\frac{df}{dt} = if(1-f) - ef(1-f) \quad \text{Equation 4.9}$$

However, if you try to set Equation 4.9 equal to zero and then solve for f , you will find there is no simple solution. Instead, the "equilibrium" depends on the relative sizes of i and e . If $i > e$, the immigration rate [$if(1-f)$] will always be greater than the extinction rate [$ef(1-f)$], so the metapopulation will "grow" until $f = 1$ (landscape saturation). Conversely, if $e > i$, the extinction rate exceeds the immigration rate, and the metapopulation will contract until $f = 0$ (regional extinction). If i and e vary stochastically, the metapopulation may fluctuate between these two equilibrium points (Hanski 1982). Finally, if i equals e , f will not change because the immigration rate will always equal the extinction rate. If some external force changes f , it will then stay at this new equilibrium value. We refer to this as a **neutral equilibrium**.

The metapopulation models that we have considered here have treated colonization as either internal or external. Similarly, extinctions were either independent or mediated by a rescue effect (Table 4.1). These four alternatives actually represent endpoints of a continuum. Colonization in most metapopulations is probably both from propagules generated from within

the system and from propagules derived from external "mainland" sources. Similarly, there are extrinsic and intrinsic forces leading to extinction. These factors can be incorporated into a more general metapopulation model, which includes the four models developed in this chapter as special cases (Gotelli and Kelley 1993).

The derivations presented here just scratch the surface of metapopulation models (Hanski and Gilpin 1991). Other metapopulation models predict N directly, rather than just the presence or absence of populations. Metapopulation models have also been extended to two-species models of competitors or predator and prey. In some cases, species may coexist regionally that cannot coexist locally in closed populations. In other instances, exposing local populations to competitors or predators can lead to extinctions that might not have occurred otherwise. In Chapter 7, we will again return to a discussion of "open" systems when we model the colonization of an island by an entire community of species. For now, we will return to our simple closed models of local populations and incorporate the effects of competitors (Chapter 5) and predators (Chapter 6).

Empirical Examples

THE CHECKERSPOT BUTTERFLY

Populations of the bay checkerspot butterfly (*Euphydryas editha bayensis*) occur in discrete patches that seem to be organized into a large metapopulation. The butterfly is somewhat of a habitat specialist; adult butterflies emerge in spring, and females prefer to lay their eggs on the annual plantain *Plantago erecta*. This host plant serves as a food source for the caterpillars, which feed for one or two weeks and then enter a summer diapause, or resting stage. Caterpillars resume feeding during the cool, rainy months of December to February, and then build cocoons. *P. erecta* grows in Northern California grasslands on serpentine soil rock outcroppings, which serve as potential population sites (Figure 4.3). Populations of the checkerspot butterfly have been studied in this area for over 30 years (Ehrlich et al. 1975).

Fluctuations in the weather can disrupt the life-cycle synchrony of the butterfly and its host plant, leading to local extinction. For example, at least three butterfly populations are known to have gone extinct following a severe drought in 1975–1977 (Murphy and Ehrlich 1980). Very small populations recorded in 1986 may represent successful recolonizations of empty sites (Harrison et al. 1988). The Morgan Hill site is a large patch of serpentine soil that supports a population of hundreds of thousands of butterflies. Because of its large size and the topographic diversity of the site, this population survived the drought and probably served as a source of colonists for empty patches.

Table 4.1 Four metapopulation models.

Colonization	Extinction	
	Independent	Mediated by rescue effect
External ("propagule rain")	$\frac{df}{dt} = pi(1-f) - pcf$	$\frac{df}{dt} = pi(1-f) - ef(1-f)$
Internal	$\frac{df}{dt} = if(1-f) - pcf$	$\frac{df}{dt} = if(1-f) - ef(1-f)$

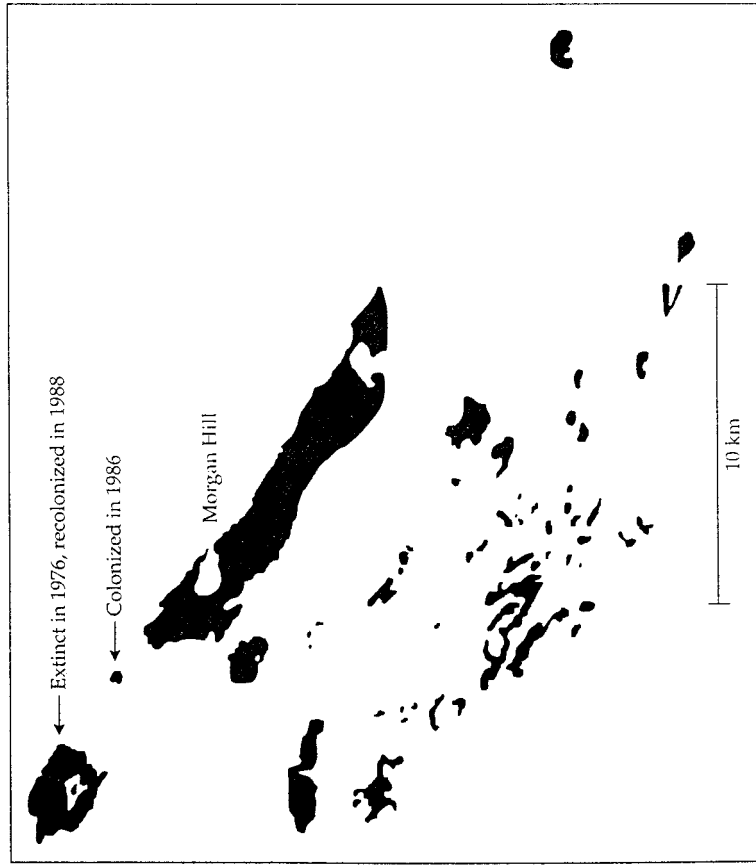


Figure 4.3 Distribution of serpentine soil grasslands in Santa Clara County, California. These habitat patches function as potential population sites for the bay checkerboard butterfly (*Euphydryas editha bayensis*). A large population of butterflies at the Morgan Hill site probably serves as a continual source of colonists for the other small patches, as in the simple island-mainland model. (After Harrison et al. 1988.)

The checkerspot metapopulation is similar, in some respects, to the island-mainland model, in which there is a persistent, external source of colonists. Although our simple metapopulation models assumed that all patches were identical, this was clearly not the case for the checkerspot butterfly. Populations were more likely to be found in sites that were close to the Morgan Hill population, had large areas of cool, north-facing slopes, and high densities of appropriate host plants (Harrison et al. 1988). For conservation purposes, preservation of the Morgan Hill population is probably essential because it provides colonists for other patches.

HEATHLAND CARABID BEETLES

Not all metapopulations occur in well-defined patches, as in the checkerspot butterfly example. Populations may be organized as a metapopulation even in the absence of discrete habitat patches. In the northern Netherlands, populations of carabid ground beetles have been studied by pitfall trapping for over 35 years (den Boer 1981). Radioactive marking revealed that most individuals moved a very limited distance. For example, 90% of the individuals of the beetle *Pterostichus versicolor* moved less than 100 meters a day. Consequently, sites separated by even modest distances effectively contain different subpopulations that are connected by migration.

Figure 4.4a shows the size of 19 subpopulations of *P. versicolor* that were studied for 21 years. Although populations fluctuated asynchronously, there were almost no extinctions recorded during this time period. This is because, at any point in time, some populations were increasing in size and acting as **source populations** that prevented the extinction of other, declining **sink populations**. In contrast, the population fluctuations of the species *Calathus melanocephalus* were much more synchronous during this period (Figure 4.4b). As a consequence, conditions were sometimes uniformly bad for all populations. At these times, there were no source populations available, so population extinctions were much more frequent. Because each subpopulation of *C. melanocephalus* behaved similarly, the risk of extinction was high. Because each subpopulation of *P. versicolor* behaved differently, the metapopulation structure effectively spread the risk of extinction. We still don't understand why the population dynamics of these two beetle species are so different, but it is clear that metapopulation structure affects local extinction and perhaps long-term persistence.

OKLAHOMA STREAM FISHES

Populations of stream fishes may behave as metapopulations because local extinctions are frequent (from floods and droughts), and because individuals migrate between sites. A 10-year study of fishes at 10 sites on the Cimmaron River, Oklahoma (Pigg 1988), provides the data needed to estimate local probabilities of extinction and colonization. For each of the 46 species in the study, the average number of sites occupied and the average probabilities of extinction and colonization can be calculated. As predicted by the metapopulation models from this chapter, p_i decreased and p_c increased as more sites were occupied (Figure 4.5). Notice that, unlike the predictions of our simple models, the graphs of p_i and p_c versus f in Figure 4.5 are not linear. This means that different kinds of equations may be needed to accurately describe the metapopulation. Also notice that each data point in

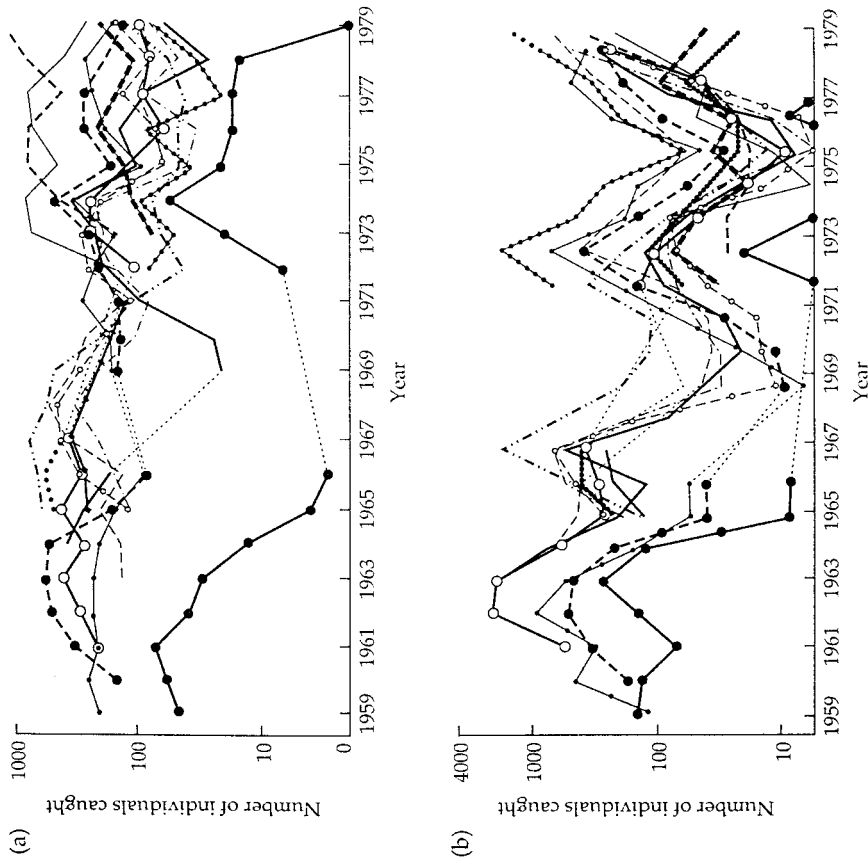


Figure 4.4 (a) Metapopulation dynamics of the ground beetle *Pterostichus versicolor* in heathlands of the northern Netherlands. Each symbol represents the track for a different subpopulation in the heath. Note the great variability in population dynamics and the relative rarity of local extinctions. Broken lines indicate gaps in sampling. Lines that touch the x axis indicate local extinctions. At each time period, some subpopulations are usually increasing, which may act as sources of migrants that prevent the extinction of declining subpopulations. (b) Metapopulation dynamics of the ground beetle *Calathus melanocephalus* in heathlands of the northern Netherlands. In contrast to *P. versicolor*, subpopulations of *C. melanocephalus* tend to fluctuate in synchrony. Consequently, there are no "source" areas to rescue declining subpopulations, so that local extinctions are more frequent. (After den Boer 1981.)

Figure 4.5 represents a different species, whereas the metapopulation model is supposed to describe the behavior of a single species. Graphs can be con-

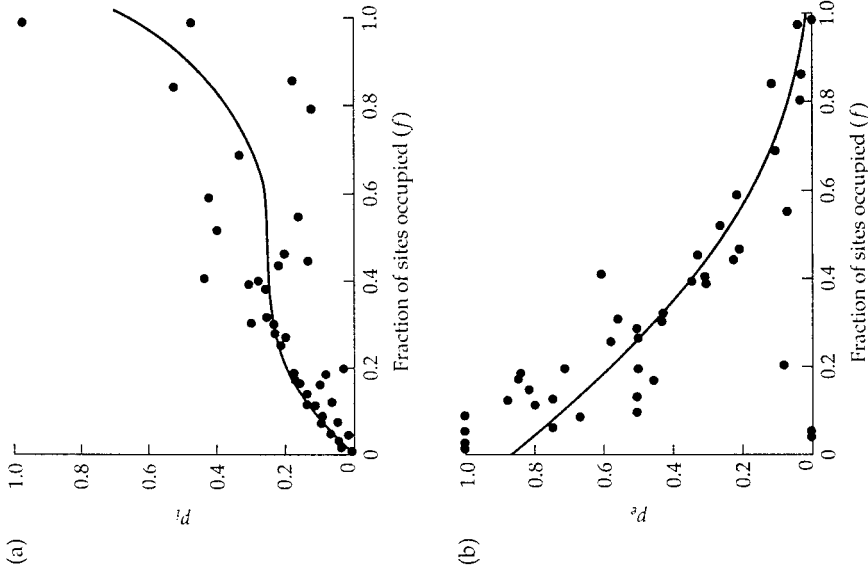


Figure 4.5 Probabilities of local colonization (p_c) and local extinction (p_e) for stream fishes of the Cimarron River, Oklahoma. Each point represents a different species, with data averaged over a 10-year collection period. As predicted by metapopulation models, the probability of colonization (a) increases as more sites are occupied and the probability of extinction (b) decreases as more sites are occupied. In both graphs, the x axis shows the fraction of sites occupied by a species. The solid lines represent best-fit curves. (Data from Pigg 1988.)

structured for individual species that generate similar patterns. As in the butterfly example, habitat quality varied in different locations. Sites near the head of the river in the Oklahoma panhandle were shallower and more variable than sites near the mouth of the river. Consequently, extinction probabilities were higher near the head of the river. Probabilities

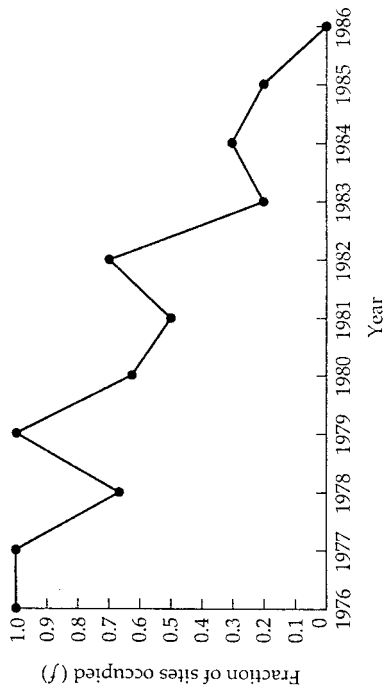


Figure 4.6 Regional extinction of the Arkansas River shiner (*Notropis girardi*) from the Cimmarron River, Oklahoma. The fraction of sites occupied declined steadily over the course of a decade until regional extinction occurred. (Data from Pigg 1988.)

of extinction may also change as a function of time. For example, the Arkansas River shiner (*Notropis girardi*) was a previously widespread species that has steadily disappeared from river drainages in Oklahoma, perhaps due to pollution and changes in water quality. Because extinction probabilities for *N. girardi* usually exceeded colonization probabilities, this species disappeared from the Cimmarron River entirely in 1986 (Figure 4.6). Although the stream fish data confirm the general features of the metapopulation model, more detailed information on population sizes and site quality would be needed to forecast the extinction of a single species such as the Arkansas River shiner.

Problems

1. You are studying a rare and beautiful species of ant lion (see cover). Populations of the ant lion live on a set of islands and on an adjacent mainland that serves as a permanent source of colonists. You can assume that the mainland is the only source of colonists and that extinctions on the islands are independent of one another.

a. If $p_i = 0.2$ and $p_e = 0.4$, calculate the fraction of islands occupied at equilibrium.

*b. A developer is preparing to pave over the mainland area for a new condominium complex. To appease local environmental groups, the developer promises to set aside the islands as a permanent “ant lion nature reserve.” Assuming that $p_i = 0.4$ and $i = 0.2$, predict the fate of the island populations after the mainland population is eliminated.

2. An endangered population of 100 frogs lives in a single pond. One proposal for conserving the frog population is to split it into three populations of 33 frogs, each in a separate pond. You know from your demographic studies that decreasing the frog population from 100 to 33 individuals will increase the yearly risk of extinction from 10% to 50%. In the short run, is it a better strategy to retain the single population or to split it into three?

*3. Suppose a metapopulation has a propagule rain and a rescue effect. The parameters are $p_i = 0.3$ and $e = 0.5$. Forty percent of the population sites are occupied. Is this metapopulation expanding or shrinking?

* Advanced problem

Solutions

1a. Because the source of colonists is external and the extinctions are independent, the metapopulation is described by the island-mainland model (Equation 4.3). In this case, the equilibrium solution is found by solving Equation 4.4:

$$\hat{f} = \frac{p_i}{p_i + p_e} = \frac{0.2}{0.2 + 0.4} = 0.33$$

Approximately one island in three (33%) will support an ant lion population.

1b. Without the mainland population, colonization is strictly internal. In this case, the metapopulation is described by Equation 4.5 (internal colonization, independent extinctions), and the equilibrium solution is from Equation 4.6:

$$\hat{f} = \left(1 - \frac{p_e}{l}\right) = \left(1 - \frac{0.4}{0.2}\right) = -1$$

Because the equilibrium is less than zero, the island populations will all go extinct. Their persistence depends on the presence of the mainland population.

2. For the single pond, the probability of persistence is $(1 - 0.1) = 0.9$. For three ponds, we use Equation 4.2, with a new p_e of 0.50. In this case, the probability that a frog population will persist in at least one of the three ponds is:

$$P_x = 1 - (p_e)^x = 1 - (0.50)^3 = 0.875$$

So, in the short run, the probability of persistence is slightly higher for the population in a single pond (0.9) than for the subdivided populations in three ponds (0.875). In the long run, the best strategy will depend on the dynamics of the frog populations. If the subdivided populations quickly increase in size to 100 or more individuals, it might be worth the short-term risk to cultivate three viable populations rather than just one.

3. Because the metapopulation has a propagule rain and a rescue effect, its dynamics are described by Equation 4.7:

$$\begin{aligned} \frac{df}{dt} &= (p_i)(1-f) - ef(1-f) \\ &= (0.3)(1-0.4) - (0.5)(0.4)(1-0.4) \\ &= 0.18 - 0.12 \\ &= 0.06 \text{ proportion of patches/time} \end{aligned}$$

Because the “growth rate” is greater than zero, the metapopulation is increasing. Also, we could have solved for the equilibrium fraction of sites occupied with Equation 4.8:

$$\hat{f} = \frac{p_i}{e} = \frac{0.3}{0.5}$$

= 0.6 proportion of patches

Because 40% of the population sites are occupied, $f = 0.4$. This is below the equilibrium value, also demonstrating that the metapopulation is expanding.

CHAPTER 5

Competition



Model Presentation and Predictions

COMPETITIVE INTERACTIONS

Chapters 1 through 4 examined single-species population growth. Although we didn't exclude the possibility that other species were important, we did not write explicit equations for populations of predators, prey, or competitors. Instead, the effects of other species were contained in constants such as K , the carrying capacity of the environment (Chapter 2), or p_c , the probability of local population extinction (Chapter 4). In this chapter, we will introduce a second population of a competing species and model the growth of two interacting populations.

Before introducing the model, we need to specify exactly what we mean by "competition." **Competitive interactions** are those in which two species negatively influence each other's population growth rates and depress each other's population sizes. This general definition encompasses a variety of population interactions. **Exploitation competition** occurs when populations depress one another through use of a shared resource, such as food or nutrients. Examples include tropical reef fish that graze on the same kinds of algae, and desert plants that compete for a limited supply of water.

Interference competition occurs when an individual or population behaves in a way that reduces the exploitation efficiency of another individual or population. Examples include song birds that maintain well-established breeding territories, and ant colonies that kill invaders at food patches. Even plants engage in a form of interference competition known as **allelopathy**. Many plant species, particularly aromatic herbs, release toxic chemicals that poison the soil for competitors. The key element in interference competition is that species suppress one another directly, not only through their indirect use of resources.

Interference competition leaves more resources for the winner to consume, so it may evolve as an adaptation when exploitation competition is severe. As an analogy for understanding these two kinds of interactions, exploitation competition is when you and a friend are sitting at a table drinking the same milkshake with straws. The "winner" in exploitation competition is the one who consumes the most milkshake. Interference competition is when you reach over and pinch your friend's straw!

Pre-emptive competition is a third category that has elements of both exploitation and interference. In pre-emptive competition, organisms compete for space as a limiting resource. Examples include birds that use tree holes for nesting and intertidal algae that must attach to stable rock surfaces. Unlike food or nutrients that are used exploitatively, space is a renewable resource that is "recycled"—as soon as an organism dies or leaves, the space is immediately available for use by other individuals.

We need to consider not only the mechanism of competition, but the extent to which competition occurs within and between species. **Intraspecific competition** is competition that occurs among members of the same species. The logistic equation (Equation 2.1) is a model of intraspecific competition because the growth rate diminishes as the population becomes more crowded. **Interspecific competition** is competition between individuals of two or more different species. In this chapter we will build a model of interspecific competition that is a direct extension of the logistic equation.

THE LOTKA-VOLTERRA COMPETITION MODEL

In the 1920s and 1930s, A. J. Lotka and V. Volterra described a simple mathematical model of interspecific competition that is the framework for competition studies in ecology. The model treats populations of two competing species, which we will designate as N_1 and N_2 . Each population grows according to the logistic, with its own intrinsic rate of increase (r_1 or r_2) and its own carrying capacity (K_1 or K_2). As in the logistic model, population growth is reduced by intraspecific competition:

$$\frac{dN_1}{dt} = r_1 N_1 \left(\frac{K_1 - N_1}{K_1} \right) \quad \text{Expression 5.1}$$

$$\frac{dN_2}{dt} = r_2 N_2 \left(\frac{K_2 - N_2}{K_2} \right) \quad \text{Expression 5.2}$$

In our new model, the population growth rate is further depressed by the presence of the second species. For now, assume that the growth is reduced by some function (f) of the number of individuals of the competitor:

$$\frac{dN_1}{dt} = r_1 N_1 \left(\frac{K_1 - N_1 - f(N_2)}{K_1} \right) \quad \text{Expression 5.3}$$

$$\frac{dN_2}{dt} = r_2 N_2 \left(\frac{K_2 - N_2 - f(N_1)}{K_2} \right) \quad \text{Expression 5.4}$$

These expressions show that population growth rate is depressed by both intraspecific and interspecific competition. There are many complicated functions that we could use in Expressions 5.3 and 5.4, but the simplest formula is to multiply the population size of the competitor by a constant number:

$$\frac{dN_1}{dt} = r_1 N_1 \left(\frac{K_1 - N_1 - \alpha N_2}{K_1} \right) \quad \text{Equation 5.1}$$

$$\frac{dN_2}{dt} = r_2 N_2 \left(\frac{K_2 - N_2 - \beta N_1}{K_2} \right) \quad \text{Equation 5.2}$$

COMPETITION COEFFICIENTS

The coefficients α and β are critical to understanding the Lotka-Volterra model. α is a measure of the effect of species 2 on the growth of species 1. If $\alpha = 1$, then individuals of the two species are interchangeable—each has an equal effect in depressing the growth of species 1. On the other hand, suppose that $\alpha = 4$. Each individual of species 2 that is added to the environment depresses the growth of N_1 by the same amount as adding four individuals of species 1. Thus, α is a measure of the relative importance *per individual* of interspecific and intraspecific competition. If $\alpha > 1$, the per capita effect of interspecific competition is greater than the per capita effect of intraspecific competition. If $\alpha < 1$, the intraspecific competition is more important—population growth of species 1 is depressed more by the addition of an individual of N_1 than by addition of an individual of the competing species. Finally, notice that if $\alpha = 0$, there is no competitive effect, and Equation 5.1 reduces to the single-species logistic equation (Equation 2.1). Thus, we can define α as the per capita effect of species 2 on the population growth of species 1, *measured in units of species 1*.

Similar reasoning applies to the interpretation of β , which is the per capita effect of species 1 on the population growth of species 2, measured in units of species 2. It is important to realize that α and β need not have the same values. Competitive effects in nature often are asymmetrical—adding an individual of one species may severely depress the population growth of a second species, whereas the reverse is not true. Although both species in our model coexist in the same location, remember that they each have separate carrying capacities (K_1 and K_2), and intrinsic rates of increase (r_1 and r_2). Although r_1 and r_2 do not affect the outcome of competition in this model, we will see in the next section that the carrying capacities and competition coefficients are critical for determining species coexistence.

A useful way to understand α and β is to return to the analogy we developed in Chapter 2 (Krebs 1985): the carrying capacity of the environment for species 1 is a square frame that holds a limited number of flat tiles (individuals). In our competition model, the tiles come in two different sizes, representing the two different species (Figure 5.1). Continuing the analogy, α is the relative area of the two tiles. For example, if $\alpha = 4$, then a single individual of species 2 consumes four times the remaining resources of the environment as a single individual of species 1. So, a tile of species 2 has four times the area of a tile of species 1. At equilibrium, the frame is filled with a mix of the

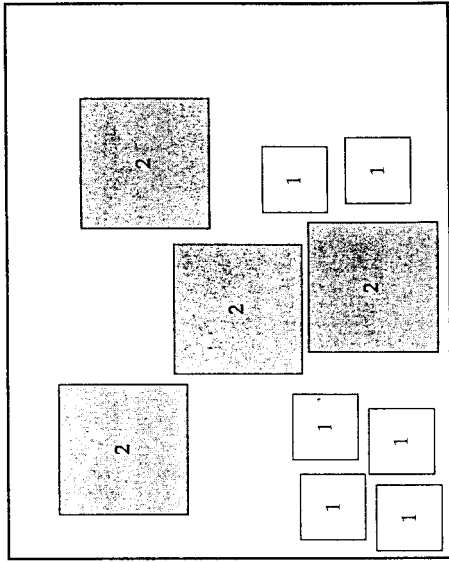


Figure 5.1 A graphical analogy for interspecific competition. The heavy square frame represents the carrying capacity for species 1 (K_1). Each individual consumes a portion of the limited resources available and is represented by a tile. Individuals of species 2 reduce the carrying capacity four times as much as individuals of species 1. Hence, the tiles for species 2 are four times larger than those for species 1, and $\alpha = 4.0$. (After Krebs 1985.)

two kinds of tile. In the next section we will solve for these equilibrium densities.

EQUILIBRIUM SOLUTIONS

As in all our previous analyses, we find the equilibrium population densities (\hat{N}) by setting the differential equations equal to zero and solving for N :

$$\hat{N}_1 = K_1 - \alpha N_2 \quad \text{Equation 5.3}$$

$$\hat{N}_2 = K_2 - \beta N_1 \quad \text{Equation 5.4}$$

These results make intuitive sense. The equilibrium for N_1 is the carrying capacity for species 1 (K_1) reduced by some amount due to the presence of species 2 (αN_2). But we have trouble putting numbers into these solutions—the equilibrium for species 1 depends on the equilibrium for species 2, and vice versa! We can make progress by substituting the equilibrium for N_2 into Equation 5.3, so that the answer will be entirely in terms of N_1 :

$$\hat{N}_1 = K_1 - \alpha(K_2 - \beta \hat{N}_1) \quad \text{Expression 5.5}$$

Similarly, we can substitute the equilibrium for N_1 into Equation 5.4:

$$\hat{N}_2 = K_2 - \beta(K_1 - \alpha\hat{N}_2) \quad \text{Expression 5.6}$$

For each of these expressions, we carry out the multiplication, move all the N terms to the left side of each equation, and arrive at the following solutions:

$$\hat{N}_1 = \frac{K_1 - \alpha K_2}{1 - \alpha\beta} \quad \text{Equation 5.5}$$

$$\hat{N}_2 = \frac{K_2 - \beta K_1}{1 - \alpha\beta} \quad \text{Equation 5.6}$$

Note that for both species to have an equilibrium population size greater than zero, the denominator of each expression must usually be greater than zero. Thus, it is usually the case that the product $\alpha\beta$ must be less than 1 for both species to coexist.

THE STATE SPACE

Although Equations 5.5 and 5.6 tell us the equilibrium conditions for the Lotka–Volterra competition models, they do not provide much insight into the dynamics of competitive interactions, or whether these equilibrium points are stable or not.

We can understand these equations much better by using the **state space**, a special kind of graph. In the state-space graph, the x axis represents the abundance of species 1, and the y axis represents the abundance of species 2. This graph takes a bit of getting used to, but it is an important tool in multispecies models. We will use it again in Chapter 6, when we explore predator–prey models.

What do points in state space represent? A point in this graph represents a *combination of abundances* of species 1 and species 2. The abundance of species 1 can be read from the x axis and the abundance of species 2 can be read from the y axis. If our point falls on the x axis, then only species 1 is present and the abundance of species 2 is zero. For points on the y axis, only species 2 is present. So, the full collection of points in this graph represents all the different combinations of species 1 and species 2 that we could put together.

We use the state-space graph to understand the population dynamics of two competitors. Imagine two competing species whose populations are changing size with time. At each point in time, we could represent their abundances by a single point in the state space (Figure 5.2a). As both populations change in size (Figure 5.2b), we would trace a line in the state space. The final equilibrium point is the end of this line, and if either species goes extinct, this

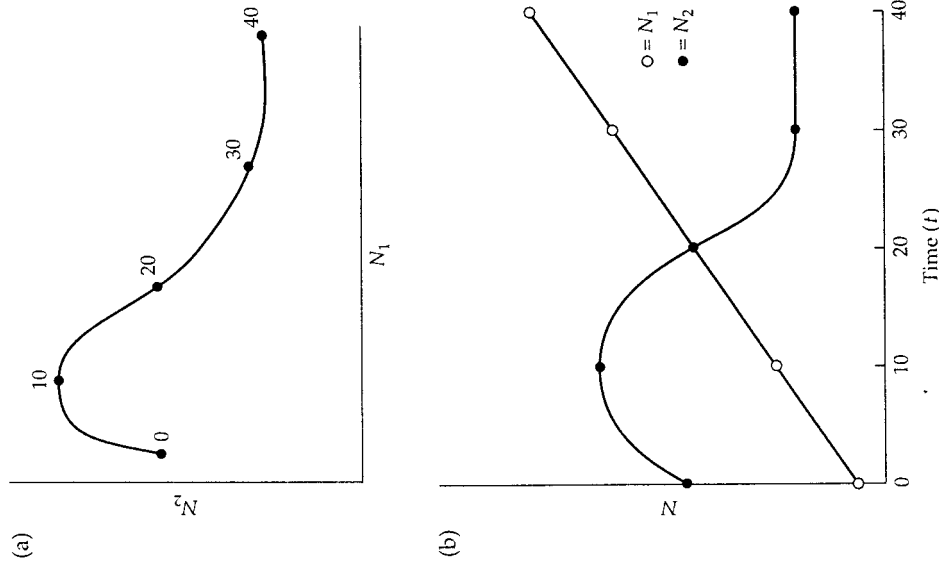


Figure 5.2 (a) A state-space graph. The axes of the state space are the abundances of the two species (N_1 and N_2). As abundances change through time, a curve is traced from left to right. The numbers on the curve indicate time, beginning at 0 and ending at 40. (b) Translation of the state-space graph in (a). The abundances of each species are read from the state-space graph at different times. Note that species 2 first increases and then decreases, whereas species 1 shows a continuous increase in population size.

point falls on one of the two axes of the state-space graph.

How can we use the state-space graph to help us understand the Lotka–Volterra equations? We will first plot Equation 5.3 in the state space. Equation 5.3 is the equilibrium solution for species 1, and its graph is a straight line.

This line represents the combinations of abundances of species 1 and species 2 for which there is zero growth of species 1. At any point on this line, the carrying capacity for species 1 is entirely "filled" with individuals of both species. This line is an **isocline**: a set of abundances for which the growth rate (dN/dt) of one species is zero.

The isocline for species 1 intersects the axes of the state-space graph in two places. The intersection on the x axis is at a value of K_1 . This equilibrium point represents the case in which species 2 is absent and species 1 has grown to its own carrying capacity. The other point is the intersection on the y axis. Here, species 1 is essentially extinct, and the carrying capacity of species 1 is entirely filled with individuals of species 2. The equilibrium solution at this point is K_1/α individuals of species 2 and zero individuals of species 1. Between these extremes are combinations of both species that fall on the isocline (Figure 5.3).

The isocline for species 1 splits the state space into two regions. If we are to the left of the isocline, the joint abundance of N_1 and N_2 is less than the car-

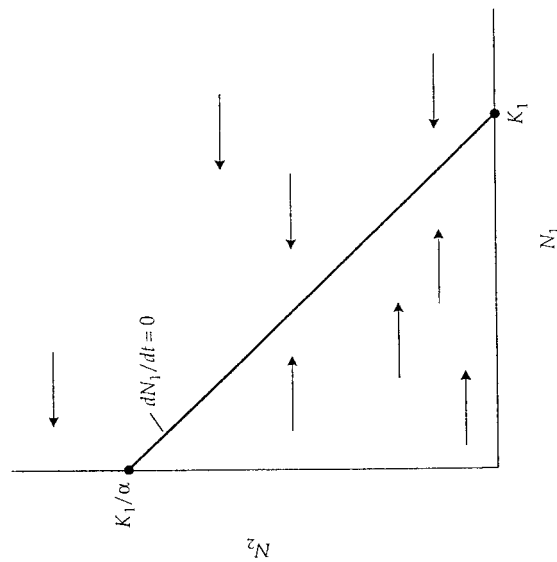


Figure 5.3 The linear isocline for species 1 in the Lotka–Volterra competition model. The isocline defines the combination of abundances for which species 1 shows zero growth. For points to the left of this line, the population of species 1 increases, indicated by the right-pointing horizontal arrow. For points to the right, the joint abundance of species 1 and species 2 exceeds the isocline for species 1, so its population decreases, indicated by the left-pointing arrows.

rying capacity for species 1, so N_1 will increase. An increase in N_1 in the state space is represented as a *horizontal* arrow pointing to the right. The arrow is horizontal because the abundance of species 1 is represented on the x axis. When you work with state-space graphs, pay close attention to which species' isocline you are plotting. Any point to the left of the isocline for species 1 generates a horizontal right-pointing arrow. Under these circumstances, we know that species 1 has a positive growth rate, so its population will increase in size. In contrast, if we are to the right of the isocline, the joint abundance of N_1 and N_2 exceeds the carrying capacity of species 1. In this case, the growth rate of N_1 is negative, and the population decreases. The decrease is represented as a left-pointing horizontal arrow in the state space. Finally, if we are at a point precisely on the isocline, N_1 neither increases nor decreases, and there is no movement in the horizontal direction.

Now we plot the isocline for species 2 in the state space. The isocline of species 2 intersects the y axis at a value of K_2 and intersects the x axis at a value of K_2/β . The first case is one in which species 1 is absent and species 2 has grown to its carrying capacity. In the second case, species 2 is absent, and its carrying capacity is occupied by K_2/β individuals of species 1. Once again, the isocline for species 2 splits the state space into two regions. If we are below the isocline, the joint abundance of species 1 and species 2 is below K_2 and N_2 will increase. Because species 2 is on the y axis, positive growth of species 2 is represented as a *vertical* arrow pointing up in state space. Similarly, if we are above the isocline, the carrying capacity of species 2 is exceeded; its population decreases, represented by a downward-pointing arrow (Figure 5.4).

It is important to recognize that there is a unique isocline for each species that dictates its population growth. By plotting both isoclines together in the state space, we can understand the dynamics of two-species competition. Of course, there are an infinite number of isoclines we could build, simply by using different values of K_1 , K_2 , α , and β . Fortunately, there are only four qualitatively different ways we can plot the isoclines. These four patterns represent the four possible outcomes of competition in the Lotka–Volterra equations.

GRAPHICAL SOLUTIONS TO THE LOTKA–VOLTERRA COMPETITION MODEL

Case 1: Species 1 wins in competition. Figure 5.5 shows one possible configuration of the two isoclines in the state space: the isocline for species 1 lies entirely above the isocline for species 2. In this case, the state space is split into three regions. If we are in the lower left-hand region of the graph, we are below the isoclines of both species, and both species can increase. This is represented by a horizontal and vertical arrow joined at their base. The joint movement of these two populations is represented by the vector sum, which is an arrow that points towards the upper right-hand corner of the graph.

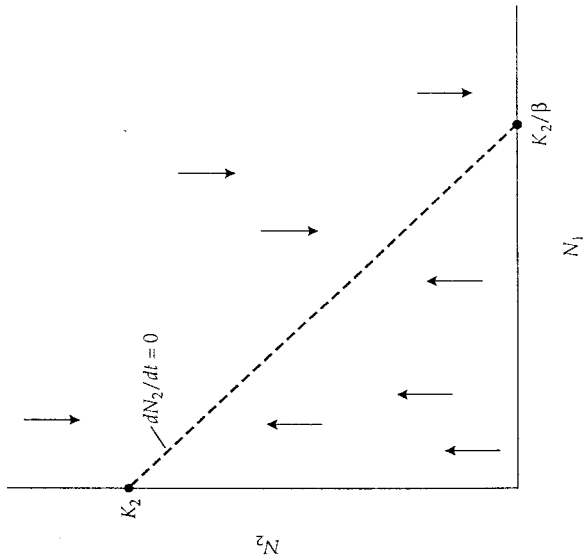


Figure 5.4 The isocline for species 2 in the Lotka-Volterra competition model. Note that the arrows point vertically for species 2, because its abundance is measured on the y axis of the state space graph.

Conversely, if we are in the upper right-hand region of the state space, we are above the isoclines of both species. Both populations will decrease, and the joint vector points towards the origin of the graph.

Things get more interesting in the interior region. Here, we are *below* the isocline of species 1, so its population increases in size, and the horizontal arrow points to the right. However, we are *above* the isocline of species 2, so its population decreases, and the vertical arrow points down. The joint vector points down and to the right, which takes the populations towards the carrying capacity of species 1. Eventually, species 2 declines to extinction, and species 1 increases to K_1 . Notice that, no matter what combination of abundances we start with, the arrows always point towards this outcome. If the isocline of species 1 lies above that of species 2, species 1 always wins in competition, and species 2 is driven to extinction.

Case 2: Species 2 wins in competition. If we graph the isocline of species 2 above that of species 1, then we reverse the conditions and species 2 wins in competition (Figure 5.6). The only difference in this graph is the vector in the interior region. In this case, we are *above* the isocline of species 1, which generates a horizontal arrow to the left, but we are *below* the isocline of species 2, which gener-

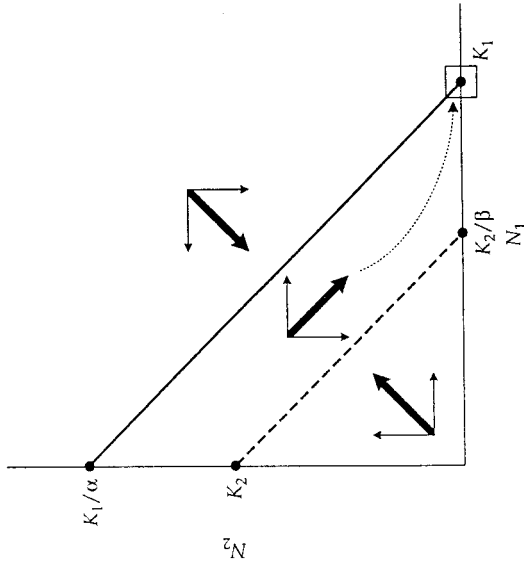


Figure 5.5 Case 1: Competitive exclusion of species 2 by species 1. The thin arrows show the trajectories of each population, and the thick arrow is the joint vector of movement. Competition results in the exclusion of species 2 and an equilibrium for species 1 at carrying capacity. The box indicates a stable equilibrium point.

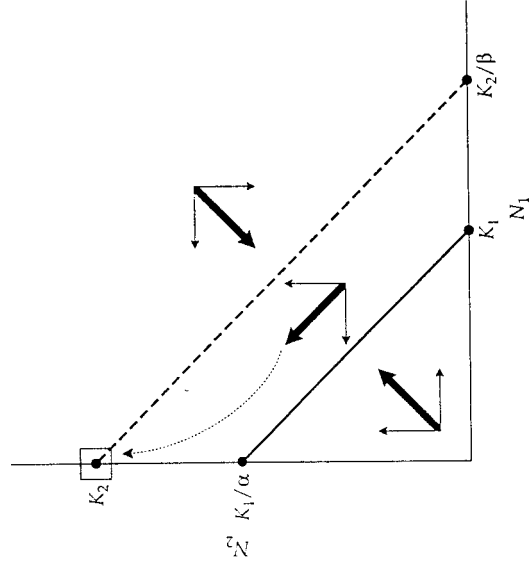


Figure 5.6 Case 2: Competitive exclusion of species 1 by species 2.

ates a vertical arrow pointing up. The joint vector points up and to the left, taking us towards the equilibrium point at K_2 , with N_1 going extinct.

Case 3: Coexistence in a stable equilibrium. The remaining two cases are slightly more complex, because they involve isoclines that cross, dividing the state space into four regions. Nevertheless, the analysis is exactly the same. We simply plot the vectors in each of the four regions to determine the outcome (Figure 5.7). First, note that because the two isoclines cross, there must be an equilibrium point—the crossing of the isoclines represents a combination of abundances for which both species 1 and species 2 have achieved zero growth. The state space analysis reveals whether that equilibrium is stable or not.

As in our previous two examples, the region close to the origin is one of joint growth of both populations, and the region in the upper right-hand corner of the graph is one of joint decrease. The vectors in these regions point towards the equilibrium intersection. If we are in the region of the graph on the lower right, we are *above* the isocline of species 1, but *below* the isocline of species 2. Here, the joint vector points towards the center, as N_1 decreases along the horizontal axis and N_2 increases along the vertical axis. Finally, if

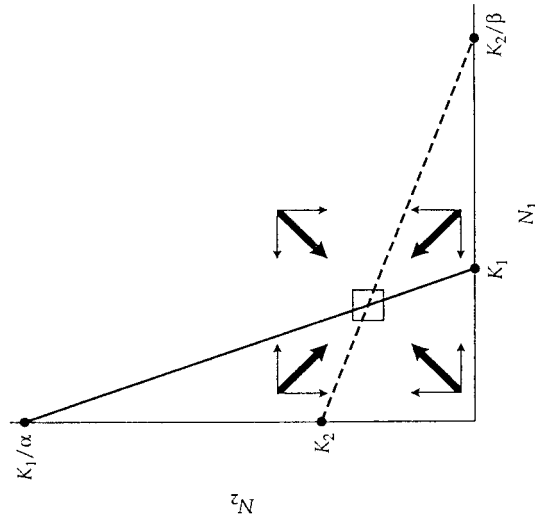


Figure 5.7 Case 3: Coexistence in a stable equilibrium. The two isoclines cross, and the joint vectors point in towards the equilibrium point. The equilibrium is stable because if the populations are displaced, they will always return to their equilibrium sizes.

we are in the region of the graph on the upper left, we are *above* the isocline of species 2, but *below* the isocline of species 1, and the joint vector again points towards the center.

This is a stable equilibrium in which all roads lead to Rome—no matter what the initial abundances of the two species are, both populations will move towards the joint equilibrium value. Although this equilibrium is stable and both species coexist, note that each species persists at a lower abundance than it would in the absence of its competitor. Competition reduces the population size of each species, but neither can drive the other extinct.

Case 4: Competitive exclusion in an unstable equilibrium. This final case is the one in which the isoclines cross in the opposite way (Figure 5.8). Once again, both populations increase in the sector closest to the origin, and both populations decrease in the upper right-hand region. But the pattern changes in the two remaining slivers of state space. In the lower right-hand region, we are *below* the isocline of species 1, but *above* the isocline of species 2. In this region of the graph, the populations move *away* from the joint equilibrium and towards K_1 . Similarly, in the fourth region of the state space, we are *above* the isocline for N_1 , but *below* that for N_2 . The populations move away from the joint equilibrium and towards K_2 .

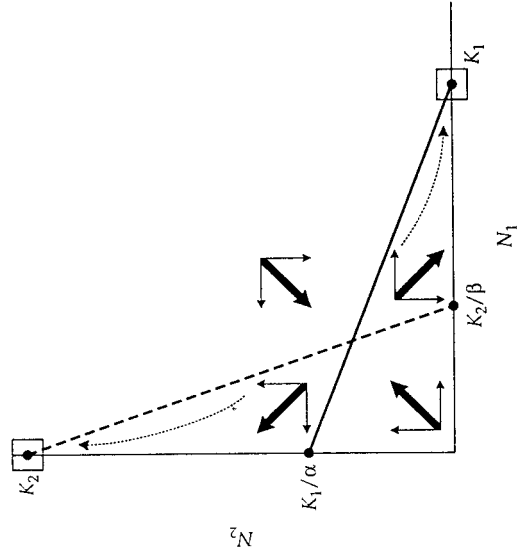


Figure 5.8 Case 4: Competitive exclusion in an unstable equilibrium. The two isoclines again cross and form an equilibrium point. However, the joint vectors point away from this equilibrium. If the populations are displaced, one species or the other will win in competition, depending on the starting abundances.

Case 4 represents an **unstable equilibrium**. If the populations are displaced from the joint equilibrium, they will eventually end up in one of the two regions of the graph that will take them to competitive exclusion. Thus, both species cannot persist in the long run, and one will be driven to extinction by competition. However, the winner is difficult to predict. The population that has a numerical advantage is the one that will probably win in competition, but the outcome depends on the initial position in the state space, and the relative growth rates of the two competitors (r_1 and r_2).

THE PRINCIPLE OF COMPETITIVE EXCLUSION

Now that we understand the four graphical solutions to the Lotka–Volterra equations, we will take another look at the algebraic solutions. We can reason that species 1 will always persist if it can increase under the worst possible circumstances. The worst scenario for species 1 is that its own abundance is close to zero ($N_1 \approx 0$), and the abundance of its competitor is close to carrying capacity ($N_2 \approx K_2$). If N_1 can achieve a positive per capita growth rate [$(dN_1/dt)/(N_1) > 0$] under these circumstances, then it should always be able to persist (MacArthur 1972). Plugging these conditions into Equation 5.1 gives:

$$\left(\frac{dN_1}{dt}\right)\left(\frac{1}{N_1}\right) = r_1\left(\frac{K_1 - 0 - \alpha K_2}{K_1}\right) \tag{Expression 5.7}$$

Since r_1 is always positive, the following inequality must hold for N_1 to increase:

$$\frac{K_1 - \alpha K_2}{K_1} > 0 \tag{Expression 5.8}$$

which reduces to:

$$\frac{K_1}{K_2} > \alpha \tag{Expression 5.9}$$

If species 1 is to successfully invade, the ratio of the carrying capacities must exceed the competitive effect of species 2 on species 1. In other words, if species 2 is a strong competitor, species 1 must have a relatively large carrying capacity to persist.

Using Equation 5.2, we can go through a similar calculation to arrive at the following inequality for the persistence of species 2:

$$\frac{K_2}{K_1} > \beta \tag{Expression 5.10}$$

Flipping the inequality makes this directly comparable with Expression 5.9:

$$\frac{1}{\beta} > \frac{K_1}{K_2} \tag{Expression 5.11}$$

Table 5.1 Algebraic inequalities defining the persistence of species and the outcome of competition in the Lotka–Volterra equations.

(a)	
Inequality	Outcome
$\frac{K_1}{K_2} > \alpha$	Species 1 persists
$\frac{K_1}{K_2} < \alpha$	Species 1 does not persist
$\frac{K_1}{K_2} < \frac{1}{\beta}$	Species 2 persists
$\frac{K_1}{K_2} > \frac{1}{\beta}$	Species 2 does not persist

(b)		
Species 1 persists	Species 2 persists	Outcome
Yes	No	Species 1 wins (Case 1)
No	Yes	Species 2 wins (Case 2)
Yes	Yes	Stable coexistence (Case 3)
No	No	Unstable equilibrium (Case 4)

Now we have expressions for whether N_1 will persist or not, and whether N_2 will persist or not. Putting these expressions together generates four algebraic inequalities that define the four graphical solutions to the Lotka–Volterra equations. For example, if species 1 can persist ($K_1/K_2 > \alpha$), but species 2 cannot ($1/\beta < K_1/K_2$), then we have defined the conditions for case 1, in which species 1 always wins in competition. If both species are able to persist, we have the stable coexistence of case 3, whereas if neither species can always persist, we have the unstable equilibrium of case 4 (Table 5.1).

These inequalities give us insight into one of ecology’s enduring proverbs, the **principle of competitive exclusion**. Briefly stated, the principle is that “complete competitors cannot coexist” (Hardin 1960). In other words, there must be some difference between species in resource use if species are able to coexist.

If two species are very similar in their resource use, then α and β should be very close to 1. Suppose, for example, that $\alpha = \beta = 0.9$. From the inequality

ity in Table 5.1, coexistence of these species requires that:

$$\frac{1}{0.9} > \frac{K_1}{K_2} > 0.9$$

Expression 5.12

$$1.1 > \frac{K_1}{K_2} > 0.9$$

Expression 5.13

Thus, if the species are very similar in their use of resources, there is only a narrow range of carrying capacities that will ensure stable coexistence. In contrast, suppose that $\alpha = \beta = 0.2$, indicating that species differ greatly in their use of common resources. In this case, coexistence will occur if:

$$5 > \frac{K_1}{K_2} > 0.2$$

Expression 5.14

In this case, the two species will coexist with a wide range of possible carrying capacities. Thus, our analysis of the Lotka–Volterra equations allows us to refine the competitive exclusion principle: the more similar species are in their use of shared resources, the more precarious their coexistence.

The Lotka–Volterra equations are the simplest two-species model of competition. As you might expect, it is even more difficult to obtain coexistence of species in models that have three or more competitors. For many years, ecologists have studied the “coexistence problem,” and discovered that species often coexist in nature with little apparent difference in resource exploitation. In these circumstances, one or more of the following assumptions of the model has been violated.

Model Assumptions

As in the logistic and exponential growth models, we assume there is no age or genetic structure to the populations, no migration, and no time lags. The following assumptions also apply to the Lotka–Volterra model:

- ✓ **Resources are in limited supply.** The result of resource limitation is both intra- and interspecific competition. If resources are not limiting, then an infinite number of species can coexist, regardless of how similar they are in resource use.
- ✓ **Competition coefficients (α and β) and carrying capacities (K_1 and K_2) are constants.** If these parameters should change with time or density, it may be difficult to predict species coexistence.

✓ **Density dependence is linear.** Adding an individual of either species produces a strictly linear decrease in per capita population growth rate. This is reflected in the linear isoclines of the Lotka–Volterra model. Models with nonlinear isoclines have more complex stability properties that are not easy to deduce from simple state-space graphs.

Model Variations

INTRAGUILD PREDATION

Ecologists classify species interactions according to their effects on population growth rate. Thus, competition is defined as both species having a net negative effect on one another ($-,-$), mutualism as both species having a net positive effect ($+,+$), and predation or parasitism as one species gaining and the other species losing ($+,-$). These classifications are convenient and natural, and they reflect our model assumptions that interaction coefficients are constant and that there is no age structure in the populations.

But when we study the natural history of many animals, we find they cannot be classified simply as “predators” or “competitors.” For example, lions prey on the young of cheetahs, wild dogs, and spotted hyenas, but also compete with these same species for prey. Flour beetles in the genus *Tribolium* compete for food, but at high densities they also consume one another’s larvae. For many predators, diet is determined strictly by their size and what they can get their jaws around. As individuals age, their diets can change radically. Anyone who has tried to raise baby fish in an aquarium can appreciate that predation is often critically tied to body size. Individuals of a single species may act as prey, competitors, or predators, depending on their age and size. **Intraguild predation (IGP)** is the ecological interaction in which two competing species also interact as predator and prey. IGP is not an isolated phenomenon; it is common in terrestrial, marine, and freshwater communities, and probably represents the rule rather than the exception in nature (Polis et al. 1989).

How can we modify our simple competition model to take account of IGP? Suppose that two species compete according to the Lotka–Volterra equations, but species 1 is also a predator on species 2. This is a simple model that does not involve age structure, reciprocal predation, or cannibalism. However, it at least illustrates the way that IGP can modify ecological interactions. The growth equation for species 1 (“predator”) is:

$$\frac{dN_1}{dt} = r_1 N_1 \left(\frac{K_1 - N_1 - \alpha N_2}{K_1} \right) + \gamma N_1 N_2 \quad \text{Equation 5.7}$$

This is identical to the original Lotka–Volterra model, except we have added an additional term. This addition represents the increase in growth rate that species 1 receives by feeding on species 2. The amount of this increase depends on the encounter rate of predator and prey (N_1N_2) and a conversion constant (γ). We will see a similar expression in Chapter 6 when we build a predator–prey model. The growth equation for species 2 (“prey”) is:

$$\frac{dN_2}{dt} = r_2N_2 \left(\frac{K_2 - N_2 - \beta N_1}{K_2} \right) - \delta N_1N_2 \quad \text{Equation 5.8}$$

Again, growth of species 2 is described by the Lotka–Volterra model, but is further reduced because of losses due to predation by species 1. These losses also depend on the encounter rate of predator and prey (N_1N_2) and a conversion factor (δ). Note that the conversion factors for predator (γ) and prey (δ) need not be equivalent. The loss of an individual to predation usually does not correspond to a symmetrical gain for the predator population. Again, these ideas are explored more thoroughly in Chapter 6.

How does IGP affect the coexistence of species? The graphical effect of IGP is to rotate the isoclines. IGP does not change the carrying capacity for either predator or prey. Instead, it changes the density of the competitor that is necessary to cause extinction. Consequently, each isocline is rotated up or down, but remains fixed at the intercept on its own axis. For the predator, the isocline swings up, because it now requires more competitors to drive the predator to extinction than before (Figure 5.9a). For the prey species, IGP swings the isocline in towards the origin, because it now requires fewer competitors to cause extinction (Figure 5.9b).

IGP can either reinforce or reverse the outcome of competition, depending on the position of the isoclines and the amount of rotation (which is ultimately controlled by the conversion factors). For example, if the inferior competitor is also the prey species, IGP merely adds the insult of predation to the injury of competition and reinforces the extinction of species 2 (Figure 5.10a). But if the inferior competitor is the predator, IGP can change the outcome from competitive exclusion (case 1) to stable coexistence (case 3; Figure 5.10b). Other outcomes are possible, and IGP may provide insight into species coexistence when simple competition and predation models fail (Polis et al. 1989).

Empirical Examples

COMPETITION BETWEEN INTERTIDAL SANDFLAT WORMS

In northern Puget Sound, many species of marine worms coexist in intertidal sandflats at very high densities. Abundances can be manipulated experi-

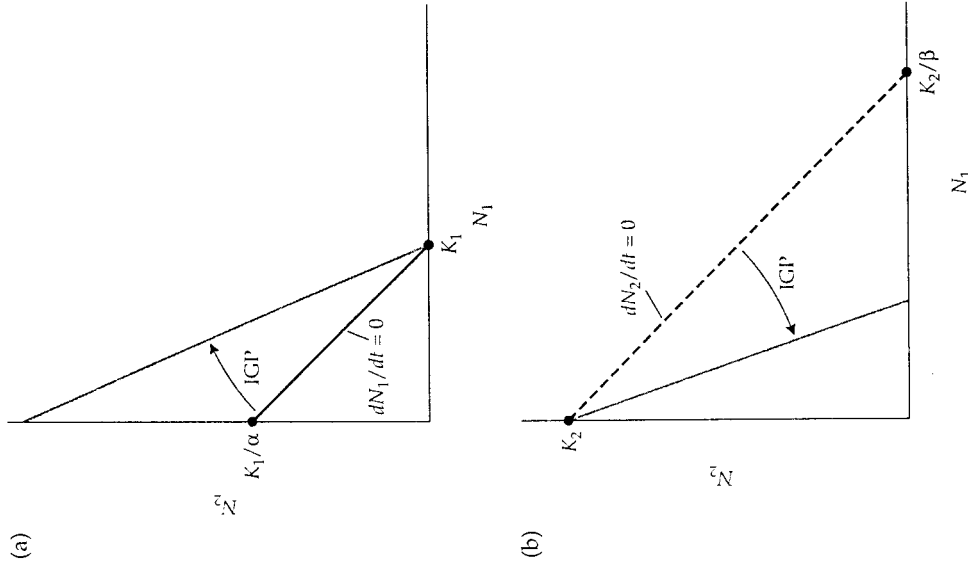


Figure 5.9 (a) Intraguild predation rotates the isocline of the “predator” species up, because it now requires more individuals of the competitor–prey to drive it to extinction. (b) Intraguild predation rotates the isocline of the “prey” species down, because it now requires fewer individuals of the competitor–predator to drive it to extinction.

mentally, allowing for a direct test of the Lotka–Volterra competition model. Gallagher et al. (1990) examined competition between juveniles of the polychaete *Hobsonia florida* and a number of closely related species of oligochaetes. Both *Hobsonia* and the oligochaetes coexist in dense aggregations in nature and feed on benthic diatoms.

Gallagher et al. (1990) used field experiments to determine whether such coexistence could be successfully predicted by the Lotka–Volterra model. By adding predatory shrimp to small (26-centimeter diameter) field enclosures, the authors were able to manipulate the densities of *Hobsonia* and the

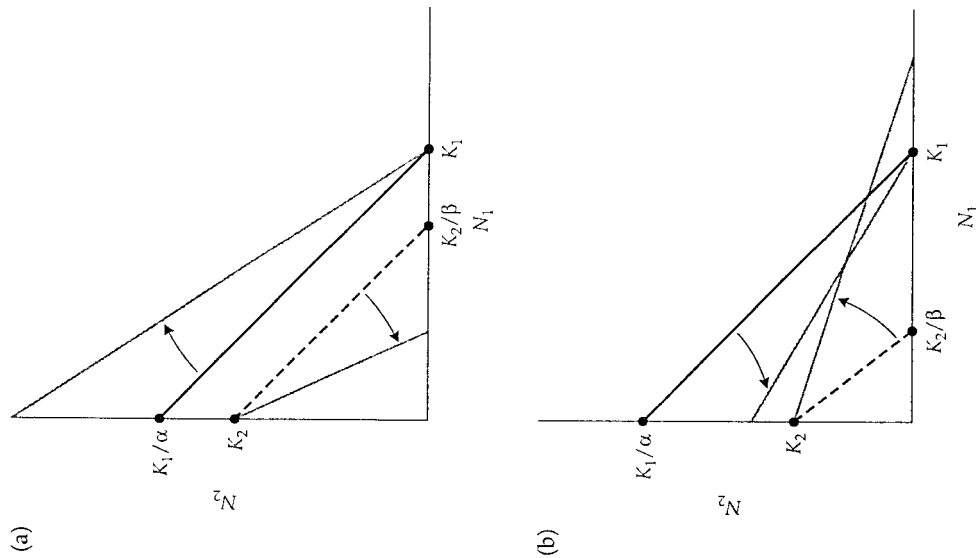


Figure 5.10 (a) Intraguild predation reinforces competitive exclusion. In this example, the superior competitor (N_1) is also the predator, so the shifted isoclines lead to the same outcome. (b) Intraguild competition reverses competitive exclusion. In this example, the inferior competitor (N_2) is now the predator. The isoclines shift from competitive exclusion (Case 1) to stable coexistence (Case 3).

oligochaetes in the patch. These starting densities represented a single point in the state space. Next, they measured the increase and decrease of each population in the patch after three days. These changes revealed the vector of population dynamics in the state space. By repeating this procedure for different starting densities, they were able to determine the placement of both isoclines. These field experiments produced the following estimates: K_1 (*Hobsonia*) = 64.2, α = 1.408; K_2 (oligochaetes) = 50.7, β = 0.754. Finally, the authors started two patches at a low initial abundance of both competitors and then followed their dynamics for 55 days.

The isoclines for both species are plotted in Figure 5.11. Superimposed on this state space is one of the trajectories for the 55-day experiment. Because the isocline for the oligochaetes lies slightly above the isocline for *Hobsonia*, the model predicts that the oligochaetes should win in competition. But the trajectory of the 55-day experiment did not reach the oligochaete carrying capacity, and in nature, both species coexist. The simple Lotka–Volterra model must be rejected for this system.

Why did the model fail to give us the correct predictions? Because the isoclines of the two species are very close to one another, the predicted time to

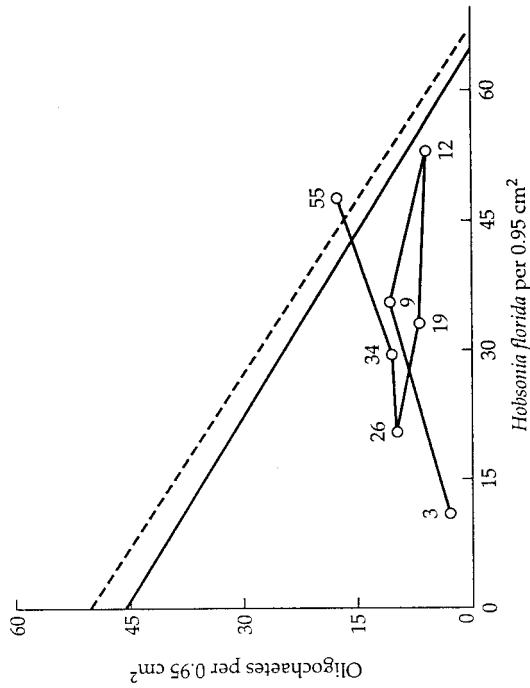


Figure 5.11 Competition between marine intertidal worms. The solid line is the estimated isocline for *Hobsonia florida*, and the dashed line is the estimated isocline for the oligochaetes. The line segments trace an experiment in the state space that was started with low abundances of both competitors. The numbers indicate the number of days since the start of the experiment. (From Gallagher et al. 1990.)

extinction is long. Moreover, there are seasonal changes in diatom abundance, so that the carrying capacities for each species are always changing. When carrying capacities change, the isoclines “wobble” through time, so that population trajectories may be continually changing. Under these conditions, there may not be enough time for one species to win in competition. Thus, the oligochaetes do not competitively exclude *Hobsonia* because the environment is always changing. As the ecologist G. E. Hutchinson (1967) wrote: “The competitors of a given genus or other higher taxon are from time to time lined up, and sometimes the race begins, but as it might be in the works of Lewis Carroll, the event is always called off before it is completed and something entirely different is arranged in its place.”

THE SHAPE OF A GERBIL ISOCLINE

Gerbils are mouse-like rodents of the deserts of Africa and the Middle East. They are nocturnal seed foragers, and the coexistence of several gerbil species may depend on their use of common food and habitat resources. Abramsky et al. (1991) studied the coexistence of *Gerbillus allenbyi* and *G. pyramidum* in the western Negev desert of Israel.

Experimental studies of vertebrate competition are particularly difficult because of the large areas needed to enclose populations, and because competition is often mediated by subtle behavioral interactions. Abramsky et al. (1991) took advantage of the fact that *G. pyramidum* is considerably larger than *G. allenbyi* (mean mass = 40 grams versus 26 grams). The authors built enclosures that were 100 meters on a side (one hectare in area). Each enclosure was separated into two plots by a common fence. This fence had small gates to permit gerbils to move between the two sides. The gates were large enough to allow *G. allenbyi* through, but too small for *G. pyramidum* to pass. Thus, the fence acted as a semipermeable membrane, allowing *G. allenbyi* to “equilibrate” its density on the two sides based on the density of *G. pyramidum*.

Although the Lotka-Volterra model predicts changes in population growth rate, these are difficult to measure in short-term experiments on vertebrates. Moreover, the effects of competition on gerbil populations are likely to be expressed more immediately in changes in behavior and foraging activity. Instead of measuring gerbil density, the authors measured the “activity density” of each species by counting gerbil footprints in clean trays of sand that were placed in the plots each night. This index was correlated with density and foraging activity of individual gerbils.

The authors established one half of each enclosure with a high density of *G. pyramidum* and the other half with a low density. The density of *G. allenbyi* was allowed to equilibrate to these differences in competitor density. The

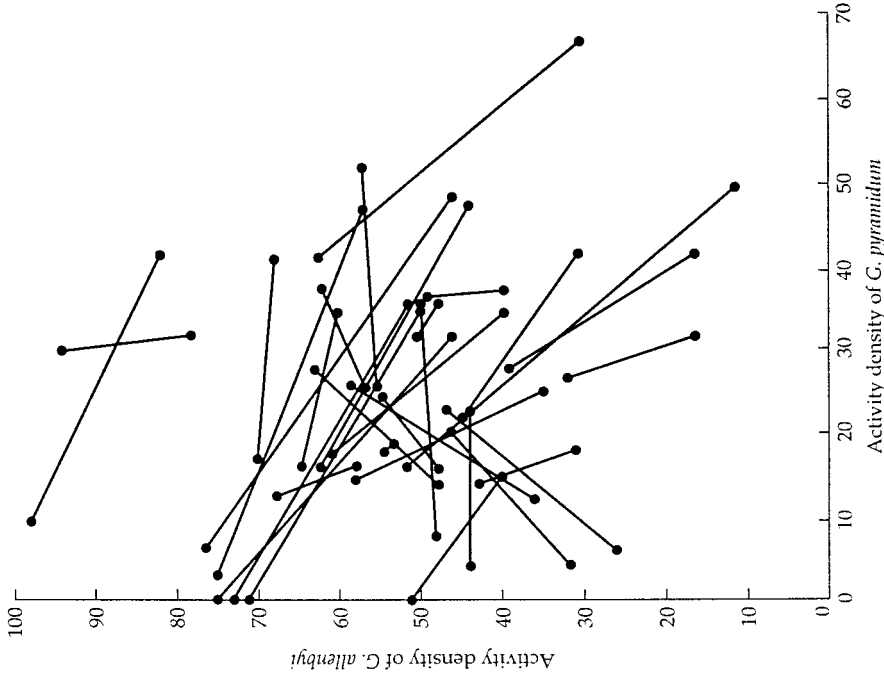


Figure 5.12 Results of gerbil competition experiments plotted in state space. Each line segment connects the points for the high-density and low-density plots in an experimental enclosure. Most of the segments have a negative slope, indicating a reduction in the activity density of *Gerbillus allenbyi* (*y* axis) in the presence of its competitor *G. pyramidum* (*x* axis).

resulting changes in activity of both species can be plotted in state space. Each line segment in this graph represents the activity density in the two halves of an enclosure. The slope of this line segment is a measure of the isocline of *G. allenbyi* in that area of the state space (Figure 5.12). Although there is considerable scatter in the data, most of these segments have a negative slope, indicating that high densities of *G. pyramidum* depressed the activity of *G. allenbyi*.

Figure 5.13 shows the isocline based on the “best fit” of all these line segments. In contrast to the predictions of the Lotka–Volterra model, the isocline for *G. allenbyi* is nonlinear, with steep declines at high and low densities of *G. pyramitidum*, but a shallow slope at intermediate competitor densities.

Why isn't the isocline of *G. allenbyi* a straight line? The answer is that activity density depends not only on the abundance of competitors, but also on the availability and use of different habitats. In the Negev Desert, there are two habitat types that the gerbils use. “Semistabilized dunes” contain little perennial vegetation, many open patches of sand, and unstabilized sand dunes. “Stabilized sand” habitats are dominated by dense shrub cover, with large areas of stable soil crust and few open patches. Both habitat types were present in approximately equal abundance within each enclosure.

Under uncrowded conditions, both gerbil species preferred the semistabilized dunes. As intraspecific density increased, both species began to use the stabilized sand in greater frequency. *G. pyramitidum* density induced a habitat shift in *G. allenbyi*, and this was responsible for the nonlinear isocline. Superimposed on the state space in Figure 5.13 are four lines (“isolegs”) that are cutpoints for changes in habitat use of the two species. At low densities (regions I and II), both species preferred the semistabilized dunes, and increased densities of *G. pyramitidum* led to a sharp decrease in the activity density of *G. allenbyi*. As the density of *G. pyramitidum* increased, *G. allenbyi* did not decrease its activity, but instead shifted into the less preferred stabilized sand habitat. Consequently, the isocline is relatively flat in this region, reflecting habitat shift, rather than a reduction in activity density. But as the density of *G. pyramitidum* increased, it was also forced to use the stabilized sand habitat. At high densities of *G. pyramitidum*, *G. allenbyi* could no longer “escape” competition by moving to an unoccupied habitat, so activity density again dropped off steeply. Additional field experiments measured the isocline of *G. pyramitidum* (Abramsky et al. 1994), and a mathematical analysis predicts stable coexistence of both competitors.

The Lotka–Volterra model generates simple predictions and provides a framework for field tests of competition. Nevertheless, it is very difficult to manipulate species densities in realistic field experiments, and it is still an open question as to whether resources are limiting. These studies show that even when resources are limiting, the model's simple predictions may fail because factors such as variable environments and habitat selection can also affect the outcome of interspecific competition.

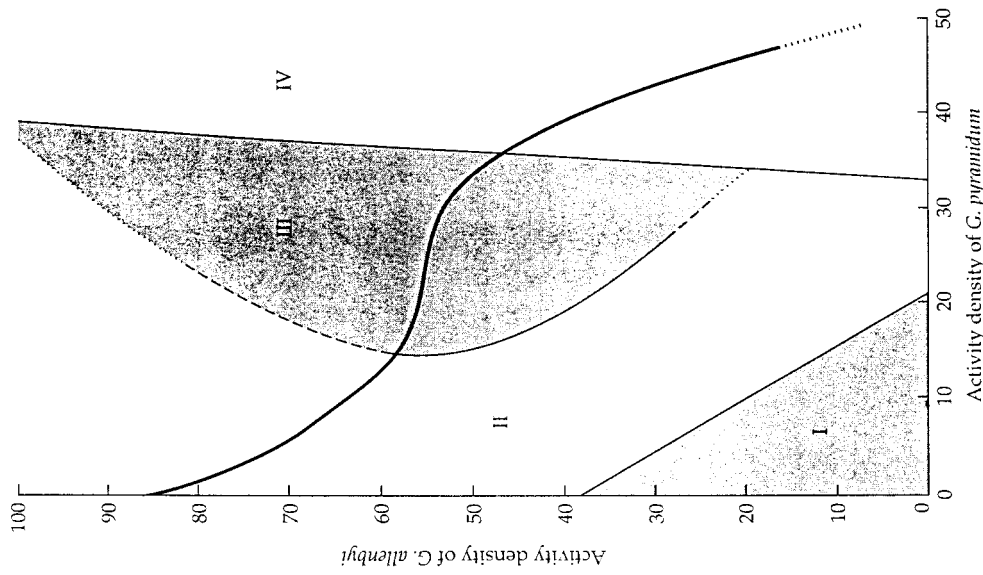


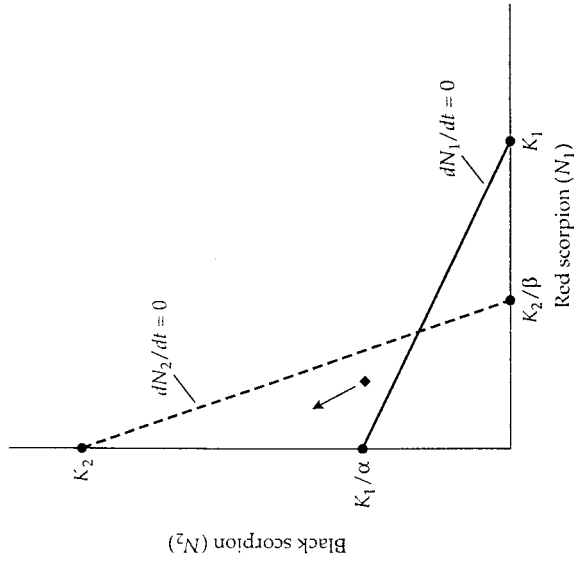
Figure 5.13 The isocline for *Gerbillius allenbyi*, estimated from the data in Figure 5.12. Note that the isocline (thick line) is not linear, but has a region of shallow slope at intermediate densities of *G. pyramitidum*. This nonlinear isocline reflects the effects of competition and habitat selection. The thin lines divide the state space into regions based on habitat use. Region 1: Both species use the preferred habitat, semistabilized dune. Region 2: *G. allenbyi* is forced to use the less preferred habitat, stabilized sand. Region 3: Increased use of the stabilized sand by *G. allenbyi*. Because *G. allenbyi* shifts to its less preferred habitat, its activity density can remain high, leading to a shallow slope for the isocline in this region of the state space. Region 4: *G. pyramitidum* is forced into the stabilized dune habitat by intraspecific competition. Because *G. allenbyi* no longer has an escape to the unoccupied habitat, its activity density drops off sharply with increases in the activity density of *G. pyramitidum*.

Problems

1. You are studying competition between red and black desert scorpions. For the red scorpion, $K_1 = 100$ and $\alpha = 2$. For the black scorpion, $K_2 = 150$ and $\beta = 3$.
Suppose the initial population sizes are 25 red scorpions and 50 black scorpions. Graph the state space and isoclines for each species, and plot these initial population sizes. Predict the short-term dynamics of each population and the final outcome of interspecific competition.
2. Suppose that, for two competing species, $\alpha = 1.5$, $\beta = 0.5$, and $K_2 = 100$. What is the minimum carrying capacity for species 1 that is necessary for coexistence? How large is the carrying capacity needed for species 1 to win in competition?
- *3. Diagram the state space for two competing species in which there is a stable equilibrium. Show how intraguild predation could shift this to exclusion by the predatory species.

Solutions

1. Here is the plot of the state space, with the initial population sizes indicated by the diamond:



These isoclines define an unstable equilibrium. From the initial densities, the black scorpion will increase and the red scorpion will decrease in the short run. Eventually, the red scorpion will be driven to extinction, and the black scorpion will persist at its carrying capacity (K_2) of 150.

2. To answer this question, we use the inequalities in Table 5.1. Coexistence requires that:

$$\frac{1}{\beta} > \frac{K_1}{K_2} > \alpha$$

$$\frac{1}{0.5} > \frac{K_1}{100} > 1.5$$

$$2 > \frac{K_1}{100} > 1.5$$

A minimum carrying capacity of 151 individuals for species 1 is necessary to satisfy this inequality and ensure coexistence.

* Advanced problem

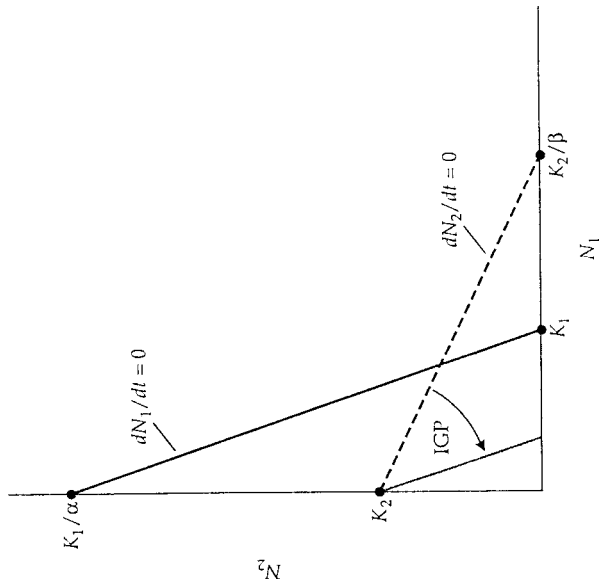
If species 1 is going to win in competition, then:

$$\frac{1}{\beta} < \frac{K_1}{K_2} > \alpha$$

$$2 < \frac{K_1}{100} > 1.5$$

For this to happen, the carrying capacity of species 1 must be greater than 200 individuals.

3. The following diagram illustrates the two isoclines for the initial stable equilibrium. The arrow indicates the shift in the "prey" isocline that allows the "predator" to win in competition:



CHAPTER 6

Predation



Model Presentation and Predictions

Competitive interactions in nature are often indirect and subtle, and may be mediated through populations of resources. In contrast, predation is a direct and conspicuous ecological interaction. The image of a wolf pack bringing down a moose, or a spider eating a fly evokes Tennyson's description of "nature red in tooth and claw." Seed predators, such as finches and harvester ants, are less dramatic in their feeding, but may be equally effective at controlling plant populations. Other animals do not consume their prey entirely. Parasites require that their hosts survive long enough for the parasite to reproduce, and many herbivores graze on plants without killing them. In all of these interactions, we can recognize a population of "predators" that benefit from feeding, and a population of "victims" that suffers. In this chapter, we will develop some simple models to give us insight into the dynamics of predation. As in our analysis of competition, the predation equations were first derived independently by A. J. Lotka and V. Volterra. Volterra's interest in the subject stemmed from his daughter's fiancé, a fisheries biologist who was trying to understand fluctuations in the catch of predaceous fish (Kingsland 1985).

MODELING PREY POPULATION GROWTH

We will use the symbol P to denote the predator population and the symbol V to denote the victim or prey population. The growth of the victim population will be some function, f , of the numbers of both victims and predators:

$$\frac{dV}{dt} = f(V, P) \quad \text{Expression 6.1}$$

Suppose that the predators are the only force limiting the growth of the victim population. In other words, if the predators are absent, the victim population increases exponentially:

$$\frac{dV}{dt} = rV \quad \text{Expression 6.2}$$

with r representing the intrinsic rate of increase (see Chapter 1). This potential for increase of the victim population is offset by losses that occur when predators are present:

$$\frac{dV}{dt} = rV - \alpha VP \quad \text{Equation 6.1}$$

The term after the minus sign says that losses to predation are proportional to

the *product* of predator and victim numbers. This is equivalent to a chemical reaction in which the reaction rates are proportional to the concentrations of molecules. If predators and victims move randomly through the environment, then their encounter rate is proportional to the product of their abundances. Note that we have now started recycling symbols: α is *not* the competition coefficient from Chapter 5! Instead, here α measures the **attack rate** of the predators on their prey. The larger α is, the more the victim population is depressed by the addition of a single predator. A filter-feeding baleen whale would have a large α , because a single whale can consume millions of plankton. In contrast, a web-building spider might have a fairly low α if the addition of a single web does not greatly depress prey populations. The product αV is the **functional response** of the predator—the rate of victim capture by a predator as a function of victim abundance (Solomon 1949). Later in this chapter, we will derive some more complicated expressions for the functional response, but for now we will represent it as a simple product of victim abundance and attack rate. Before we explore the solutions to the equation for victim growth, we will develop an analogous equation to describe the growth of the predator population.

MODELING PREDATOR POPULATION GROWTH

The growth of the predator population is affected by the numbers of both predators and victims:

$$\frac{dP}{dt} = g(P, V) \quad \text{Expression 6.3}$$

We use the symbol g for this function to distinguish it from the function f that is used for the victim population in Expression 6.1.

The predator we are modeling is an extreme specialist. It will feed only on the victim population and has no alternative source of prey. Consequently, if the victim population is absent, the predator population declines exponentially:

$$\frac{dP}{dt} = -qP \quad \text{Expression 6.4}$$

where q is the death rate. (This is equivalent to the death rate d from the exponential growth model described in Chapter 1; we have changed symbols here to avoid confusion.)

Positive growth occurs only when the victim population is present:

$$\frac{dP}{dt} = \beta VP - qP \quad \text{Equation 6.2}$$

Here βVP indicates random encounters of predators and victims. β is a measure of **conversion efficiency**—the rate at which the predator converts victim biomass into new predator offspring. We expect β to be high when a single prey item is particularly valuable, such as a moose that is captured by wolves. In contrast, β will be low when a single prey item does not contribute much to growth of the predator population; think of a single seed consumed by a granivorous bird. βV reflects the **numerical response** of the predator population—the growth rate of the predator population as a function of victim abundance.

EQUILIBRIUM SOLUTIONS

To find the equilibrium for the victim and predator populations, we set each equation equal to zero and solve for population size. Beginning with Equation 6.1:

$$0 = rV - \alpha VP \quad \text{Expression 6.5}$$

$$rV = \alpha VP \quad \text{Expression 6.6}$$

$$r = \alpha P \quad \text{Expression 6.7}$$

$$\hat{P} = \frac{r}{\alpha} \quad \text{Equation 6.8}$$

Although we tried to solve for the victim equilibrium, the solution is in terms of P , the predator population! The important result is that a specific number of predators (\hat{P}) will maintain the victim population at zero growth. This predator level is determined by the ratio of the growth rate of the victims (r) to the attack rate of the predators (α). The faster the growth rate of the victim population, the more predators are needed to keep the victim population in check. Conversely, the higher the attack rate, the fewer predators needed for control.

Solving the equilibrium for the predators (Equation 6.2) yields an expression in terms of the victim population size:

$$0 = \beta VP - qP \quad \text{Expression 6.9}$$

$$\beta VP = qP \quad \text{Expression 6.10}$$

$$\beta V = q$$

$$\hat{V} = \frac{q}{\beta} \quad \text{Equation 6.4}$$

Thus, the predator population is controlled by a fixed number of victims (\hat{V}). The greater the death rate of the predators (q), the more victims needed to keep the predator population from declining. Conversely, the greater the conversion efficiency of predators (β), the fewer victims needed to maintain the predators at equilibrium. Because Equations 6.3 and 6.4 give the conditions for zero growth, they represent the victim and predator isoclines, respectively.

GRAPHICAL SOLUTIONS TO THE LOTKA–VOLTERRA PREDATION MODEL

As in our analysis of the competition model (Chapter 5), we can plot the isoclines for each species in state space to evaluate the joint equilibrium. Plotting the victim population on the x axis yields a horizontal victim isocline, representing the number of predators needed to hold the victim population in check. If the predator population is less than this number, the victim population can increase in size, represented by horizontal arrows pointing to the right. Conversely, if the predator population is above the victim isocline, the victim population declines, represented by horizontal arrows pointing to the left (Figure 6.1).

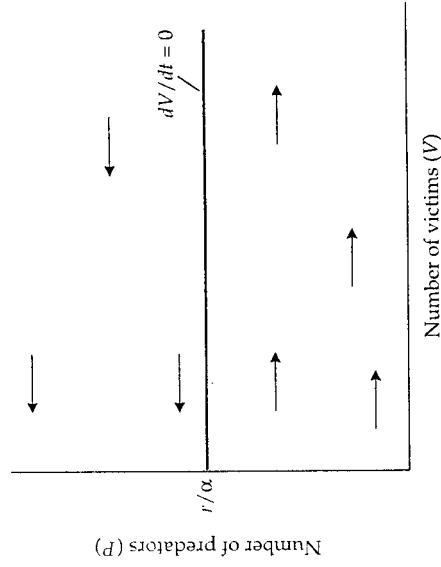


Figure 6.1 The victim isocline in state space. The Lotka–Volterra predation model predicts a critical number of predators (r/α) that controls the victim population. If there are fewer predators than this, the victim population increases (right-pointing arrows). If there are more predators, the victim population decreases (left-pointing arrows). The victim population has zero growth when $P = r/\alpha$.

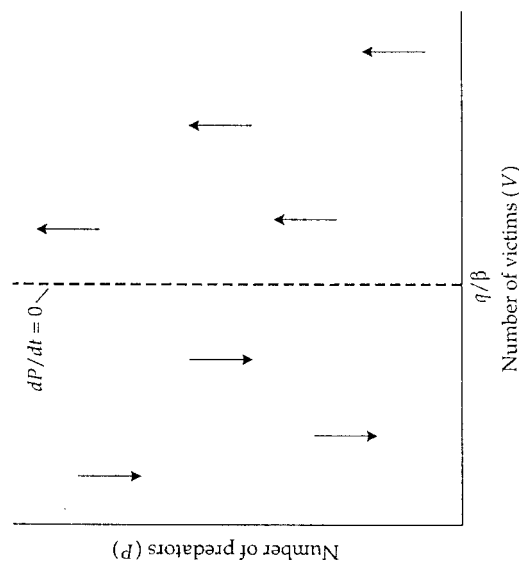


Figure 6.2 The predator isocline in state space. The Lotka–Volterra predation model predicts a critical number of victims (q/β) that controls the predator population. If there are fewer victims than this, the predator population decreases (downward-pointing arrows). If there are more victims, the predator population increases (upward-pointing arrows). The predator population has zero growth when $V = q/\beta$.

Similar reasoning applies to the analysis of the predator isocline. This isocline is a vertical line, representing a critical size of the victim population. To the left of the isocline, there are not enough victims to support the predator population. In this region of the state space graph, the predator population declines, represented by downward-pointing vertical arrows. To the right of the isocline, there is an excess supply of victims, and the predator population increases, represented by upward-pointing vertical arrows (Figure 6.2).

In our analysis of competition models, there were four ways that the pair of isoclines could be placed in the state-space graph. For the predation model, there is only one possible pattern: the isoclines cross at 90° angles (Figure 6.3). However, we will see that the dynamics are more complex than in the competition model.

The predator and victim isoclines divide the state space into four regions. Beginning in the upper right-hand corner, we are in a region where both predator and victim are abundant. Because we are above the predator isocline, prey are abundant enough for the predator to increase. However, we are to the right of the vertical victim isocline. Consequently, there are too many predators, and the victim population declines. The vector of net move-

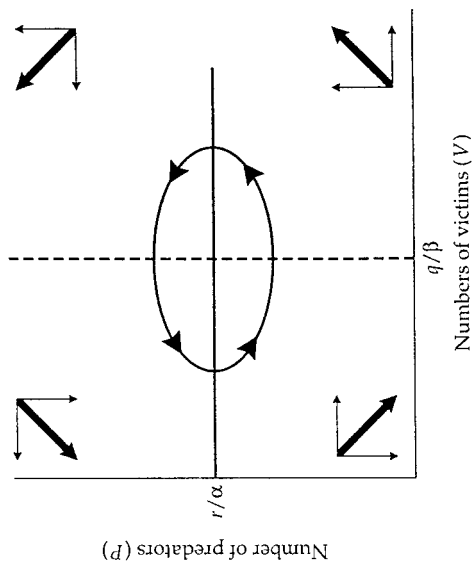


Figure 6.3 The dynamics of predator and victim populations in the Lotka–Volterra model. The vectors indicate the trajectories of the populations in the different regions of the state space. The populations trace a counterclockwise path that approximates an ellipse.

ment points towards the upper left-hand quadrant. As the victim abundance continues to decline, we cross the vertical isocline into the upper left-hand region of state space.

Now the victim population has declined to the point where the predator population can no longer increase. Both predator and victim populations decrease, and the vector moves into the lower left-hand quadrant. In this region, the predator population continues to decline, but the victim population starts to increase again. The net movement is down and to the right, taking the trajectory into the fourth quadrant. Here, the victim population continues to grow, but it has now become large enough for the predators to also increase. The system again moves back to the starting point, the upper right-hand quadrant.

Thus, the predator and victim populations trace an approximate ellipse in state space. Unless the predator and victim populations are precisely at the intersection of the isoclines, their trajectories will continue to move in this counterclockwise ellipse.

How does this ellipse translate into growth curves for the predator and victim populations? Both populations cycle periodically, increasing and decreasing smoothly from minimum to maximum. The ellipse indicates that the peak of the predator population occurs when the victim population is at

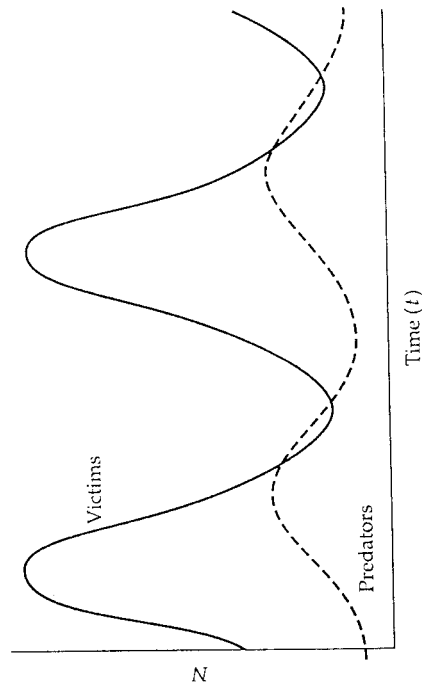


Figure 6.4 Cycles of predator and prey in the Lotka–Volterra model. Each population cycles with an amplitude that is determined by the starting population sizes and a period of approximately $2\pi/\sqrt{rq}$. The predator and victim populations are displaced by one-quarter of a cycle, so that the predator population peaks when the victim population has declined to half its maximum, and vice versa.

its midpoint, and vice versa. In other words, the peaks of the predator and victim populations are displaced by one-quarter of a cycle (Figure 6.4).

What would happen if the predator and victim populations had a different starting point in the state space? This would correspond to different initial densities of predator and victim, and a new ellipse would be traced. Both populations would again exhibit cycles, although with a different amplitude. The closer the ellipse is to the isocline intersection, the smaller the amplitude of the predator and victim cycles. Thus, the Lotka–Volterra cycles are **neutrally stable**—the amplitudes are determined solely by the initial conditions.

There are only two exceptions to population cycling: (1) if the victim and predator populations are precisely at the isocline intersection, they will not change, although if they are displaced any distance from this point, they will begin cycling; or (2) if the starting point of the ellipse is too extreme, it will hit one of the axes of the state space graph. In this case, the amplitude of the cycle is so large that either predator or the victim population will crash. Although the amplitude of the cycle is determined by the initial population sizes, the period of the cycle (C) is approximately

$$C \approx \frac{2\pi}{\sqrt{rq}} \quad \text{Equation 6.5}$$

Thus, the greater the prey growth rate (r) and/or the predator death rate (q), the faster the populations cycle between high and low values. The essential feature of the Lotka–Volterra predation model is that the predator and victim populations cycle because they reciprocally control one another’s growth.

Model Assumptions

The Lotka–Volterra predation model carries with it the standard assumptions of no immigration, no age or genetic structure, and no time lags. In addition, the model makes the following assumptions about predators, victims, and the environment:

- ✓ **Growth of the victim population is limited only by predation.** Equation 6.1 shows that the victim population grows exponentially in the absence of the predator.
- ✓ **The predator is a specialist that can persist only if the victim population is present.** Equation 6.2 shows that the predator population will starve in the absence of the victim.
- ✓ **Individual predators can consume an infinite number of victims.** Because the horizontal victim isocline ($dV/dt = 0$) implies a constant number of predators, each predator must be able to increase its consumption as the victim population increases in size. An infinite capacity for consuming prey also implies that there is no interference or cooperation among predators.
- ✓ **Predator and victim encounter one another randomly in an homogeneous environment.** The interaction terms (αVP and βVP) imply that predators and victims move randomly through the environment, and that victims do not have spatial or temporal refuges for avoiding predators.

Model Variations

The unique prediction of the Lotka–Volterra predation model is cycles of predator and victim populations. However, these cycles are very sensitive to the restrictive assumptions and linear isoclines of the model. In the following sections, we will incorporate more realistic assumptions about predators and victims that bend the isoclines and produce other dynamics. We will not solve the equations for these more complex models, although we will analyze their behavior with state-space graphs.

INCORPORATING A VICTIM CARRYING CAPACITY

The victim isocline tells us how many predators are needed to hold the victim population in check. Notice that as we move to the right in the state-space graph (Figure 6.1), the same number of predators will control the victim population. This is not realistic. We expect that as the victim population becomes more crowded, it will start to be limited by other resources that have nothing to do with predators. We can modify the victim isocline to incorporate a victim carrying capacity by including another term with a new constant c :

$$\frac{dV}{dt} = rV - \alpha VP - cV^2 \quad \text{Equation 6.6}$$

Now the growth of the victim population is decreased by the presence of predators (αVP) and by its own density (cV^2). When graphed in the state space, this new isocline is a straight line with a negative slope, in contrast to the horizontal victim isocline of the simple Lotka–Volterra model. The new isocline crosses the x axis at r/c , which is the maximum population size achieved by the victims when no predators are present.

How does the interaction of predator and prey change when the victim population is limited by its own carrying capacity? Figure 6.5 shows that the trajectory for the predator and prey populations spirals inwards to the equilibrium intersection. This is a stable equilibrium point, and the equilibrium

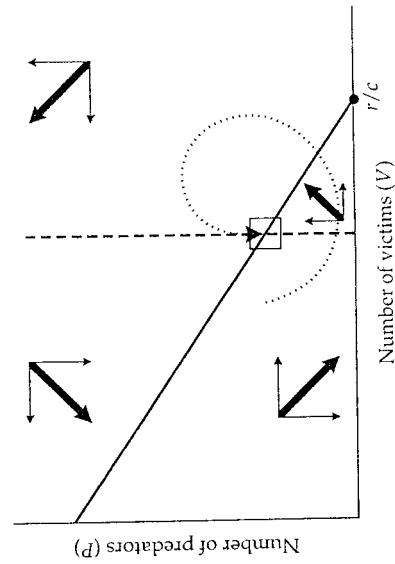


Figure 6.5 The effect of a victim carrying capacity on the victim isocline. The victim isocline slopes downward with a carrying capacity incorporated. The intersection with the vertical predator isocline forms a stable equilibrium point.

abundance for the victim population is lower when the predators are present than when they are absent. The presence of a victim carrying capacity stabilizes the predator–prey interaction. This makes intuitive sense—if the victims are limited by factors other than their predators, then there would be less of a tendency for the two populations to cycle.

MODIFYING THE FUNCTIONAL RESPONSE

One of the most unrealistic assumptions of the Lotka–Volterra model is that individual predators can always increase their prey consumption as the victim population increases. This type of foraging is illustrated in a graph of the functional response (Figure 6.6), which plots the rate of prey captured per individual predator (n/t) as a function of prey density (V). The Lotka–Volterra model assumes a “Type I” functional response, in which the predator consumes more as prey density increases. The slope of this curve is α , the attack rate.

A Type I functional response is unrealistic for two reasons. First, predators will eventually become satiated (stuffed) and stop feeding. Second, even in the absence of satiation, predators are limited by the **handling time** needed to catch and consume each prey item. Consequently, there is a limit to the rate at which individual predators can process prey.

We can construct a more realistic “Type II” functional response by model-

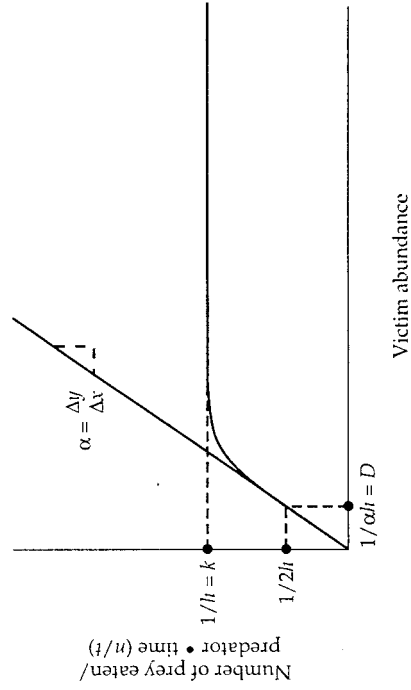


Figure 6.6 The functional response of predators is the feeding rate per predator as a function of prey abundance. The shape of these curves depends on the prey attack rate (α), the maximum predator feeding rate (k), and the prey abundance for which the predator feeding rate is half of the maximum (D).

ing the components that contribute to the rate at which individual predators capture victims (n/t) (Royama 1971). The total amount of time that a predator spends feeding (t) is the time spent searching for the prey (t_s), plus the time spent "handling" or consuming the prey (t_h):

$$t = t_s + t_h \quad \text{Expression 6.11}$$

If we let n equal the number of prey captured in time t and h equal the handling time per prey item, the total handling time is:

$$t_h = h n \quad \text{Expression 6.12}$$

Similarly, we can derive an expression for the search time. The total number of prey captured by a predator (n) is simply the product of the victim abundance (V), the attack rate (α), and the total search time (t_s):

$$n = V \alpha t_s \quad \text{Expression 6.13}$$

We can rearrange this to give us an expression for the search time:

$$t_s = \frac{n}{\alpha V} \quad \text{Expression 6.14}$$

Substituting Expressions 6.12 and 6.14 into 6.11, we have:

$$t = \frac{n}{\alpha V} + h n \quad \text{Expression 6.15}$$

Multiplying the second term by $(\alpha V / \alpha V)$ gives:

$$t = \frac{n}{\alpha V} + \frac{\alpha V h n}{\alpha V} \quad \text{Expression 6.16}$$

$$t = n \left(\frac{1 + \alpha V h}{\alpha V} \right) \quad \text{Expression 6.17}$$

Finally, this can be rearranged to give us an expression for the feeding rate (n/t):

$$n/t = \frac{\alpha V}{1 + \alpha V h} \quad \text{Equation 6.7}$$

Equation 6.7 describes the feeding rate per predator as a function of the attack rate, the victim abundance, and the handling time. Note that if the victim abundance is very low, the term $\alpha V h$ in the denominator is small, so the feeding rate is close to αV , as in the simple Lotka-Volterra model. But as the victim abundance increases, the feeding rate approaches a saturation value of

$1/h$. This value represents the maximum feeding rate that the predator can achieve because of the constraints of handling time. Equation 6.7 is sometimes referred to as the "disc equation" because it fits data from an experiment in which human subjects were blindfolded and required to find and pick up small discs of sandpaper scattered on a flat surface (Holling 1959).

We can simplify Equation 6.7 somewhat by letting $k = 1/h$, the maximum feeding rate. We can also define the constant D as $1/\alpha h$. This value turns out to be the "half-saturation constant," which is the abundance of prey at which the feeding rate is half-maximal. If we first multiply the numerator and denominator of Equation 6.7 by $1/\alpha h$, we have:

$$n/t = \frac{\frac{\alpha V}{\alpha h}}{1 + \frac{\alpha V h}{\alpha h}} \quad \text{Expression 6.18}$$

Substituting in the two new constants k and D yields:

$$n/t = \frac{kV}{D + V} \quad \text{Equation 6.8}$$

This Type II functional response increases to a maximum and constant rate of prey consumption per predator (k). The half-saturation constant (D) controls the rate of increase to this maximum. This equation is identical to the Michaelis-Menten equation of enzyme kinetics (Real 1977).

Finally, a Type III functional response can be described by:

$$n/t = \frac{kV^2}{D^2 + V^2} \quad \text{Equation 6.9}$$

For a Type III functional response, the feeding rate also reaches an asymptote at k , but the curve has a sigmoid shape, similar to the logistic curve (see Chapter 2). Consequently, the feeding rate is accelerated at low prey density, but decreases at high prey density as the asymptote is reached (Figure 6.7). This functional response can occur if predators switch to prey items that become more common, if they develop a search image that increases capture efficiency as victim abundance increases, or if there are fixed and variable costs to foraging (Holling 1959, Mitchell and Brown 1990).

The functional response has important consequences for the ability of predators to control victim populations. Figure 6.8 shows the proportion of the prey population that is consumed by an individual predator as victim density increases. For the Type I response of the simple Lotka-Volterra

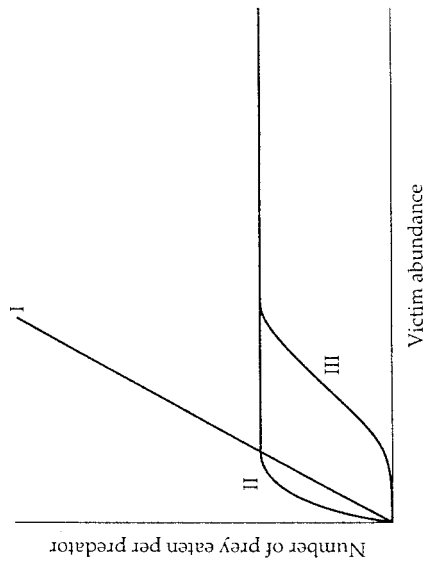


Figure 6.7 Type I, Type II, and Type III functional responses.

model, this proportion remains a constant, because each predator increases its individual feeding as victim abundance increases. For the Type II response, the proportion decreases steadily because each predator can only process prey at a maximum rate k . The Type III response shows an initial increase because of the accelerated feeding rate, but this quickly decreases and converges on the Type II curve. These curves show that, at high victim abundance, predators with a Type II or Type III response may not be able to

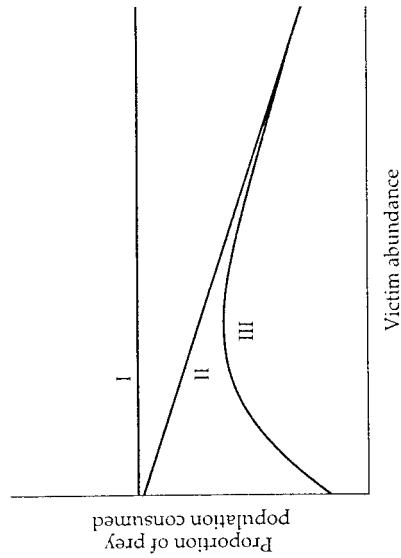


Figure 6.8 The proportion of the prey population consumed by an individual predator as a function of victim density.

effectively control victim populations. Control is possible with the Type III response, but only at relatively low victim abundance. In contrast, the Type I functional response ensures effective control over all levels of victim abundance.

Incorporating a Type II or Type III functional response into the equation for the victim growth rate gives:

$$\frac{dV}{dt} = rV - \left(\frac{kV}{V + D} \right) P \quad \text{Equation 6.10}$$

$$\frac{dV}{dt} = rV - \left(\frac{kV^2}{V^2 + D^2} \right) P \quad \text{Equation 6.11}$$

Figure 6.9 shows that the isoclines for these growth equations increase in the state space, with an upward swing at low victim abundance for the Type III functional response. Because each predator is limited by a maximum consumption rate, more predators are required to hold large victim populations at zero growth. When these increasing victim isoclines intersect a vertical predator isocline, the equilibrium is unstable, and the predator and victim will not coexist.

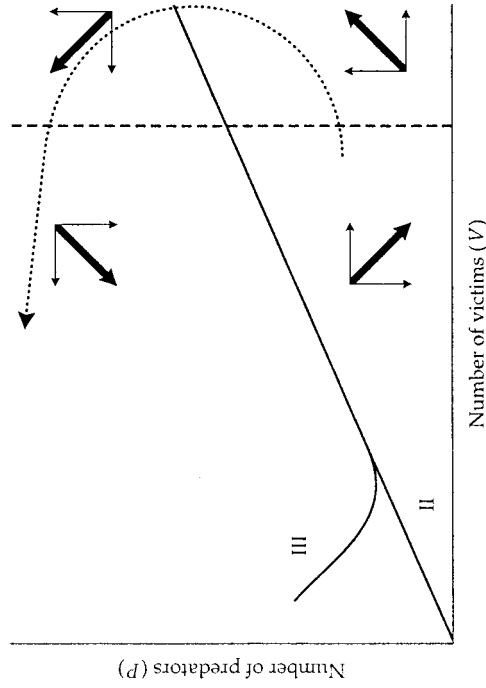


Figure 6.9 Victim isoclines incorporating a Type II or a Type III functional response. The intersection of an increasing victim isocline with a vertical predator isocline generates an unstable equilibrium point.

THE PARADOX OF ENRICHMENT

The victim isocline may also increase because of an Allee effect (see Chapter 2) for the victim population. If larger victim populations are more effective at reproducing, obtaining food, or defending themselves from predators, more predators would be needed to control the prey population. Because of a victim carrying capacity, predator functional response, Allee effects, and a variety of other reasons, the victim isocline may have a hump in the middle (Rosenzweig and MacArthur 1963), turning downward at both low and high prey densities.

How does this more realistic victim isocline affect predator–prey dynamics? The answer depends on precisely where the vertical predator isocline intersects the victim isocline. If the intersection is at the peak of the victim isocline, the predator and victim populations will cycle as in the simple Lotka–Volterra model (Figure 6.10a). However, if the predator isocline crosses to the right of the hump, the predator and victim populations converge on a stable equilibrium point, without population cycles (Figure 6.10b). In this case, the predator is relatively inefficient. Thus, from Equation 6.4, the predator population has a relatively high death rate (q) and/or a low conversion efficiency (β). In contrast, if the predator is relatively efficient (low q and/or high β), the isoclines intersect to the left of the hump. In this case, the equilibrium is unstable. The predator population will overexploit the victim population, drive it to extinction, and then starve (Figure 6.10c).

This instability due to a relatively efficient predator has been termed “the paradox of enrichment” (Rosenzweig 1971). The paradox may explain why some artificially enriched agricultural systems are vulnerable to pest outbreaks. Suppose the “victim” population is a crop plant that coexists in a stable equilibrium with a “predator” population of an herbivorous insect. If the productivity of the crop plant is increased with fertilizers, the victim isocline may shift to the right to a new, higher carrying capacity (Figure 6.11). If the predator isocline remains stationary, the dynamics may shift from a stable equilibrium to an unstable outbreak of the “pest.” This paradox depends on the unrealistic assumption of a strictly vertical predator isocline. More realistic predator isoclines, described later in this chapter, may enhance stability of predator and prey over a wider range of victim abundances (Berryman 1992).

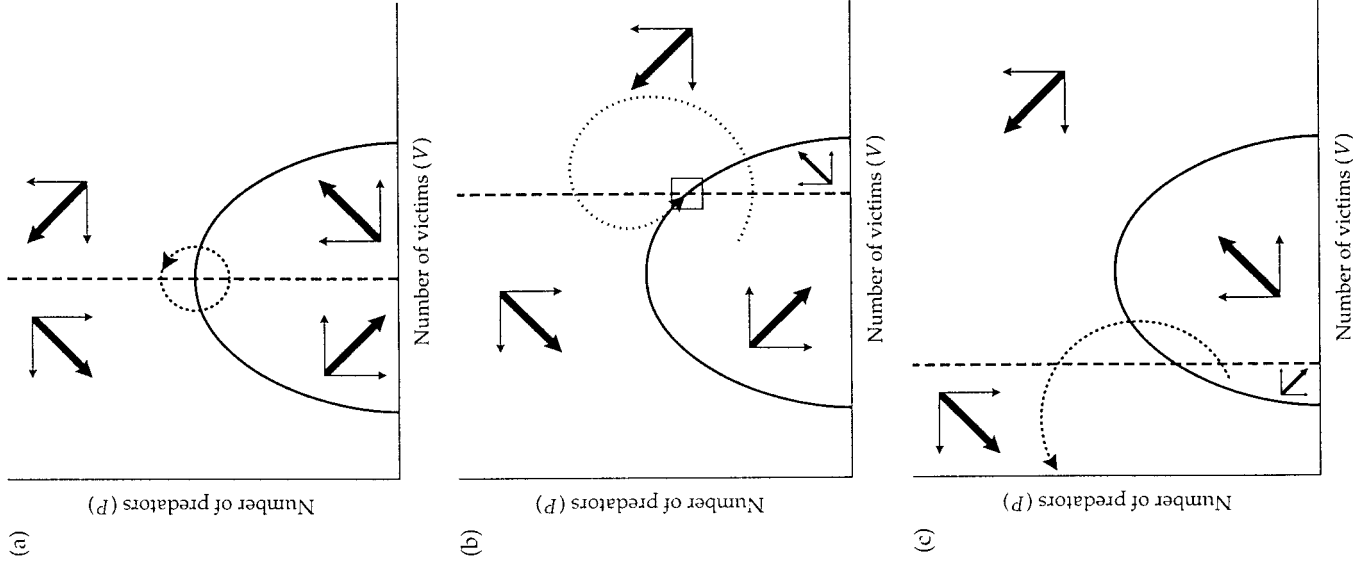


Figure 6.10 (a) Predator–prey cycles with a humped prey isocline. As in the Lotka–Volterra model, the predator and victim populations cycle as long as the predator and prey isoclines are perpendicular where they intersect. (b) If the predator is relatively inefficient, the predator isocline intersects to the right of the peak of the victim isocline. In this case, predator and victim coexist in a stable equilibrium. (c) If the predator is relatively efficient, the predator isocline intersects to the left of the peak of the victim isocline. In this case, the predator overexploits the prey population, drives it to extinction, and starves.

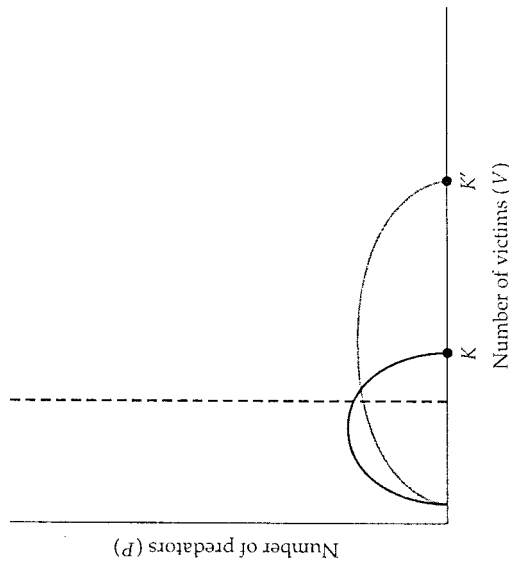


Figure 6.11 The paradox of enrichment. If the victim population has its carrying capacity enhanced from K to K' , the system moves from a stable equilibrium to over-exploitation by the predator.

INCORPORATING OTHER FACTORS IN THE VICTIM ISOCLINE

The victim isocline may also turn upward at low victim densities, generating different population dynamics. There are three reasons for an upturn of the victim isocline. First, the isocline will turn up if there are a fixed number of victim refuges that are secure from predators. For example, fish that live in rock crevices and songbirds that establish territories in areas protected by cover have spatial refuges from predation. In this case, no matter how large the predator population gets, the victim population can always persist at low density in the refuges. Second, the victim isocline may turn upwards if there is a constant number of victim immigrants that arrive each generation. With immigration, the victim population always has the potential to increase at low numbers. Finally, the isocline may turn upward at low victim densities because of a Type III functional response, as explained earlier.

This upward turn of the victim isocline has the potential to stabilize predator-prey interactions. For example, suppose that the predator is relatively efficient, but there is a victim carrying capacity and there are refuges from predation for the victim population (Figure 6.12). In this case, the predators quickly consume all the available victims, as in the destabilized case (Figure 6.10c). But once all the victims outside the shelters are consumed, the predator population begins to starve, and its abundance declines. When the pred-

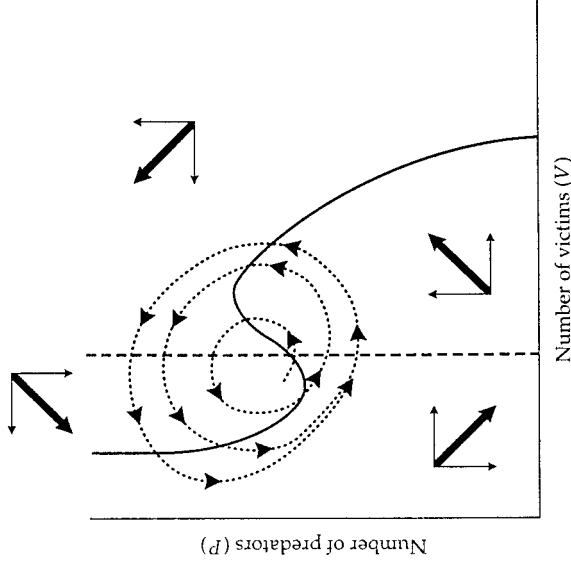
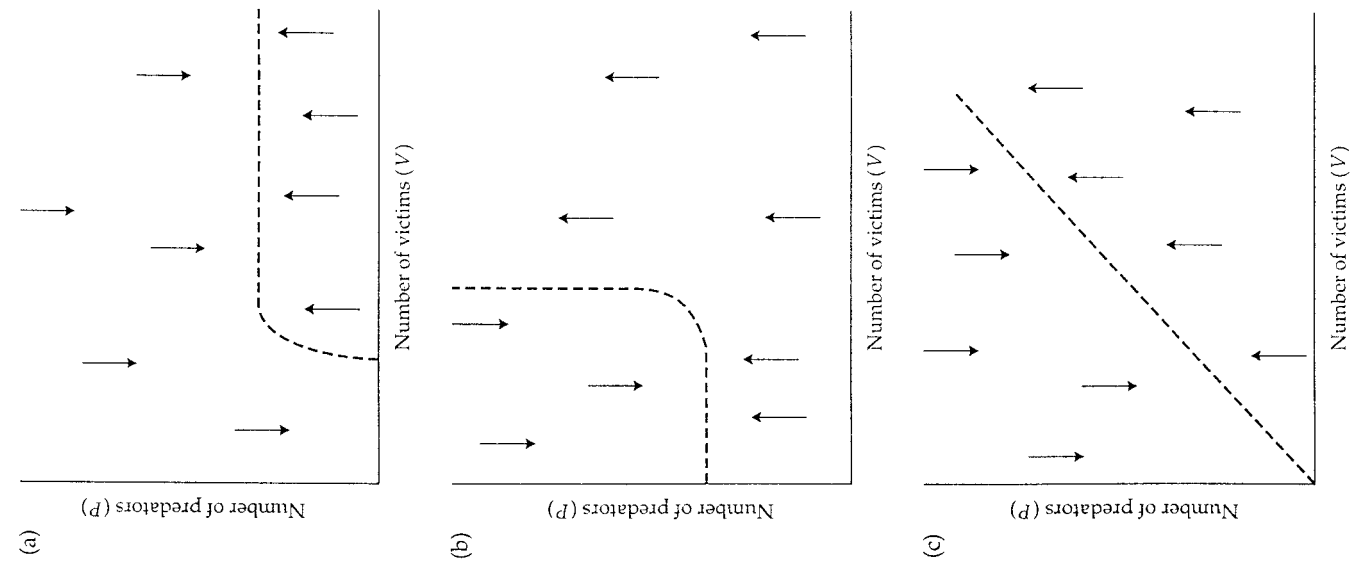


Figure 6.12 Cycling of predator and victim populations because of refuges. If there are spatial refuges from predation, the victim isocline becomes vertical at low densities. In this case, the efficient predator cannot overexploit its prey, and begins to starve once all the victims outside of the refuges have been consumed. After the predator population declines below a certain point, the victim population begins to increase again, repeating the cycle.

tor population declines below a certain point, the victim population in the refuges starts to increase, and the cycle repeats itself. In contrast to the simple Lotka-Volterra model, these cycles are stable, because no matter what the starting density, the predator population will eventually consume all the victims not in refuges, and the cycle will repeat.

MODIFYING THE PREDATOR ISOCLINE

We can also modify the vertical predator isocline to make it more realistic. These modifications involve changes in the numerical response of Equation 6.2, which will be described qualitatively. For example, the Lotka-Volterra predation model assumes that the predator population can always increase in size if there is an excess of prey available. It is more realistic to suppose that the predator population has its own carrying capacity, so that its growth is limited by other factors. A carrying capacity for the predator bends the predator isocline to the right (Figure 6.13a).



◀ **Figure 6.13** (a) Effects of carrying capacity on the predator isocline. If the predator population is limited by factors other than prey density, the predator isocline bends to the right. No matter how large the victim population, the predator population becomes limited when it reaches its own carrying capacity. (b) Effects of the availability of alternative prey on the predator isocline. If the predator is not a specialist on the victim, the predator population may be able to increase even when the victim density declines to zero. (c) Effects of victim density on the predator isocline. If the size of the victim population acts as a carrying capacity for the predators, the predator isocline increases with increasing victim density.

Another unrealistic assumption of the Lotka–Volterra model is that the predator is a specialist on the victim. Suppose instead that the predator has alternative prey sources. Then, when the victim population becomes less abundant, the predator population can continue to increase by feeding on other prey items. This will tip the predator isocline towards the horizontal at low prey abundance (Figure 6.13b). Thus, with alternative prey and a predator carrying capacity, the predator isocline can shift from vertical to horizontal. As we noted earlier, the availability of other prey may shift the victim isocline as well.

As an intermediate case, suppose that the size of the victim population determines the size of the predator population. In other words, the victim population functions as a “carrying capacity” for the predators. In this case, the predator isocline will be a line with a positive slope, intermediate between the vertical isocline of the Lotka–Volterra model and the horizontal isocline of a predator with an independent carrying capacity and alternative prey (Figure 6.13c).

How will these alterations of the predator isocline affect the stability of the model? As a general rule, anything that rotates either the predator or the victim isocline in a *clockwise* direction will tend to stabilize the interaction, whereas anything that rotates the isoclines *counterclockwise* will be destabilizing. These rotations can be compared to the neutral stability of a horizontal victim isocline and a vertical predator isocline in the Lotka–Volterra model (Figure 6.14). For example, giving the victim population a carrying capacity rotates the victim isocline clockwise, leading to a stable equilibrium on the right side of the hump (Figure 6.10b). But adding predator satiation rotates the victim isocline counterclockwise at low abundances, leading to an unstable equilibrium on the left side of the hump (Figure 6.10c). Rotating the predator isocline also increases the stability of the interaction. Whereas a vertical predator isocline generates population cycles in a neutral equilibrium, an increasing predator isocline generates damped cycles, and a horizontal predator isocline generates a stable equilibrium point (Figure 6.15).

These geometrical rules make intuitive biological sense. The more independent the predator and prey are of one another, the more stable the inter-

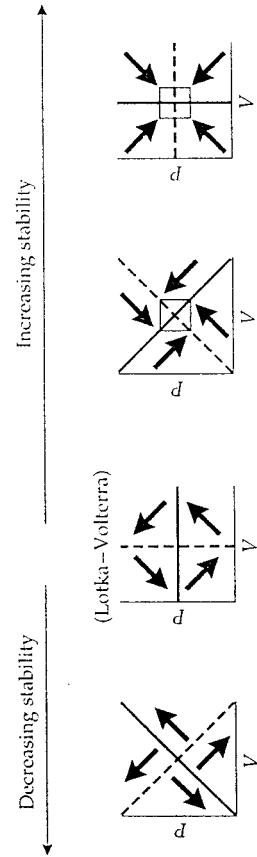


Figure 6.14 Effects of rotating the predator and victim isoclines on the stability of the equilibrium. Relative to the neutral equilibrium of the Lotka–Volterra model, clockwise rotations of the isoclines lead to more stable equilibria; counterclockwise rotations lead to less stable equilibria.

action. For example, suppose the victim isocline is vertical and the predator isocline is horizontal (Figure 6.14). In this case, the carrying capacities of the predator and victim are completely independent of one another, and both species coexist in a very stable equilibrium. Cycles are difficult to generate with simple predator–victim models, and require a special dependence of

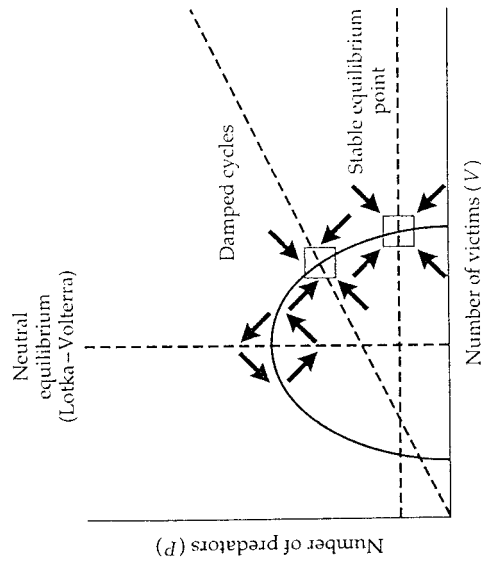


Figure 6.15 Effects of clockwise rotation of the predator isocline. As the predator isocline is rotated, the dynamics change from cycles with a neutral equilibrium, to damped cycles, to a stable equilibrium point. Biologically, the three predator isoclines correspond to a predator that is a complete specialist on the victim, to one whose carrying capacity is proportional to victim abundance, to one whose carrying capacity is independent of victim abundance.

predator and victim populations upon each other, as in the original Lotka–Volterra model.

Empirical Examples

POPULATION CYCLES OF HARE AND LYNX

The basic prediction of the Lotka–Volterra model is the regular cycling of predator and prey populations. The most famous example of this cycling is the case of the Canada lynx (*Lynx canadensis*) and its principal prey, the snowshoe hare (*Lepus americanus*). The ecologist Charles Elton analyzed fur-trapping records from the Hudson’s Bay Company in Canada and found a long-term record of population cycles (Elton and Nicholson 1942). The major source of hare mortality is predation (Smith et al. 1988), and the hare population cycles with a peak abundance approximately every 10 years (Figure 6.16). The lynx population is highly synchronized with the hare and usually peaks one or two years later. These are not the only prey and predator species that cycle in the boreal north. Populations of muskrat, ruffed grouse, and ptarmigan exhibit 9 to 10 year cycles, whereas smaller herbivores such as voles and lemmings cycle with peaks every 4 years. Predators such as foxes, mink, owls, and martens also cycle synchronously with their prey.

What is the explanation for the striking hare–lynx cycle? An early suggestion that the hare cycles were correlated with sunspot activity was dismissed because sunspot activity peaks every 11 years, whereas the hare cycle is approximately 10 years in length (Moran 1949). For many years, the hare–lynx cycle was the classic textbook example of predator and prey populations that cycled according to the Lotka–Volterra model. More recently,

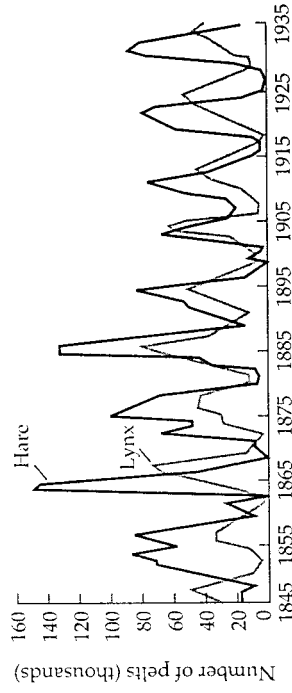


Figure 6.16 One-hundred-year record of population cycles of the snowshoe hare (*Lepus americanus*) and the Canada lynx (*Lynx canadensis*), based on pelt records of the Hudson’s Bay Company in Canada.

ratio-dependent predator-prey models have been applied to the hare-lynx data (Akçakaya 1992). These models are based on the assumption that the functional response of the predators depends not simply on prey density (V), but on the ratio of prey to predator density (V/P).

Unfortunately, two additional pieces of data complicate the story. First, the hare-lynx cycles seem to be broadly synchronized within a year or two over wide areas of North America (Smith 1983). If the predator-prey models were correct, we would expect cycles of different amplitude and period to arise in different local populations. Second, there are places on the coast of British Columbia and on Anticosti Island, Quebec, where there are no lynx, but the hare population cycles nonetheless!

These results suggest that the hare and lynx do not reciprocally influence each other. Instead, the lynx population is probably "tracking" the hare cycle. The hare cycle seems to be caused, in part, by interactions with its food supply. Heavily grazed grasses produce shoots with high levels of toxins that make them less palatable to hares (Keith 1983). This chemical protection persists for two or three years after grazing, further contributing to the hare decline. A single-species logistic model with a time lag (see Chapter 2) would qualitatively describe this sort of cycle. However, as most hares die of predation, not starvation, food quality probably contributes to their susceptibility to predation.

Finally, recent evidence again suggests that sunspots may indeed contribute to the cycles. Sunspot activity is associated with hare browse marks in tree rings and with periods of low snow accumulation (Sinclair et al. 1993). Sunspot activity may serve as a phase-locking mechanism through indirect influences on climate and plant growth. These broad climatic effects could be responsible for the synchrony of hare-lynx cycles over large areas of Canada and Alaska. Whatever the ultimate explanation, it is clear that the hare-lynx cycle is more complex than suggested by the superficial match of the data to the simple predictions of the Lotka-Volterra model.

POPULATION CYCLES OF RED GROUSE

Interactions between hosts and parasites represent a special kind of "predation" in which the life history of the predator is intimately tied to that of its host. Whereas most predators benefit from rapidly killing and consuming their prey, a parasite must keep its host alive at least long enough to successfully reproduce and infect a new host. To understand the population dynamics of hosts and parasites, we must therefore model the dynamics of the egg or larval stages, as well as those of the host and the adult parasite (Anderson and May 1978).

A nice example illustrating these complexities is the case of the parasitic

nematode *Trichostrongylus tenuis*, which infects red grouse (*Lagopus lagopus scoticus*) on the moors of England and Scotland. Adult worms inhabit the large caeca of red grouse, and their eggs pass out of the host with the feces. If the environment is warm and moist, the eggs hatch and develop into a larval stage. The larval nematode moves to the growing tips of heather plants, where it is consumed by a new host, and the life cycle repeats itself. A single bird may be host to over 10,000 worms. As the intensity of the parasite infection increases, winter mortality, egg mortality, and chick losses all increase (Figure 6.17). Thus, *T. tenuis* has the potential to regulate the population growth of red grouse.

Because red grouse are an important game bird in England and Scotland, there are detailed records on its population dynamics and the prevalence of parasite infection (Hudson et al. 1992). Figure 6.18 shows a 14-year record of host and parasite populations at Gunnerside, North Yorkshire. The red grouse population cycles, with a period of approximately 5 years. Parasite burden (number of worms per host) also cycles, with peaks occurring near the low point of the red grouse cycle.

Even a relatively simple model of the grouse-nematode interaction requires a minimum of three differential equations: one for the host (H), one for the adult worms (P), and one for the free-living egg and larval stages (W ; Dobson and Hudson 1992). The growth of the host population can be modeled as:

$$\frac{dH}{dt} = (b - d - cH)H - (\alpha + \delta)P \quad \text{Expression 6.19}$$

The first term $[(b - d - cH)H]$ represents the growth of the red grouse population in the absence of the parasite. The constants b and d represent intrinsic birth and death rates, and cH is a density-dependent term. The first part of this equation is really a model of logistic growth, with a carrying capacity of $[(b - d)/c]$. A finite carrying capacity is realistic for the grouse population because the birds are territorial. The second part of the equation $[(\alpha + \delta)P]$ represents the losses due to parasites. α is the reduction in host population growth due to effects of the parasite on the survivorship of grouse, and δ is the reduction due to parasite effects on the reproduction of grouse. We distinguish between these two mechanisms because α and δ appear separately in other equations in the model.

Next, we write an equation for the growth rate of the free-living stages:

$$\frac{dW}{dt} = \lambda P - \gamma W - \beta WH \quad \text{Expression 6.20}$$

Here, λ is the per capita fecundity of the parasite in the host, γ is the death rate

of the egg and larval stages in the field, and βWH is the rate at which larvae are transmitted to new hosts. Note the similarity of this latter expression to the "random encounter" term in the Lotka-Volterra model (Equations 6.1 and 6.2). Finally, we can describe the dynamics of the adult worm population as:

$$\frac{dP}{dt} = \beta WH - (\mu + d + \alpha)P - \alpha \frac{P^2}{H} \left(\frac{k+1}{k} \right) \quad \text{Expression 6.21}$$

The first term (βWH) represents the increase in the adult worm population from transmission. This is equivalent to the loss component of the egg-larva population. The second term $[(\mu + d + \alpha)P]$ represents decreases in growth of the worm population due to parasite death (μ), intrinsic host mortality (d), and host mortality from parasitism (α). The final term, $[\alpha(P^2/H)][(k+1)/k]$, represents losses due to the spatial dispersion of the worms among hosts. The constant k describes the distribution of worms among hosts. The smaller k is, the more aggregated the worms are in a few hosts. Aggregation will tend to decrease the growth of the worm population as the few heavily infected hosts die and take their parasites with them! In contrast, if the worms are distributed randomly or evenly among hosts, the growth rate of the parasite population

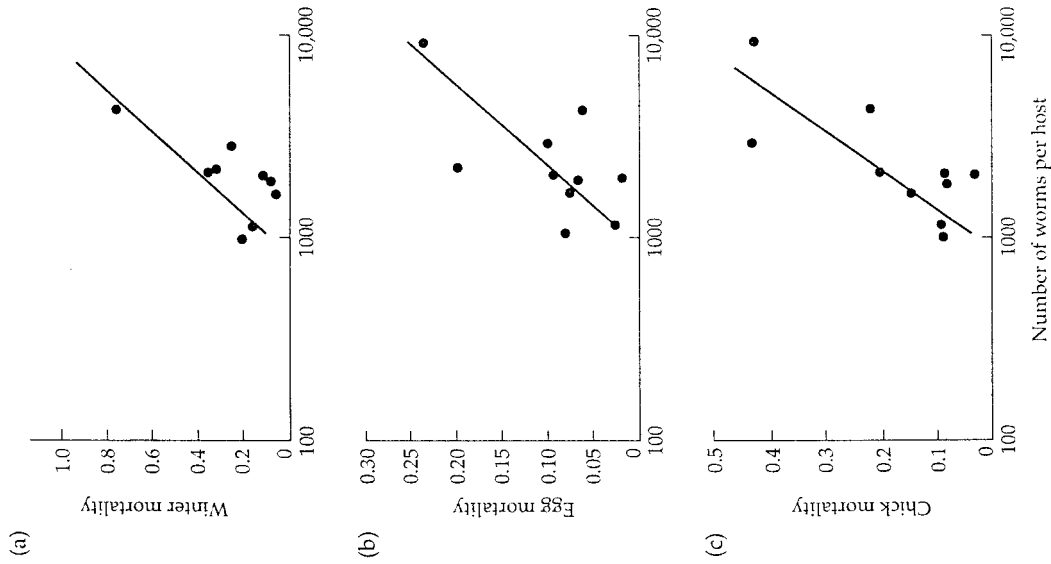


Figure 6.17 Effects of parasite load on (a) the winter mortality, (b) egg mortality, and (c) chick losses of red grouse (*Lagopus lagopus scoticus*). The x axis is the average parasite load (worms per host), and the y axis is the proportional mortality caused by each factor. Because the nematode *Trichostrongylus tenuis* reduces both the survivorship and reproduction of red grouse, it has the potential to regulate host numbers. (From Hudson et al. 1992.)

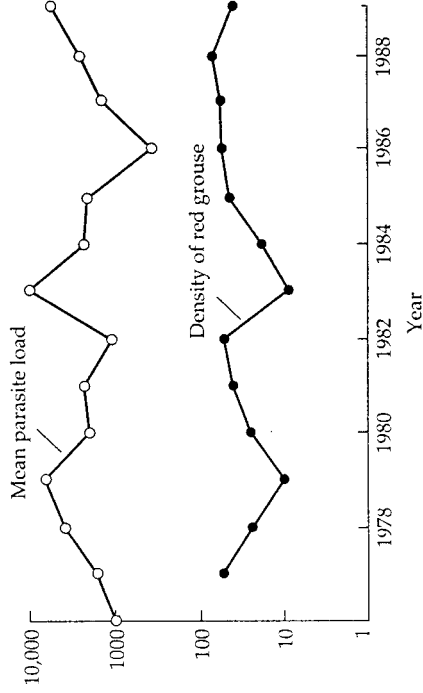


Figure 6.18 Changes in red grouse (*Lagopus lagopus scoticus*) density (breeding hens per square kilometer) and mean parasite load (worms per host) over 14 years at Gunnerside, North Yorkshire. Both the grouse and the nematode populations cycle with a period of approximately 5 years. Note the logarithmic scale on the y axis. (From Dobson et al. 1992; data from Hudson et al. 1992.)

lation is increased.

With ten different parameters in the model, there are a variety of possible outcomes. If parasite and host fecundity are not high enough, the parasite will go extinct, and the grouse population will rise to its carrying capacity. If the larval life of the parasite is relatively short, the grouse and parasite populations will coexist in a stable equilibrium. But if the larval and egg stages are fairly long-lived, the model generates stable cycles of host and parasite populations. Cycles in this model arise when $\alpha/\delta > k$. In other words, the ratio of parasite effects on survivorship (α) to parasite effects on reproduction (δ) must exceed the degree of parasite aggregation among hosts (k).

Field data were used to independently estimate the parameters of Expressions 6.19–6.21. The resulting model predicted population cycles with a period of approximately five years, which was observed at Gunnerside (Dobson and Hudson 1992). The model also provides insight into other grouse populations in England and Scotland. Not all grouse and nematode populations cycle, and these noncycling populations are in areas of relatively low rainfall (Hudson et al. 1985). Under these circumstances, the survival of eggs and larvae outside of the host is poor, and the model does not predict cycles.

The interaction of red grouse and its nematode parasite is one of the few well-documented cases of a predator and victim that cause each other's populations to cycle. However, the biology of the system is considerably more complex than that described by the simple Lotka–Volterra model. Models of host–parasite interactions have also been used to predict the dynamics of HIV (the AIDS virus) that infects humans.

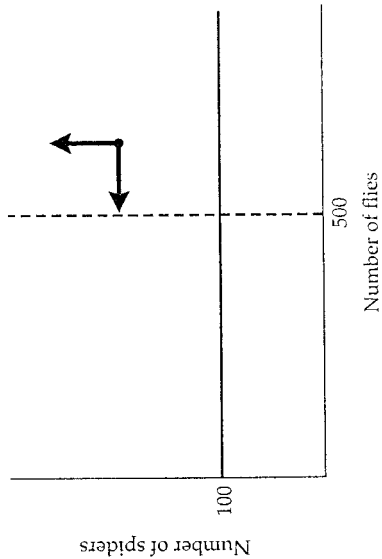
Problems

1. Suppose that spider and fly populations are governed by Lotka–Volterra dynamics, with the following coefficients: $r = 0.1$, $q = 0.5$, $\alpha = \beta = 0.001$. If the initial population sizes are 200 spiders and 600 flies, what are the short-term population dynamics predicted by the model?
2. Suppose that hawk and dove populations cycle with a peak every 10 years, and $r = 0.5$. If q is doubled in size, what happens to the period of the cycle?
- *3. Draw the state-space graph for a predator isocline with a carrying capacity and alternative prey, and a victim isocline with a carrying capacity and an Allee effect. Discuss predator–prey dynamics at the two intersection points.
- *4. You are studying an insect-eating bird with a Type II functional response for which $k = 100$ prey/hour and $D = 5$.
 - a. What is the attack rate, α ?
 - b. If the prey density is 75, what is the feeding rate?

* Advanced problem

Solutions

- From Equation 6.3, the solution for the victim isocline is $0.1/0.001 = 100$ spiders, and the solution for the predator isocline (Equation 6.4) is $0.5/0.001 = 500$ flies. Plotting the isoclines and the initial population sizes gives:



Because we are above the victim isocline, there are too many predators, so the fly population will decline. However, we are to the right of the predator isocline, so there are enough victims to allow the spider population to increase. Both populations will undergo smooth cycles.

- From Equation 6.5, we first can solve for the missing value of q :

$$10 = \frac{2\pi}{\sqrt{0.5q}}$$

$$100 = \frac{4\pi^2}{0.5q}$$

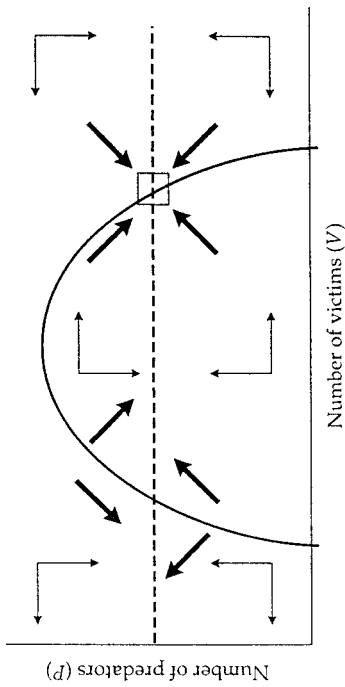
$$50q = 4\pi^2$$

Thus, $q = 0.7896$. If q is doubled in size and r is 0.5, the period of the cycle becomes:

$$\text{period} = \frac{2\pi}{\sqrt{(0.5)(2)(0.7896)}} = \frac{6.283}{0.889} = 7.07 \text{ years}$$

So, the period of the cycle decreases to approximately 7 years.

- The state-space drawing is as follows:



From this drawing, it is easy to see that the equilibrium on the right is stable, and predator and prey will coexist. However, the equilibrium on the left is unstable, and if the victim population falls below a critical minimum, the predators will drive it to extinction. Note the similarity between this analysis and that of the Allee effect in the single-species logistic model (see Problem 2.3). However, do not confuse the state-space graphs with the single-species graphs of density-dependent birth and death rates!

- Because $k = 1/h$, we have:

$$D = \frac{1}{\alpha h}$$

$$\alpha D = \frac{1}{h}$$

$$\alpha D = k$$

$$\alpha = \frac{k}{D} = \frac{100}{5} = 20 \text{ prey/hour}$$

- From Equation 6.8:

$$\frac{dV}{dt} = \frac{kV}{D+V} = \frac{100(75)}{5+75} = \frac{7500}{80} = 93.8 \text{ prey/hour}$$

CHAPTER 7

Island
Biogeography



Model Presentation and Predictions

THE SPECIES-AREA RELATIONSHIP

One of the few genuine "laws" in ecology is the **species-area relationship**: large islands support more species than small islands. The pattern holds for most assemblages of organisms, everything from vascular plants of the British Isles to reptiles and amphibians of the West Indies. The "islands" need not even be oceanic. Fish that live in lakes, mammals that occupy patches of forested mountaintops, and insects that visit thistle-heads all show a species-area relationship for their respective habitat islands. Because a national park or a nature reserve effectively is an island in a sea of disturbed habitat, studies of the species-area relationship may be relevant to the preservation of species in a fragmented landscape. This chapter explores in detail the relationship between area and the number of species in the community (species richness).

Figure 7.1a shows a typical species-area relationship for species of breeding land-birds on islands of the West Indies. The x axis shows the area of island, and the y axis shows the number of breeding land-bird species. You can see that the relationship is not linear: species number increases rapidly with area for small islands, but more slowly for large islands. For many oceanic islands, a rule of thumb is that a tenfold increase in island area results in a doubling of species number (Darlington 1957). Mathematically, the species-area relationship for many communities can be described by a simple power function:

$$S = cA^z \quad \text{Equation 7.1}$$

In this equation, S is the number of species, A is the island area and z and k are fitted constants, which we will explain in a moment. If we take the logarithm (base 10) of each side of the equation, we have:

$$\log(S) = \log(c) + z \log(A) \quad \text{Equation 7.2}$$

The logarithmic transformation turns the species-area curve into a straight line when plotted on logarithmic axes. The constant $\log(c)$ is the intercept of that line, and the constant z is the slope of the line. Figure 7.1b shows the West Indian bird data replotted with both axes transformed to logarithms. The data conform fairly well to a straight line, suggesting a good fit to the power function.

The area of an island is not the only factor that affects species richness.

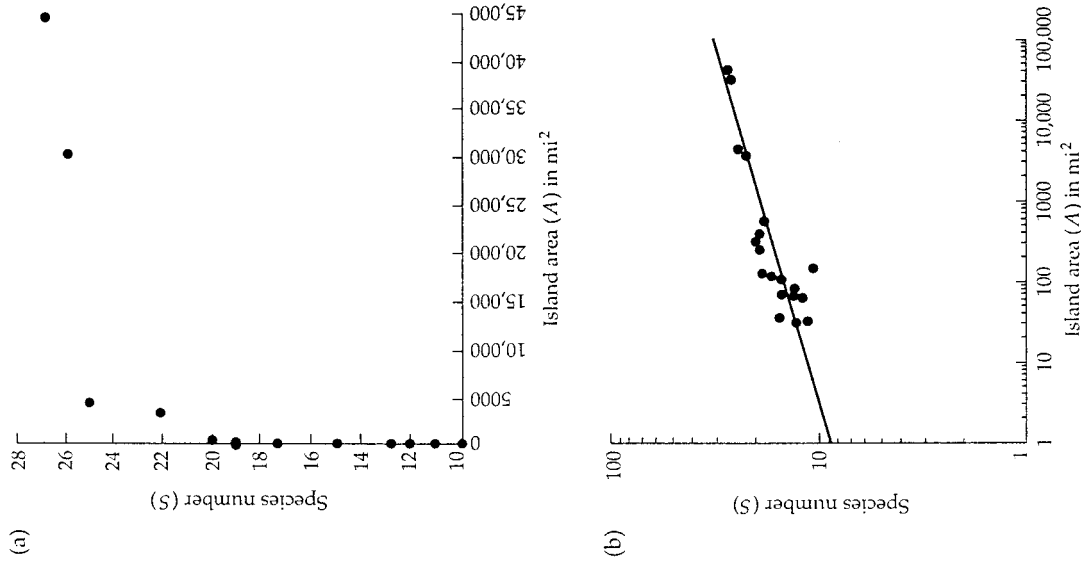


Figure 7.1 (a) Species-area relationship for breeding land-birds of the West Indies. Each point is a different island. Note that species number increases rapidly for small islands, but more slowly for large islands. (Data from Gotelli and Abele 1982.) (b) Logarithmic (base 10) transformation of the species-area relationship. The data in (a) have been plotted on a double-log plot. The best-fitting power function is shown by the straight line $\log(S) = 0.942 + 0.113 \log(A)$. Equivalently, the power function is $S = 8.759(A)^{0.113}$.

Figure 7.2 shows the effects of distance on species richness for birds of the Bismarck Islands in the tropical Pacific. New Guinea serves as the probable "source pool" for these islands, because all of the bird species found on the Bismarck Islands are a subset of the New Guinea avifauna. The x axis of this graph gives the distance from each island to New Guinea. The y axis shows the observed number of species divided by the number expected for a "near" island (< 500 km from New Guinea) of comparable area. You can see that relative species richness decreases with increasing distance from the source pool. In general, species richness is reduced for communities in small or isolated areas. In the following sections, we develop several models that attempt to explain the **area effect** (more species on large islands than on small islands) and the **distance effect** (more species on near islands than on far islands).

THE HABITAT DIVERSITY HYPOTHESIS

The most straightforward explanation for the species-area relationship is that large islands contain more habitat types than small islands. Therefore, species that are restricted to certain habitat types may occur only on large islands with those habitats. The species-area relationship for West Indian land-birds can be

explained, in part, by this phenomenon. The largest islands in the chain are the Greater Antilles (Puerto Rico, Cuba, Hispaniola, and Jamaica). These islands include many unique habitat types, such as extensive swampland (Cuba) and high-elevation pine forest (Hispaniola), that do not occur on any of the smaller islands. Habitat specialists such as the Zapata wren of Cuba (*Fermilia cervina*) and the white-winged crossbill of Hispaniola (*Loxia leucoptera*) occur only in these particular habitats. Intermediate-sized islands, such as Guadeloupe and St. Lucia, are steep volcanic plugs that have fewer habitats and species than islands of the Greater Antilles. Some of the smaller islands, such as Antigua and Barbuda, are flat coral atolls. They are arid islands with structurally simple vegetation, and they support even fewer bird species.

Although habitat diversity can account for many species-area relationships, it is not always the correct explanation. For one thing, most species are not extreme habitat specialists, and their distribution may not always be limited by available habitat. In addition, there are many examples of species-area relationships in which there is little, if any, habitat variation. Within patches of identical habitat, species number is still greater on large islands than small, suggesting that other forces may be at work. In the next section, we develop the "equilibrium model" of island biogeography as an alternative hypothesis that accounts for the species-area relationship. Later in this chapter, we describe a third hypothesis, the passive sampling model, which can also explain the species-area relationship.

THE EQUILIBRIUM MODEL OF ISLAND BIOGEOGRAPHY

The **equilibrium model of island biogeography** was popularized by R. H. MacArthur and E. O. Wilson (1963, 1967). It is sometimes referred to as the "equilibrium model" or the "MacArthur-Wilson model." The model's basic premise is that the number of species occurring on an island represents a balance between recurrent *immigration* of new species onto the island, and recurrent *extinction* of resident species. When immigration and extinction rates are equal, the number of species is at an equilibrium. The concept is similar to the equilibrium N in a local population (Chapter 2), and to the equilibrium fraction of sites occupied by a metapopulation (Chapter 4).

The equilibrium model assumes there is a permanent mainland source pool of species that can potentially colonize an island. There are P species in the mainland pool, and we assume for now they are all similar to one another in colonization and extinction potential. We define the immigration rate, λ_s , as the number of new species colonizing the island per unit time. The extinction rate, μ_s , is the number of species already present on the island going extinct per unit time. The rate of change in species number on the island (dS/dt) is the difference between the immigration rate and the extinction rate.

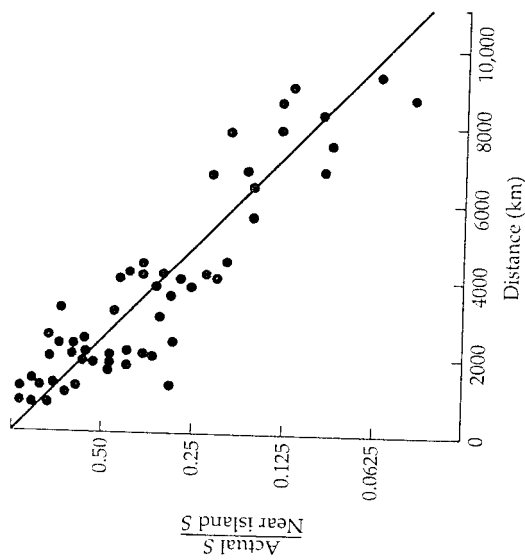


Figure 7.2 Distance effects for birds of the Bismarck Archipelago. The x axis gives the distance from each island to New Guinea, the presumed source pool. The y axis shows the observed species richness divided by the expected species richness for a "near" island (< 500 kilometers from New Guinea) of comparable size. (From Diamond 1972.)

Thus:

$$\frac{dS}{dt} = \lambda_s - \mu_s \quad \text{Equation 7.3}$$

First, we will define the functions for λ_s and μ_s . Next, we will set Equation 7.3 equal to zero and solve for the equilibrium number of species. Finally, we will modify the extinction and immigration curves to account for the effects of area and isolation on species richness.

Figure 7.3 illustrates the immigration curve for the equilibrium model. The x axis of the graph shows the number of species present on the island. The y axis shows the rate of immigration. The maximum immigration rate, I , occurs when the island is empty. The immigration rate *decreases* as more species are added to the island. This is because as more species are present, fewer *new* species remain in the source pool as potential colonists. Finally, suppose that all of the species in the source pool are present on the island. By definition, there can be no further immigration, so the immigration curve crosses the x axis at the point $S = P$. Thus, the immigration curve is a decreasing straight line, with a maximum rate of I , and a minimum rate of zero, when $S = P$.

Remember that a straight line can be described by the equation $y = a + bx$, where a is the intercept and b is the slope. In this case, the intercept is I , and the slope (rise over run) is $-I/P$. Thus, the equation for the immigration rate is:

$$\lambda_s = I - \left(\frac{I}{P}\right)S \quad \text{Expression 7.1}$$

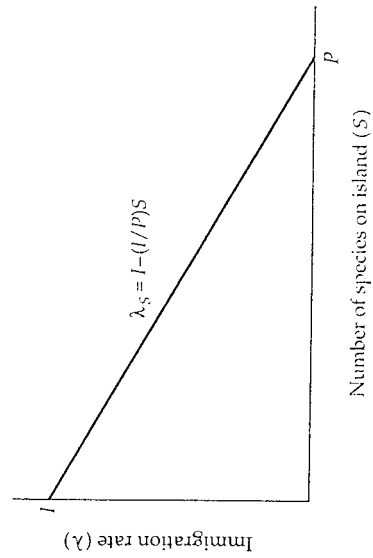


Figure 7.3 The immigration rate in the MacArthur–Wilson model. The immigration rate (number of species per unit time) decreases as more species are added to the island.

Now we turn to the extinction rate, μ_s . We expect μ_s to *increase* with increasing S : the more species present on the island, the greater the rate at which species disappear. This relationship occurs because each species has a constant probability of disappearance, so species disappear at a faster rate when there are more species present on the island. The maximum extinction rate, E , will occur when all the species in the source pool are present on the island ($S = P$). Conversely, if no species are present on the island ($S = 0$), the extinction rate must equal zero. Thus, the extinction curve is also a straight line with an intercept of zero, and a maximum rate of E , which occurs when $S = P$ (Figure 7.4):

$$\mu_s = \left(\frac{E}{P}\right)S \quad \text{Expression 7.2}$$

Now that we have derived expressions for linear immigration and extinction rates, we can substitute these into Equation 7.3 to model the change in species richness on an island:

$$\frac{dS}{dt} = I - \left(\frac{I}{P}\right)S - \left(\frac{E}{P}\right)S \quad \text{Expression 7.3}$$

The number of species on an island reaches an equilibrium when (dS/dt) equals zero. Setting Expression 7.3 equal to zero and solving for S yields:

$$S\left(\frac{I+E}{P}\right) = I \quad \text{Expression 7.4}$$

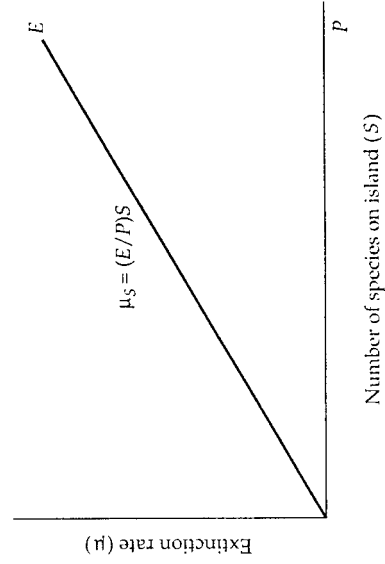


Figure 7.4 The extinction rate in the MacArthur–Wilson model. The extinction rate (number of species extinctions per unit time) increases as more species are added to the island.

The equilibrium number of species, \hat{S} , is thus:

$$\hat{S} = \frac{IP}{I + E} \quad \text{Equation 7.4}$$

The equilibrium depends on the size of the source pool (P) and the maximum immigration (I) and extinction (E) rates. Graphically, this equilibrium species number corresponds to the point on the x axis beneath the intersection of the immigration and extinction curves (Figure 7.5). At the intersection, the rate at which new species arrive is matched by the rate at which species present on the island go extinct.

This equilibrium point is stable. If we are below \hat{S} , we are to the left of the intersection point. In this region of the graph, the immigration rate exceeds the extinction rate, so species number increases. To the right of the intersection, extinctions exceed immigrations, so species number declines.

Equation 7.4 shows that species richness is increased by larger source pools and higher immigration rates, and decreased by higher extinction rates. Note the similarity between this equilibrium and the equilibrium in the island-mainland metapopulation model (Equation 4.4) we derived in Chapter 4. The intersection of the immigration and extinction curves also resembles the intersection of density-dependent birth and death rate curves in our derivation of the logistic growth equation (Figure 2.1) in Chapter 2.

Figure 7.5 also shows that the equilibrium is characterized by a **turnover rate**, which is measured on the y axis of the equilibrium graph. This turnover rate, T , is the number of species arriving (or disappearing) per unit time at equilibrium. T can be measured as either the extinction or the immigration rate, because these two are equal at equilibrium. Using some simple geometry, we see in Figure 7.5 that:

$$\frac{\hat{T}}{\hat{S}} = \frac{E}{P} \quad \text{Expression 7.5}$$

Therefore:

$$\hat{T}P = \hat{S}E \quad \text{Expression 7.6}$$

Rearranging and substituting Equation 7.4 for \hat{S} gives:

$$\hat{T} = \frac{\left(\frac{IP}{I + E}\right)E}{P} \quad \text{Expression 7.7}$$

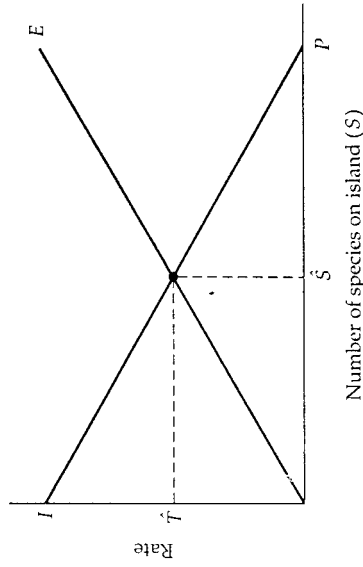


Figure 7.5 Equilibrium species number in the MacArthur–Wilson model. The intersection of the immigration and extinction curves determines the equilibrium number of species (\hat{S}) and the turnover rate (\hat{T}) at equilibrium.

$$\hat{T} = \frac{IE}{I + E} \quad \text{Equation 7.5}$$

Note that the turnover rate at equilibrium depends only on the maximum immigration and extinction rates (I and E), not on the size of the source pool (P). As you might expect, increasing either the maximum immigration or extinction rate increases the turnover at equilibrium.

This turnover of island populations at equilibrium is a key feature of the MacArthur–Wilson model. In contrast to many of the ecological models we have studied, the MacArthur–Wilson model does not predict stable population sizes. Instead, there is ongoing colonization and stochastic extinction of island populations. Species composition on the island is continually changing, although total species number remains relatively constant.

So far, we have constructed an equilibrium model of island species richness, but we still haven't explained the species–area effect. To do so, we must incorporate two additional assumptions about the demography of the colonizing species. The first assumption is that total population size for each species is proportional to island area. In other words, the *density* of populations (number of individuals per unit of area) is the same on islands of different size. The second assumption is that the probability of population extinction decreases with increasing population size. This assumption follows directly from our models of demographic and environmental stochasticity developed in Chapter 1. Because population sizes will be larger on big islands than on small islands, the extinction rates will correspondingly be lower on big islands.

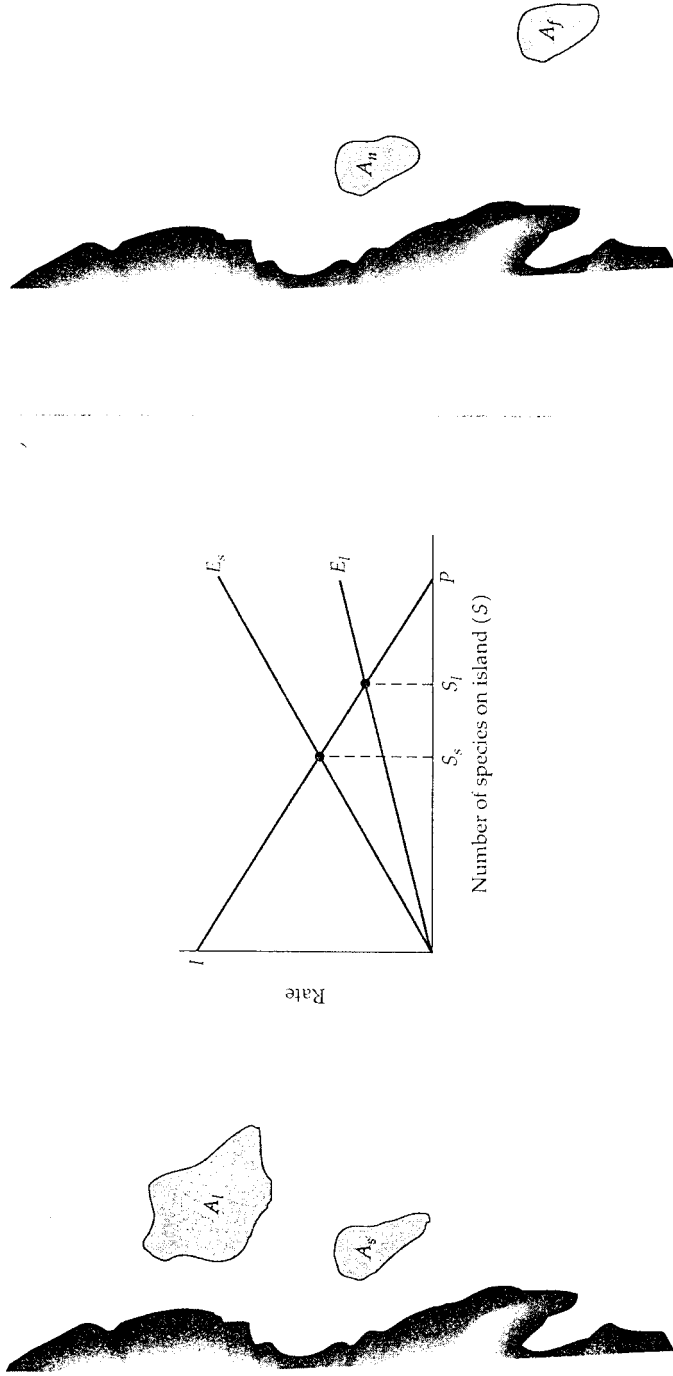


Figure 7.6 The area effect in the MacArthur–Wilson model. Smaller islands have smaller population sizes, which increases the extinction rate and leads to a lower species equilibrium. I_n is the maximum immigration rate for small islands; E_l is the maximum extinction rate for large islands.

Suppose we have a large island (A_l) and a small island (A_s) that differ only in area, but are identical in habitat diversity and distance from the source pool (Figure 7.6). Because both islands are equidistant from the mainland and colonized by the same source pool of P species, they have the same immigration curve. However, maximum extinction rates on the large island (E_l) are lower than on the small island (E_s) because population sizes are greater on the large island. Therefore, the equilibrium number of species is greater on the large island, with a lower rate of turnover.

We can also account for the “distance effect” by modifying the immigration curves for near and distant islands. Suppose two islands have identical areas and habitats, but differ in their distance from the source pool (Figure 7.7). Because the areas are equal, the two islands have the same extinction curve. But the maximum immigration rate will be higher on the near island

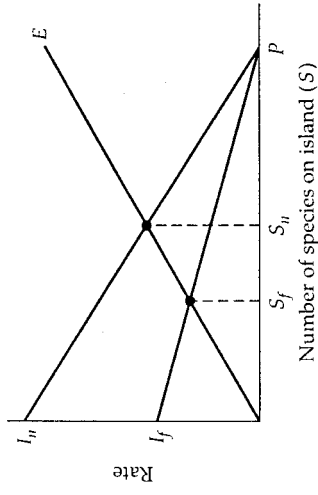


Figure 7.7 The distance effect in the MacArthur–Wilson model. Islands that are distant or isolated from the source pool have a reduced immigration rate, leading to a lower species equilibrium. I_n is the maximum immigration rate for near islands; I_f is the maximum immigration rate for far islands.

(I_n) than on the far island (I_f). Consequently, the near island will have more species at equilibrium than the far island. The near island will also be characterized by greater turnover than the far island.

Thus, island species richness in the MacArthur–Wilson model is uniquely determined by the geometry of an island—its area determines the extinction rate, and its distance or isolation determines its immigration rate. The intersection of these two curves controls the equilibrium number of species and the turnover rate.

Model Assumptions

Although the equilibrium model predicts patterns of species richness, its underlying assumptions are at the population level. These assumptions are:

- ✓ *An island potentially can be colonized by a set of P source pool species that have similar colonization and extinction rates.*
- ✓ *The probability of colonization is inversely proportional to isolation or distance from the source pool.*
- ✓ *The population size of a given species is proportional to the area of the island.*
- ✓ *The probability of a population becoming extinct is inversely proportional to its size.*
- ✓ *Extinction of local populations is independent of species composition on the island.*

Model Variations

NONLINEAR IMMIGRATION AND EXTINCTION CURVES

The linear immigration curve implies that all species have identical potential for dispersal and colonization of islands. But suppose that some species are much better at dispersal and colonization than others. These species would

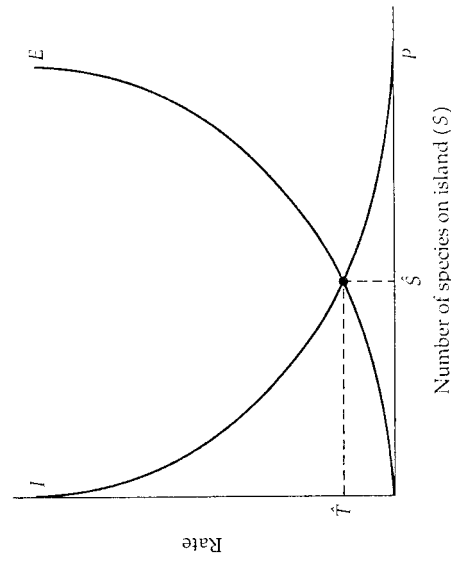


Figure 7.8 Nonlinear immigration and extinction curves in the MacArthur–Wilson model. These curves may reflect the influence of species interaction on the extinction rate and differential colonization ability on the immigration rate. The qualitative predictions of this nonlinear model are similar to those of the linear model described in the text (Figure 7.5).

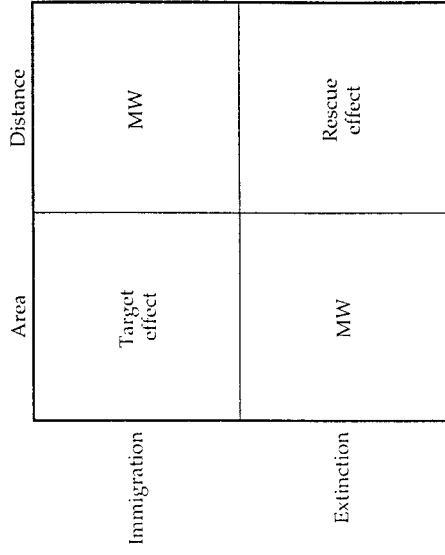


Figure 7.9 Area and distance effects in the MacArthur–Wilson model. The basic model (MW) considers the effects of area on the extinction rate and distance on the immigration rate. The model can be extended to incorporate the effects of distance on the extinction rate (rescue effect), and the effects of area on the immigration rate (target effect).

be among the first to colonize an empty island, whereas the poor dispersers would arrive later in the colonization sequence. With differential dispersal, the immigration curve would be exponential, with a steep decline initially and a slower rate of decrease as later species are added (Figure 7.8).

Similarly, the linear extinction curve implies that species extinctions are independent of one another. It might be more realistic to assume that competition increases the extinction rate when more species are present. In this case, the extinction curve would increase exponentially with S (Figure 7.8). In textbooks, the MacArthur–Wilson model is usually presented with these nonlinear immigration and extinction curves. Fortunately, the basic predictions of the equilibrium model remain the same, whether the linear or nonlinear rate curves are used.

AREA AND DISTANCE EFFECTS

Both area and distance affect extinction and immigration in the MacArthur–Wilson model. But the basic model describes only two mechanisms: the effect of area on extinction, and the effect of distance on immigration (Figure 7.9). In the next two sections, we briefly explore the effect of distance on extinction (the “rescue effect”), and the effect of island area on immigration (the “target effect”). These modifications incorporate more biological realism, but they also complicate the predictions of the simple MacArthur–

Wilson model. We then develop a "passive sampling" model that may also account for the species-area relationship without invoking habitat specialization or a species equilibrium.

THE RESCUE EFFECT

The MacArthur-Wilson model assumes that the only effect of distance or isolation is on the immigration rate. However, as we saw in Chapter 4, isolation can also affect the probability of extinction. In the metapopulation models of Chapter 4, we defined the rescue effect as the reduction in the probability of local extinction as the frequency of occupied patches increases. For the island model, we can define the rescue effect as the reduction in the species extinction rate for near versus far islands (Brown and Kodric-Brown 1977). Figure 7.10 illustrates the change in the immigration and extinction curves when a rescue effect is present. The basic prediction that large islands have more species than small ones remains unchanged with the rescue effect. However, the original MacArthur-Wilson model predicted *less* turnover on more isolated islands because they received fewer immigrants. In contrast, the rescue effect may generate *greater* turnover on more isolated islands because of the increase in the extinction rate.

THE TARGET EFFECT

The MacArthur-Wilson model considers only the effects of area on the extinction rate. However, island area might affect the immigration rate as well. To the extent that islands function as targets that intercept colonizing individu-

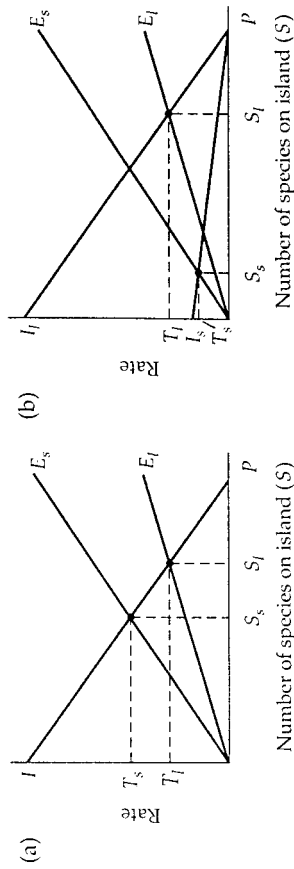


Figure 7.11 The target effect is the increase in the immigration rate on large islands versus small ones. Whereas the simple MacArthur-Wilson model predicts higher turnover on small islands (a), the target effect may increase turnover on large islands (b). T_s is the turnover rate on the small island; T_l is the turnover rate on the large island.

als, large islands may have higher immigration rates than small islands (Lomolino 1990). We can incorporate this **target effect** by assuming that the immigration rate is higher on large islands than on small. As in our analysis of the rescue effect, this change does not alter the pattern of species richness on large and small islands. If the target effect is strong enough, the model still predicts a species-area relationship, but turnover is now greater on large islands than on small (Figure 7.11).

THE PASSIVE SAMPLING MODEL

The nonlinear rate curves, rescue effect, and target effect are straightforward variations of the MacArthur-Wilson equilibrium model. All of these variations still describe species richness as a balance between ongoing immigration and ongoing extinction. But might there not be a simpler explanation for the species-area relationship? Suppose that islands function as passive "targets" that randomly accumulate individuals. Even in the absence of equilibrium turnover or habitat effects, we would still expect large islands to accumulate more species, by chance alone.

A useful analogy is to think of the islands as a set of targets. The area of each island is equivalent to the area of each target. Each individual organism is a dart, which is tossed randomly at the set of targets. The different species are represented by different colors of darts. Suppose we toss a handful of these darts at the targets. By chance, we expect the larger targets to accumulate more darts, and hence more colors, than the smaller targets. Similarly, if individuals colonize islands randomly, large islands should accumulate more individuals and species than small islands.

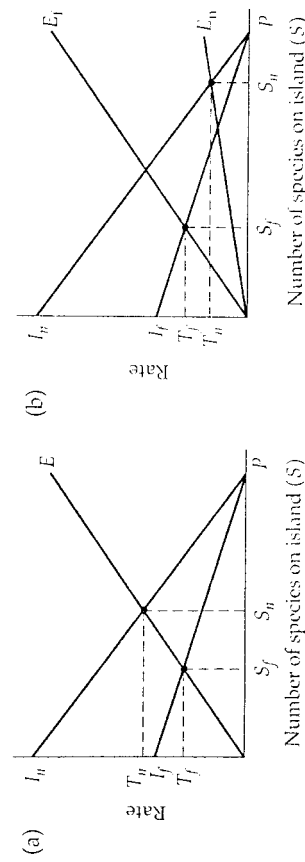


Figure 7.10 The rescue effect is the reduction in the extinction rate of near islands versus distant ones. Whereas the simple MacArthur-Wilson model predicts higher turnover on near islands (a), the rescue effect may increase turnover on more distant islands (b). T_n is the turnover rate on the near island; T_f is the turnover rate on the far island.

We can use some simple principles of probability theory to develop this **passive sampling model** (Coleman et al. 1982). First, assume we have a set of k islands. We will use the counter i to denote the i th island. The area of the i th island in the list is denoted as a_i . For example, if the fifth island on our list has an area of 100 square miles, $a_5 = 100$. Similarly, we assume a set of s species. We will use the counter j to denote the j th species. The total abundance of species j (summed across all islands) is n_j . If there are a total of 300 individuals of the sixth species that occur in the archipelago, $n_6 = 300$.

Let A equal the summed area of all the islands:

$$A = \sum_{i=1}^k a_i \quad \text{Expression 7.8}$$

Next, define x_i as the proportional area of the i th island:

$$x_i = \frac{a_i}{A} \quad \text{Expression 7.9}$$

Note that these proportional areas sum to 1.0:

$$\sum_{i=1}^k x_i = 1.0 \quad \text{Expression 7.10}$$

x_i can also be interpreted as the *probability* that a randomly placed individual will intercept an island of area a_i . Therefore, the probability that a single individual will *not* reach a particular island is:

$$P(\text{1 miss}) = 1 - x_i \quad \text{Expression 7.11}$$

For species j , the probability that *all* n_j individuals miss the island is:

$$P(n_j \text{ misses}) = (1 - x_i)^{n_j} \quad \text{Expression 7.12}$$

Expression 7.12 gives the probability that none of the n_j individuals of species j land on the island. Therefore, the probability that *at least* one individual of species j is present on the island is:

$$P(\text{species } j \text{ occurs on island } i) = 1 - (1 - x_i)^{n_j} \quad \text{Expression 7.13}$$

Finally, if we sum these probabilities across all species, we obtain the expected species richness on island i [$E(S_i)$]:

$$E(S_i) = \sum_{j=1}^s [1 - (1 - x_i)^{n_j}] \quad \text{Equation 7.6}$$

Why should the expected species richness equal the sum of the probabilities

of species occurrence? Suppose that the probability of occurrence of each species was 0.5. Intuitively, you would expect to find about half of the species in the archipelago occurring on the island. This expectation has a variance associated with it (Coleman et al. 1982), but the derivation is beyond the scope of this primer.

Like the MacArthur–Wilson model, the passive sampling model predicts more species on large islands than on small. However, the MacArthur–Wilson model predicts recurrent extinction and turnover of island populations, whereas the passive sampling model does not invoke turnover. Instead, the passive sampling model predicts that an abundant species will have a greater chance of occurring on an island than a sparse species. In fact, if an island is extremely small, rare species may be unlikely to ever occur there. Thus, the passive sampling model gives us more predictive power about species composition than does the MacArthur–Wilson model. The passive sampling model does not explicitly account for the distance effect, although we could extend the theory by modifying the relative target area as a function of distance from the source pool.

Empirical Examples

INSECTS OF MANGROVE ISLANDS

The most famous test of the equilibrium model was by E. O. Wilson and his student, D. Simberloff. These authors studied the insects that colonized small mangrove “islands” in the Florida Keys (Wilson and Simberloff 1969, Simberloff and Wilson 1969). Each island consisted of one to several red mangrove trees (*Rhizophora mangle*) that grow in shallow seawater. The total arthropod source pool for these islands was approximately 250 species, and each island supported 20 to 50 species. There are thousands of these mangrove islands in the Florida Keys that differ in their area and distance from colonization sources.

Simberloff and Wilson chose six islands for experimental manipulation and carefully censused them at the start of the experiment. The islands were then covered with canvas and all the resident arthropods were killed with methyl bromide, an insecticide. Over the next year, the authors repeatedly censused the islands and recorded the presence of different insect species during recolonization. The basic predictions of the equilibrium model were confirmed: after 250 days, species number on most islands had returned to approximately the same level as before defaunation (Figure 7.12). Large, near islands accumulated more species than small, distant islands.

Perhaps more important, the census data revealed considerable turnover in species composition, which is the essential prediction of the MacArthur–Wilson

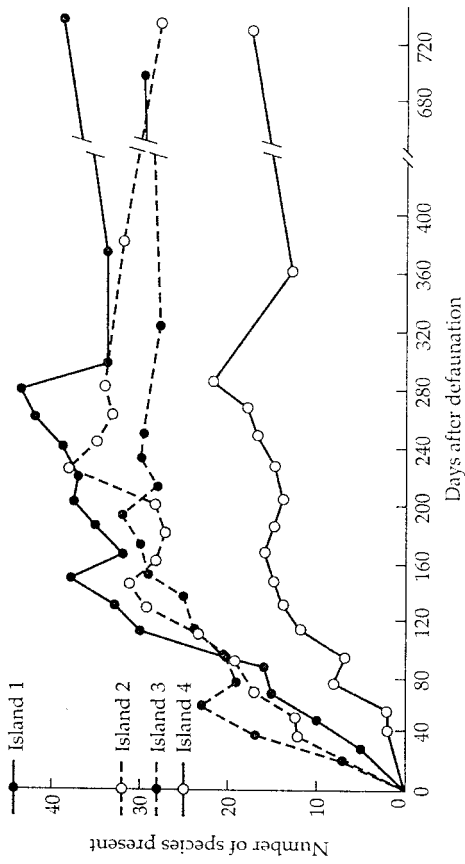


Figure 7.12 Insect recolonization of four defaunated mangrove islands. The y axis indicates the predefaunation species richness of each island. Most of the islands reached an equilibrium species number after 250 days that was approximately the same as the initial richness. (From Simberloff and Wilson 1969.)

model. Figure 7.13 shows part of the recolonization records for one of the six experimental islands. Although species number returned to an approximate equilibrium, species identity changed considerably from one census to the next, with an estimated turnover rate of 0.67 species per day.

However, Simberloff (1976) re-analyzed the mangrove data and questioned whether there was actually this much turnover. He pointed out that it is important to distinguish between local extinctions of relatively isolated breeding populations, and transient, short-term movements of individuals between islands. From the original census records, he eliminated those populations that were represented by only one or two individuals because they were unlikely to represent breeding populations. He also eliminated those populations that disappeared before they would have had time to reproduce. The corrected estimate of turnover was only 1.5 extinctions per year! Simberloff (1976) concluded that a test of the equilibrium theory required careful definitions of what constituted a true "colonization," and that much of the observed turnover in the mangrove insect community was among transient species.

BREEDING BIRDS OF EASTERN WOOD

Although the immigration and extinction curves (Figure 7.5) are the heart of the equilibrium model, they have rarely been measured in the field. An interesting exception is a long-term study of bird populations in a small plot with-

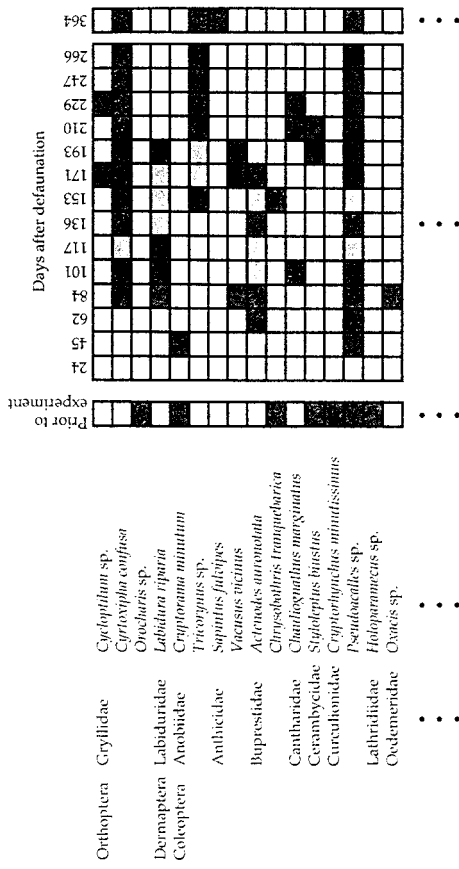


Figure 7.13 Colonization and extinction records for a single mangrove island. Each row is a species and each column is a census date. Records are shown for 16 of the 90 arthropod species that colonized the island. Open squares indicate species absence; darker squares indicate species presence. Lightly shaded squares indicate a species was not seen, but its presence was inferred from other evidence. Note the substantial turnover and change in species composition from one census to the next. (From Simberloff and Wilson 1969.)

in an oak forest (Williamson 1981). From 1947 through 1975, a team of ornithologists annually censused Eastern Wood, a 16-hectare plot of oak woodland in Surrey, England. The record of extinctions and colonizations can be plotted as a function of the number of species present each year, which varied from 27 to 36. The immigration curve matched the basic prediction of the equilibrium model, declining from an estimated maximum of 16 species per year to a value of zero at 40 resident species. This is somewhat less than the source pool estimate of 44 species. As predicted by the MacArthur–Wilson model, the extinction rate increased with S, although there was so much scatter in the data the trend was not statistically significant (Figure 7.14).

As in Simberloff's (1976) analysis, a detailed consideration of breeding status complicates the picture of species equilibrium. A core group of 14 species bred in the wood every year. A second group of 19 species did not establish substantial populations. These included species whose breeding status had not been confirmed (6), species that were represented by only 1 or 2 pairs in the plot (9), and species whose territories were larger than the area of the plot (4). The remaining 11 species were casual breeders that underwent frequent extinction.

The fit of the MacArthur–Wilson model to these data is somewhat ambigu-

ous. On the one hand, the existence of ecological turnover and the qualitative appearance of the immigration and extinction curves (Figure 7.14) matches the basic assumptions of the equilibrium model. On the other hand, the occurrence of 14 core species that bred every year in the woods could not have been predicted by the equilibrium model. This type of community structure may be fairly typical—one set of species with stable, persistent populations, and a second set of species with transient populations, which frequently go extinct and re-colonize. The models we have developed in Chapters 1, 2, 3, 5, and 6 may be more appropriate for the persistent populations, whereas the models in this chapter and in Chapter 4 may be more appropriate for transient populations.

BREEDING BIRDS OF THE PYMATUNING LAKE ISLANDS

Although the passive sampling model was introduced over 70 years ago (Arrhenius 1921), it has only received widespread attention since the early 1980s. Coleman et al. (1982) developed the mathematical predictions of the passive sampling model, and tested it with data on island breeding birds. A

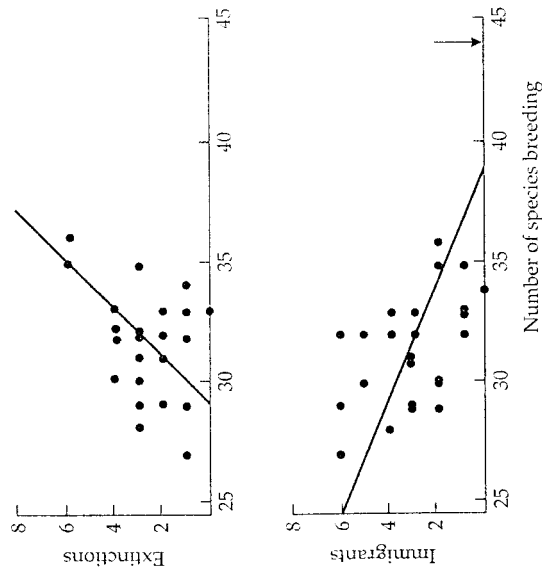


Figure 7.14 Immigration and extinction rates (species per year) for breeding land-birds of Eastern Wood. The immigration curve declined significantly with increasing species richness, but the extinction curve showed only a weak positive association with species richness. The arrow indicates 44 species, the source pool estimate. Compare to Figures 7.3 and 7.4. (From Williamson 1981.)

number of islands in Pymatuning Lake on the Ohio–Pennsylvania border were thoroughly censused for nests and bird territories. These islands were originally hilltops before a reservoir was created in 1932. The islands retain their deciduous forest vegetation, and the archipelago supports a pool of approximately 36 breeding land-bird species.

Coleman et al. (1982) knew the area of each island, and they were able to estimate the abundance of each species on the islands. They used these data to predict island species richness with the passive sampling model. In Figure 7.15, the solid line shows the predicted species richness and a confidence interval based on the passive sampling model. Species richness on most

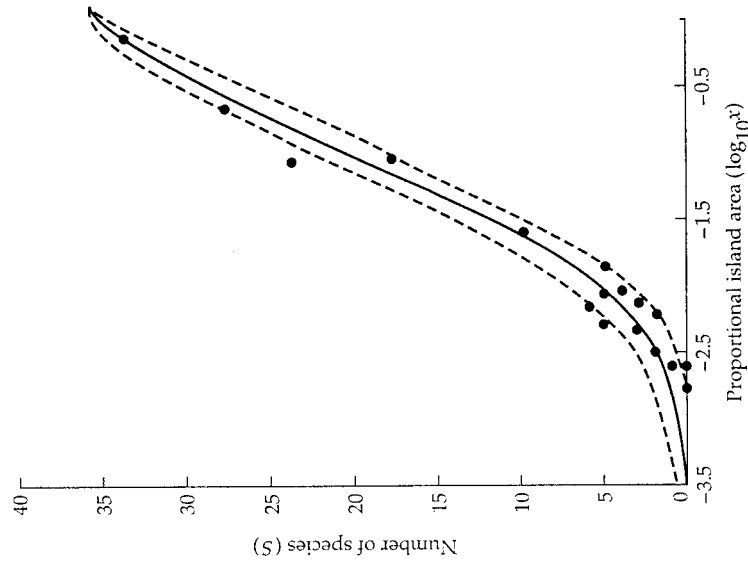


Figure 7.15 Observed and expected species richness for breeding land-birds on islands in Pymatuning Lake. The x axis gives the logarithm of the fractional area of each island. The solid line is the expected species richness and the dashed line is the confidence interval from the passive sampling model. Each circle is the observed species richness for each island. Note the good match between the observed data and the model predictions. (From Coleman et al. 1982.)

islands matched this prediction fairly well. In fact, the passive sampling model did a better job of predicting island species richness than did the power function.

One drawback of the passive sampling model is that it requires estimates of the abundance of all species on all islands, and these may be difficult to obtain. A second drawback is that the target analogy is conceptually simple, but biologically not very realistic. Many factors other than island area affect individual colonization, including weather and current patterns, seasonal migration, food resources, and the presence of other species of predators and competitors.

In conclusion, the species–area relationship represents one of the few general patterns in ecology, but its causes remain elusive. The habitat diversity hypothesis, the MacArthur–Wilson equilibrium model, and the passive sampling model are not mutually exclusive explanations—each may contribute to the species–area relationship. Additional data on habitat diversity, population turnover, and source pool structure are needed to gauge their relative contributions to the species–area relationship.

Problems

- For the West Indian land-bird data in Figure 7.1, the best-fitting power function has the constants $c = 8.759$ and $z = 0.113$. The island of Grenada has an area of 120 square miles, and supports 17 land-bird species.
 - What is the predicted number of species from the power function?
 - Suppose that half of the island's area disappears in a volcanic eruption. From the power function, how many species would be expected to remain?
- Your colleague returns from the South Pacific with data on island lizards. "Look," she says, "my data show that there are more species of lizards on small islands than large islands. This disproves the MacArthur–Wilson equilibrium model!" Using an appropriate set of immigration and extinction curves, show how more species could occur on a small island (A_2) than on a large island (A_1) in the MacArthur–Wilson model.
- Suppose that an island in MacArthur–Wilson equilibrium supports 75 species, out of a source pool of 100 species. The maximum extinction rate (E) is 10 extinctions per year. Calculate the maximum immigration rate (I). If I is doubled, what is the new species equilibrium and new turnover rate?
- Here are some hypothetical data on the abundances of six species of cactus on four small desert islands:

	Island 1 (110 ha)	Island 2 (100 ha)	Island 3 (10 ha)	Island 4 (5 ha)
Cactus A	3	0	0	0
Cactus B	1	0	0	0
Cactus C	4	2	3	1
Cactus D	2	0	2	2
Cactus E	1	0	1	0
Cactus F	1	0	0	3

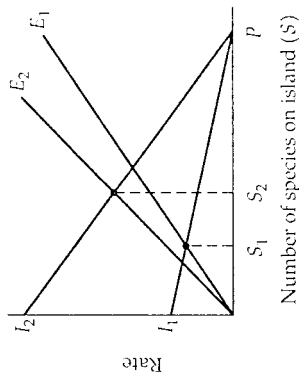
Calculate the expected number of species on each island for the passive sampling model. How close are the expected values to the observed number of species?

* Advanced problem

Solutions

- a. Plugging into Equation 7.1 gives $S = (8.759)(120)^{0.113} = 15.04$, which is close to the observed richness of 17 species.

b. With only half of the available area, the predicted species richness is $S = (8.759)(60)^{0.113} = 13.9$. So, roughly 14 species should be present, if the equation is completely accurate. Can you list several reasons why this forecast might be seriously incorrect?
- The equilibrium depends on both the immigration and the extinction curves. Therefore, if the large island (A_1) is very isolated, it may contain fewer species than the small island (A_2):



- First, we need to use Equation 7.4 to solve for I . Equation 7.4 is written in terms of \hat{S} , but we can rearrange it to give a solution for I :

$$\hat{S} = \frac{IP}{I + E}$$

$$\hat{S}(I + E) = IP$$

$$\hat{S}E = IP - \hat{S}I$$

$$I = \frac{\hat{S}E}{P - \hat{S}}$$

Plugging into this last expression yields $I = (75)(10)/(100 - 75) = 30$ species immigrations per year. If the immigration rate is doubled, $I = 60$. From Equation 7.4, the new species equilibrium is $(100)(60)/(60 + 10) = 85.7$ species. From Equation 7.5, the new turnover rate is $(60)(10)/(60 + 10) = 8.57$ species per year.

- The calculation from Equation 7.6 is tedious, so we will illustrate it in detail only for Island 1. First, we must calculate x_1 , the relative area of Island 1.

From Expression 7.9:

$$x_1 = \frac{a_1}{A} = \frac{110}{110 + 100 + 10 + 5} = 0.489$$

Now we can use Expression 7.13 to find the probability of occurrence for each of the six species. Remember that n_j in this expression is the total abundance of each species, which we obtain by summing the rows of the data matrix:

- Species A $1 - (1 - 0.489)^3 = 0.867$
- Species B $1 - (1 - 0.489)^1 = 0.489$
- Species C $1 - (1 - 0.489)^{10} = 0.999$
- Species D $1 - (1 - 0.489)^6 = 0.982$
- Species E $1 - (1 - 0.489)^2 = 0.739$
- Species F $1 - (1 - 0.489)^4 = 0.932$

From Equation 7.6, the expected species richness on Island 1 is:

$$E(S_1) = 0.867 + 0.489 + 0.999 + 0.982 + 0.739 + 0.932 = 5.008$$

If we continue these calculations for all four islands, we obtain:

	Observed S	Expected S
Island 1	6	5.008
Island 2	1	4.835
Island 3	3	1.020
Island 4	3	0.540

The expected species richness does not match the observed very well. For example, the second largest island had only a single species on it, whereas the passive sampling model predicted almost 5 species. The two smallest islands each supported 3 species, but the passive sampling model predicted 1 or zero species. These data suggest that a random placement of individuals cannot account for the observed species richness data.

Literature Cited

- Abramsky, Z., M. L. Rosenzweig and B. Pinshow. 1991. The shape of a gerbil isocline measured using principles of optimal habitat selection. *Ecology* 72: 329-340. [5]
- Abramsky, Z., O. Ovadia and M. L. Rosenzweig. 1994. The shape of a *Cerbittus pyramidum* (Rodentia: Gerbillinae) isocline: an experimental field study. *Oikos* 69: 318-326. [5]
- Açkaya, H. R. 1992. Population cycles of mammals: evidence for a ratio-dependent predation hypothesis. *Ecological Monographs* 62: 119-142. [6]
- Allee, W. C., A. E. Emerson, O. Park, T. Park and K. P. Schmidt. 1949. *Principles of Animal Ecology*. W. B. Saunders, Philadelphia. [2]
- Anderson, R. M. and R. M. May. 1978. Regulation and stability of host-parasite population interactions. I. Regulatory processes. *Journal of Animal Ecology* 47: 219-249. [6]
- Arceese, P. and J. N. M. Smith. 1988. Effects of population density and supplemental food on reproduction in the song sparrow. *Journal of Animal Ecology* 57: 119-136. [2]
- Arrhenius, O. 1921. Species and area. *Journal of Ecology* 9: 95-99. [7]
- Berryman, A. 1992. The origins and evolution of predator-prey theory. *Ecology* 73: 1530-1535. [6]
- Boerema, L. K. and J. A. Gulland. 1973. Stock assessment of the Peruvian anchovy (*Engraulis ringens*) and management of the fishery. *Journal of the Fisheries Research Board of Canada* 30: 2226-2235. [2]
- Brown, J. H. and A. Kodric-Brown. 1977. Turnover rates in insular biogeography: effect of immigration on extinction. *Ecology* 58: 445-449. [7]
- Caswell, H. 1989. *Matrix Population Models*. Sinauer Associates, Sunderland, Mass. [3]
- Caughley, G. 1977. *Analysis of Vertebrate Populations*. Wiley, New York. [3]
- Coleman, B. D., M. A. Mares, M. R. Willig and Y.-H. Hsieh. 1982. Randomness, area, and species richness. *Ecology* 63: 1121-1133. [7]
- Darlington, P. J. 1957. *Zoogeography: The Geographical Distribution of Animals*. Wiley, New York. [7]
- den Boer, P. J. 1981. On the survival of populations in a heterogeneous and variable environment. *Oecologia* 50: 39-53. [4]
- Dennis, B., P. L. Munholland and J. M. Scott. 1991. Estimation of growth and extinction parameters for endangered species. *Ecological Monographs* 61: 115-144. [1]
- Diamond, J. M. 1972. Geographic kinetics: estimation of relaxation times for avifaunas of southwest Pacific islands. *Proceedings of the National Academy of Sciences, USA* 69: 3199-3203. [7]
- Dobson, A. P. and P. J. Hudson. 1992. Regulation and stability of a free-living host-parasite system: *Trichostrongylus tenuis* in red grouse. II. Population models. *Journal of Animal Ecology* 61: 487-498. [6]
- Elton, C. and M. Nicholson. 1942. The ten-year cycle in numbers of the lynx in Canada. *Journal of Animal Ecology* 11: 215-244. [6]
- Ehrlich, P. R., R. R. White, M. C. Singer, S. W. McKechnie and L. E. Gilbert. 1975. Checkerspot butterflies: a historical perspective. *Science* 188: 221-228. [4]
- Fenchel, T. 1974. Intrinsic rate of natural increase: the relationship with body size. *Oecologia* 14: 317-326. [1]
- Fisher, R. A. 1930. *The Genetical Theory of Natural Selection*. Clarendon Press, Oxford. [3]
- Gallagher, E. D., G. B. Gardner and P. A. Jumars. 1990. Competition among the pioneers in a seasonal soft-bottom benthic succession: field experiments and analysis of the Gilpin-Ayala competition model. *Oecologia* 83: 427-442. [5]
- Gotelli, N. J. and L. G. Abele. 1982. Statistical distributions of West Indian land-bird families. *Journal of Biogeography* 9: 421-435. [7]
- Gotelli, N. J. and W. G. Kelcey. 1993. A general model of metapopulation dynamics. *Oikos* 68: 36-44. [4]
- Hanski, I. 1982. Dynamics of regional distribution: the core and satellite species hypothesis. *Oikos* 38: 210-221. [4]
- Hanski, I. and M. Gilpin. 1991. Metapopulation dynamics: brief history and conceptual domain. *Biological Journal of the Linnean Society* 42: 3-16. [4]
- Hardin, G. 1960. The competitive exclusion principle. *Science* 131: 1292-1297. [5]
- Hardin, G. 1968. The tragedy of the commons. *Science* 162: 1243-1248. [2]
- Harrison, S., D. D. Murphy and P. R. Ehrlich. 1988. Distribution of the bay checkerspot butterfly, *Euphydryas editha bayensis*: evidence for a metapopulation model. *The American Naturalist* 132: 360-382. [4]
- Holling, C. S. 1959. The components of predation as revealed by a study of small mammal predation of the European pine sawfly. *Canadian Entomologist* 91: 293-320. [6]
- Hudson, P. J., A. P. Dobson and D. Newborn. 1985. Cyclic and noncyclic populations of red grouse: a role for parasitism? In D. Rollinson and R. M. Anderson (eds.), *Ecology and Genetics of Host-Parasite Interactions*, pp. 77-89. Academic Press, London. [6]
- Hudson, P. J., D. Newborn and A. P. Dobson. 1992. Regulation and stability of a free-living host-parasite system: *Trichostrongylus tenuis* in red grouse. I. Monitoring and parasite reduction experiments. *Journal of Animal Ecology* 61: 477-486. [6]
- Hutchinson, G. E. 1967. *A Treatise on Limnology, Vol. II. Introduction to Lake Biology and Limnoplankton*. Wiley, New York. [5]
- Keith, L. B. 1983. Role of food in hare population cycles. *Oikos* 40: 385-395. [6]

- Kingsland, S. E. 1985. *Modeling Nature: Episodes in the History of Population Ecology*. University of Chicago Press, Chicago. [6]
- Krebs, C. J. 1985. *Ecology: The Experimental Analysis of Distribution and Abundance*, 3rd Ed. Harper & Row, New York. [2,5]
- Lack, D. 1967. *The Natural Regulation of Animal Numbers*. Clarendon Press, Oxford. [1]
- Lefkovich, L. P. 1965. The study of population growth in organisms grouped by stages. *Biometrics* 21: 1-18. [3]
- Leslie, P. H. 1945. On the use of matrices in certain population mathematics. *Biometrika* 35: 183-212. [3]
- Levins, R. 1969. The effect of random variations of different types on population growth. *Proceedings of the National Academy of Sciences, USA* 62: 1061-1065. [2]
- Levins, R. 1970. Extinction. In M. Gerstenhaber (ed.), *Some Mathematical Questions in Biology. Lecture Notes on Mathematics in the Life Sciences*, pp. 75-107. The American Mathematical Society, Providence, R.I. [4]
- Lomolino, M. V. 1990. The target area hypothesis: the influence of island area on immigration rates of non-volant mammals. *Oikos* 57: 297-300. [7]
- Luckinbill, L. S. 1979. Selection of the r/K continuum in experimental populations of protozoa. *The American Naturalist* 113: 427-437. [3]
- MacArthur, R. H. 1972. *Geographical Ecology*. Harper & Row, New York. [5]
- MacArthur, R. H. and E. O. Wilson. 1963. An equilibrium theory of insular zoogeography. *Evolution* 17: 373-387. [7]
- MacArthur, R. H. and E. O. Wilson. 1967. *The Theory of Island Biogeography*. Princeton University Press, Princeton, N.J. [3,7]
- May, R. M. 1973. *Stability and Complexity in Model Ecosystems*. Princeton University Press, Princeton, N.J. [2]
- May, R. M. 1974a. Ecosystem patterns in randomly fluctuating environments. *Progress in Theoretical Biology* 3: 1-50. [1,2]
- May, R. M. 1974b. Biological populations with non-overlapping generations: stable points, stable cycles, and chaos. *Science* 186: 645-647. [2]
- May, R. M. 1976. Models for single populations. In R. M. May (ed.), *Theoretical Ecology: Principles and Applications*, pp. 4-25. W. B. Saunders, Philadelphia. [2]
- Mertz, D. B. 1970. Notes on methods used in life-history studies. In J. H. Connell, D. B. Mertz and W. W. Murdoch (eds.), *Readings in Ecology and Ecological Genetics*, pp. 4-17. Harper & Row, New York. [3]
- Mitchell, W. A. and J. S. Brown. 1990. Density-dependent harvest rates by optimal foragers. *Oikos* 57: 180-190. [6]
- Moran, P. A. P. 1949. The statistical analysis of the sunspot and lynx cycles. *Journal of Animal Ecology* 18: 115-116. [6]
- Murphy, D. D. and P. R. Ehrlich. 1980. Two California bay checkerspot subspecies: one new, one on the verge of extinction. *Journal of the Lepidopterist Society* 34: 316-320. [4]
- Murphy, G. I. 1968. Pattern in life history and the environment. *The American Naturalist* 102: 391-403. [3]
- Pianka, E. R. 1970. On r - and K -selection. *The American Naturalist* 104: 592-597. [3]
- Pielou, E. C. 1969. *An Introduction to Mathematical Ecology*. Wiley, New York. [1]
- Pigg, J. 1988. Aquatic habitats and fish distribution in a large Oklahoma river, the Cimarron, from 1976-1986. *Proceedings of the Oklahoma Academy of Science* 68: 9-31. [4]
- Pitt, D. E. 1993. Talks at U.N. combat threat to oceans' species from overfishing. *The New York Times*, 6/23/93. [2]
- Polis, G. A., C. A. Myers and R. D. Holt. 1989. The ecology and evolution of intraguild predation: potential competitors that eat each other. *Annual Review of Ecology and Systematics* 20: 297-330. [5]
- Ray, C., M. Gilpin and A. T. Smith. 1991. The effect of conspecific attraction on metapopulation dynamics. *Biological Journal of the Linnean Society* 42: 123-134. [4]
- Real, L. 1977. The kinetics of functional response. *The American Naturalist* 111: 289-300. [6]
- Rose, M. R. 1984. The evolution of animal senescence. *Canadian Journal of Zoology* 62: 1661-1667. [3]
- Rosenzweig, M. L. 1971. Paradox of enrichment: destabilization of exploitation ecosystems in ecological time. *Science* 171: 385-387. [6]
- Rosenzweig, M. L. and R. H. MacArthur. 1963. Graphical representation and stability conditions of predator-prey interactions. *The American Naturalist* 47: 209-223. [6]
- Roughgarden, J. 1979. *Theory of Population Genetics and Evolutionary Ecology: An Introduction*. Macmillan, New York. [2,3]
- Royama, T. 1971. A comparative study of models of predation and parasitism. *Researches in Population Ecology (Supplement)* 1: 1-91. [6]
- Simberloff, D. 1976. Species turnover and equilibrium island biogeography. *Science* 194: 472-478. [7]
- Simberloff, D. S. and E. O. Wilson. 1969. Experimental zoogeography of islands: the colonization of empty islands. *Ecology* 50: 278-289. [7]
- Sinclair, A. R. E., J. M. Gosline, G. Holdsworth, C. J. Krebs, S. Boutin, J. N. M. Smith, R. Boonstra and M. Dale. 1993. Can the solar cycle and climate synchronize the snowshoe hare cycle in Canada? Evidence from tree rings and ice cores. *The American Naturalist* 141: 173-198. [6]
- Slade, N. A. and D. F. Balph. 1974. Population ecology of Uinta ground squirrels. *Ecology* 55: 989-1003. [3]

- Smith, C. H. 1983. Spatial trends in Canadian snowshoe hare, *Lepus americanus*: population cycles. *Canadian Field-Naturalist* 97: 151-160. [6]
- Smith, J. N. M., C. J. Krebs, A. R. Sinclair and R. Boonstra. 1988. Population biology of snowshoe hares. II. Interaction with winter food plants. *Journal of Animal Ecology* 57: 269-286. [6]
- Smith, J. N., P. Arcese and W. M. Hochachka. 1991. Social behaviour and population regulation in insular bird populations: implications for conservation. In C. M. Perrins, J.-D. Lebreton and G. J. M. Hiron (eds.), *Bird Population Studies: Relevance to Conservation and Management*, pp. 148-167. Oxford University Press, Oxford. [2]
- Solomon, M. E. 1949. The natural control of animal populations. *Journal of Animal Ecology* 18: 1-35. [6]
- Stearns, S. C. 1992. *The Evolution of Life Histories*. Oxford University Press, Oxford. [3]
- Swane, I. 1984. Observations on the long-term population dynamics of the perennial ascidian, *Ascidia mentula* O. F. Muller, on the Swedish west coast. *Biological Bulletin* 167: 630-646. [2]
- Taylor, C. E. and C. Condra. 1980. *r* selection and *K* selection in *Drosophila pseudoobscura*. *Evolution* 34: 1183-1193. [3]
- Werner, P. A. 1977. Colonization success of a "biennial" plant, teasel (*Dipsacus sylvestris* Huds.): experimental field studies in species cohabitation and replacement. *Ecology* 58: 840-849. [3]
- Werner, P. A. and H. Caswell. 1977. Population growth rates and age vs. stage distribution models for teasel (*Dipsacus sylvestris* Huds.). *Ecology* 58: 1103-1111. [3]
- Williamson, M. 1981. *Island Populations*. Oxford University Press, Oxford. [7]
- Wilson, E. O. and W. H. Bossert. 1971. *A Primer of Population Biology*. Sinauer Associates, Sunderland, Mass. [3]
- Wilson, E. O. and D. S. Simberloff. 1969. Experimental zoogeography of islands: defaunation and monitoring techniques. *Ecology* 50: 267-295. [7]
- A, *see* Area
- Activity density, gerbil competition study, 132-135
- Age class, 56-59
- age structure description and, 65-66
- fertility of, 66-67
- survival probability, 66
- Age distribution, 69-71
- Age structure (*n*), 56-57, 65-71
- describing, 65-66
- AIDS virus 166
- Allee effect
- in ascidian populations, 46-47
- defined, 29
- victim population size and, 154
- Allolopathy, 112
- α (alpha), *see* Competition coefficient;
- Encounter rate
- Amplitude, of population cycle, 35-36
- Anchovy fisheries, 47-49
- Annual plants, 59
- Area (*A*), number of species and, 172-174
- Area effect, 174, 180-181, 183-184
- Arkansas River shiner, 106
- Ascidia mentula*, 45-47
- Ascidians, population fluctuations, 45-47
- Aticemia marina*, doubling time, 8
- b*, *see* Birth rate
- B*, *see* Births
- Bacteria, doubling time, 8
- Bay checkerspot butterfly, 101-102
- Bears, grizzly, 20-22
- Beetles, metapopulation study, 103, 104
- β (beta), *see* Competition coefficient;
- Conversion efficiency
- Birds
- immigration and extinction study, 188-190
- migratory, 90
- passive sampling model study, 190-192
- population fluctuations, 43-45
- species-area relationship, 172-175
- Birth rate (*b*)
- age structure and, 56
- defined, 4-5
- fertility schedule [*b(x)*], 57-59, 62, 68
- life history strategies and, 75-76
- population density and, 28-29
- stable equilibrium, 31
- Birth-flow model, 66
- Birth-pulse model, 66
- Births, number of (*B*), 2-4
- in Euler equation, 72-73
- Bismarck Archipelago, species-area relationships, 174
- Black-throated blue warbler, 90
- Body size
- intrinsic rate of increase and, 8
- predation and, 127
- Bos taurus*, doubling time, 8-9
- Butterfly, metapopulation study, 101-102
- Calidris melanoccephalus*, 103, 104
- Canada lynx, 161-162
- Carabid beetles, metapopulation study, 103, 104
- Carrying capacity (*K*), 30-32
- Lotka-Volterra competition model and, 113-114, 126
- optimal yield and, 47-49
- of predator population, 157-159
- variation in, 40-42
- of victim population, 148-149
- Chaos, population fluctuations, 37-39
- Checkerspot butterfly, 101-102
- Cimmaron River, 103-106
- Closed population, 3, 9, 90
- Coexistence, 122-123, 125-126
- Cohort
- defined, 59-60
- number surviving [*S(x)*], 59-60
- see also* Age class
- Cohort life table, 72
- Colonization
- internal, 97-98
- island-mainland model, 96-97
- rescue effect, 98-101
- see also* Immigration
- Combination of abundances, 116-119
- Commercial fisheries, 47-49
- Competition
- defined, 112, 127
- inter- vs. intraspecific, 113, 114
- Lotka-Volterra model, *see* Lotka-Volterra competition model

Index

- state-space graphs, 116-123
types of, 112-113
- Competition coefficient (α, β)
assumptions underlying, 126
defined, 114-115
Competitive coexistence (case 3), 122-123,
125-126
- Competitive exclusion
algebraic principle, 124-126
case 1, 119-120, 121, 125
case 2, 120-123, 125
case 4, 123-124, 125
- Conservation biology
reproductive value and, 75
- Constraints, on life history strategies, 75-76
- Continuous differential equation
for population growth (dN/dt), 4-6
- Conversion efficiency (β)
population cycles and, 147
of predator population, 142
- Coral, size-structured model, 78, 79
- Cows, doubling time, 8-9
- Curves
immigration and extinction, 176-177,
181-182, 189-190
survivorship, 61-62, 76
- Cycles
in carrying capacity, 40-42
hare-lynx populations, 161-162
in logistic growth model, 37-39
Lotka-Volterra predation model, 145-147
in population size, 35-37
red grouse population, 159-160
stability of, 159-160
- d*, see Death rate
- D*, see Deaths
- Damped oscillations, 35-36
- Darwin, Charles, 11
- Death rate (d), 4-5
age structure and, 56
population density and, 28-29
predator population equilibrium and,
141-143
stable equilibrium, 31
- Deaths, number of (D), 2-4
- Defaunation experiments, 187-188, 189
- Delay differential equation, 35-37
- Demographic stochasticity, 16-19
MacArthur-Wilson model and, 179
- Dendroica caerulescens*, 90
- Density dependence
in ground squirrels, 80-82
linear, 33, 34
Lotka-Volterra competition model and,
127
migration and, 90
population growth and, 28-29
- see also Population density
- Deterministic model, 13-14
- Differential equation, see Continuous differ-
ential equation
- Dipsacus sylvestris*, 82-85
- Discrete difference equation, 11-13
- Discrete population growth, 11-13
logistic model and, 37-39
- Distance effect, 174, 180-181, 183-184
- dN/dt , see Growth rate
- Doubling time, 6, 8-9
- e (base of natural logarithm), 6
- E*, see Emigrants; Extinction rate
- Eastern Wood, immigration/extinction study,
188-190
- Emigrants
number of (E), 2-3
- Encounter rate (α)
functional response and, 149-151
population cycles and, 147
predator-prey population model, 141
random, 147
- Endangered populations
grizzly bears, 20-21
modeling, 21-22
- Engaulis ringens*, 47-49
- Enrichment, 154, 156
- Environmental stochasticity
defined, 14
MacArthur-Wilson model and, 179
population growth and, 13-16
- Equilibrium
carrying capacity and, 31
Lotka-Volterra competition model,
115-116, 121-124
number of species, 175, 178-179
Equilibrium model of biogeography, 175-182
- Escherichia coli*, doubling time, 8
- Euler, L., 64
- Euler equation, 58, 64-65
derivation of, 72-73
- Euphydryas editha bayensis*, 101-102
- Exploitation competition, 112
- Exponential growth model, 5-6
age structure and, 56, 69-70, 72
assumptions underlying, 9-11
vs. logistic model, 33-34
theory of natural selection and, 11
trajectories, 7
- External colonization, 96, 98-101
- Extinction
Allee effect and, 29
demographic stochasticity and, 18-19
environmental stochasticity and, 16
local vs. regional, 91, 93
of metapopulations, 91-92, 93
rescue effect, 98-100
- Immigrants, number of (I), 2-3
- Immigration rate (I, λ)
distance effects and, 180-181
MacArthur-Wilson model, 175-176
metapopulation model, 93-94
nonlinear, 182-183
rescue effect and, 184
Insects
colonization experiment, 187-188, 189
stage-structured population model, 77-78,
79
- Instantaneous birth rate, see Birth rate
- Instantaneous death rate, see Death rate
- Instantaneous rate of increase (r)
age structure and, 56, 58, 64-66
body size and, 8
discrete generations and, 11-13
exponential model, 5-9
compared to net reproductive rate, 63
Leslie matrix and, 68, 79
transition matrix and, 70-71, 79
- Logistic model, 37-39
- Lotka-Volterra competition model,
113-114
in prey populations, 140
time lags and (τ), 35-37
transition matrix and, 79
- Interference competition, 112
- Internal colonization, 97-98, 100
- Interspecific competition, 113-114; see also
Competitive exclusion
- Intertidal worms, competition studies,
128-132
- Intraguild predation, 127-128, 129, 130
- Intraspecific competition, 113-114
- Intrinsic rate of increase, see Instantaneous
rate of increase
- Island-mainland model, 96-97
compared to MacArthur-Wilson model,
178
- Isoclines
intraguild predation, 128, 129, 130
Lotka-Volterra competition models,
119-124
of predator and prey populations,
143-145, 148, 153, 154-160
see also State-space graph
- Iteroparous reproduction, 59, 76
- K , see Carrying capacity
- K selection, 76
- $l(x)$, see Survivorship schedule
- Lagopus lagopus scoticus*, 162-166
- λ (lambda), see Finite rate of increase
- Larvae, population cycles and, 163, 164-166
- Lepus americanus*, 161-162
- Leslie matrix, 67-68
calculating r from, 69-71
describing age distribution, 68-69
- see also Endangered populations
- Extinction probability, see Probability
- Extinction rate (E, μ_s)
area effect and, 177
MacArthur-Wilson model, 177
metapopulation model, 93-94
nonlinear, 182-183
target effect and, 184-185
- f*, see Population sites
- F_0 , see Fertility
- Ferminia cerasini*, 175
- Fertility, of age class i (F_i), 66-67, 68
- Fertility schedule $f(x)$, 57-59
- Leslie matrix and, 68
- life-table calculations and, 62
life history strategies and, 75
- Finite rate of increase (λ), 11-13, 70-71
compared to net reproductive rate, 63
- Leslie matrix and, 68, 79
transition matrix and, 79
- Fish, metapopulation studies, 103-106
- Fishing industry, optimal yield and, 47-49
- Florida Keys, island colonization experi-
ments, 187-188, 189
- Flour beetles, see *Tribolium*
- Functional response, of predator populations,
141, 149-153
- G*, see Generation time
- $g(x)$, see Survival probability
- Generation time (G), 58, 63
- Gerbils (*Citellus allenthyi*, *C. pyramidum*),
competition studies, 132-135
- Grizzly bears, 20-22
- Ground squirrels, 80-82
- Grouse, population cycles, 162-166
- Growth rate (dN/dt)
exponential model, 4-6, 33, 34
isocline, 118-119, 120
logistic model, 28-34
per capita $[(1/N)(dN/dt)]$, 33-34
- Gunnerside (England), grouse population,
162-166
- Habitat availability, competition studies and,
134-135
- Habitat diversity, species-area relationship
and, 174-175
- Half-saturation constant, 151
- Handling time, 149-150
- Hare, population cycles, 161-162
HIV, 166
- Hobsonia floridæ*, 128-132
- Horizontal life table, 72
- Hutchinson, G. E., 132
- Hydra*, doubling time, 8
- I*, see Immigration rate; Immigrants

- reproductive value and, 71, 74-75
for stage-structured growth (transition matrix), 77-79, 82-85
- Leslie, P. H., 67
- Life history strategies, 75-76
- loop diagrams, 78-79, 83
- Life-table analysis, 57-71
- fertility, 66-67, 68
- life history schedule, 57-59, 72
- generation time, 63
- of ground squirrels, 80-82
- horizontal vs. vertical, 72
- intrinsic rate of increase, 64-65
- net reproductive rate, 62-63
- survival probability, 66, 68
- survivorship schedule, 58, 59-62, 68, 72
- table of standard calculations, 58
- of teal, 82-85
- Linear density dependence, 33, 34
- Logarithm (base 10), 20
- species-area relationship and, 172-173
- see also* Natural logarithm (base e)
- Logistic growth model, 28-32
- assumptions underlying, 32-33
- vs. exponential growth model, 33-34
- r - K selection and, 76
- Lotka, A. J., 113, 140
- Lotka-Volterra competition model, 113-114
- algebraic solutions, 124-126
- assumptions underlying, 126-127
- case 1, 119-120, 121, 125
- case 2, 120-122, 125
- case 3, 122-123, 125
- case 4, 123-124, 125
- competition coefficients, 114
- equilibrium solutions, 115-116
- graphical solutions, *see* State-space graph
- for intraguild predation, 127-130
- assumptions underlying, 147
- cycles, 145-147
- graphical solutions, *see* State-space graph
- Loxia leucoptera*, 175
- Lynx canadensis*, 161-162
- MacArthur, R. H., 175
- MacArthur-Wilson model, 175-182
- assumptions underlying, 181-182
- Malthus, Thomas Robert, 5, 11
- Malthusian parameter, *see* Instantaneous rate of increase
- Mandarte Island, sparrow population, 43-45
- Mangrove, doubling time, 8
- Mangrove islands, colonization experiments, 187-188, 189
- Marine worms, competition studies, 128-132
- Maternity schedule [$m(x)$], *see* Fertility schedule
- Matrix, *see* Leslie matrix
- Maximum sustained yield, *see* Optimal yield
- Melospiza melodia*, 43-45
- Metapopulation
- defined, 90-91
- dynamics, 93-96
- extinction risk, 91-92, 93
- internal colonization model, 97-98, 100-101
- island-mainland model, 96-97
- rescue effect, 98-101
- table of models, 100
- Migration, population growth and, 90
- Modular organism, size-structured analysis, 77-79
- Monocarpic reproduction, 59
- Morgan Hill, metapopulation studies, 101-102
- Mortality
- density-dependent vs. density-independent, 43-47
- net reproductive rate and, 63
- see also* Death rate
- Mutualism, defined, 127
- n , *see* Age structure
- N , *see* Population size
- Natural logarithm (ln)
- base e defined, 6
- doubling time and, 6, 8
- see also* Logarithm
- Natural selection, 11
- life history strategies and, 75
- Negev Desert, competition studies, 132-135
- Nematode worms, population cycle studies, 162-166
- Netherlands, metapopulation studies, 103, 104
- Net reproductive rate (R_0), 58, 62-63
- Neutral equilibrium, 100
- Neutrally stable cycles, 146
- New Guinea, 174
- Notifagus fuscus*, doubling time, 8
- Notropis girardi*, 106
- Oklahoma, fish metapopulation study, 103-106
- Oligochaete worms, competition studies, 128-132
- Optimal yield
- for commercial fishing, 47-49
- reproductive value and, 74-75
- Oscillations, in population growth, 35-37
- Overexploitation, 47-49
- P*, *see* Predator population; Probability
- oscillations in, 35-37
- projecting, 6-9
- stable equilibrium ($N = K$), 31
- Postbreeding census, 66
- Predation, defined, 127
- Predator population (P)
- cycles of, 145-147
- equilibrium, 142-143
- growth of, 141-142
- parasites, 162-166
- Pre-emptive competition, 112
- Prey population, *see* Victim population
- Probability (P)
- demographic stochasticity and, 16-17
- of local colonization (p_i), 94-96, 98-100
- of local extinction (p_e), 91-92, 93, 95-96, 98-100
- passive sampling model and, 186-187
- of regional persistence, 93
- of survival to age x [$g(x)$], 58, 60-62
- of survival within age class i (P_i), 66-68
- Propagule rain, 96, 100
- rescue effect and, 98-101
- Protection Island, pheasant population, 19-20
- Pterostichus versicolor*, 103, 104
- Puget Sound, competition studies, 128-132
- Pymatuning Lake islands, passive sampling model study, 190-192
- r*, *see* Instantaneous rate of increase
- r selection, 76
- R_0 , *see* Net reproductive rate
- Random variation
- in carrying capacity, 40, 41
- Rats, doubling time, 8
- Red grouse, population cycles, 162-166
- Refuge effect, 156-157
- Reproduction
- iteroparous, 59, 72
- types of, 59
- Reproductive rate, *see* Net reproductive rate
- Reproductive value [$v(x)$], 71, 73-75
- distribution of, 71, 74, 84
- transition matrix and, 79
- Rescue effect, 98-100, 184
- Resource limitation
- competitive interactions and, 112
- Lotka-Volterra competition model and, 126
- prey population as, 140
- Rhizophora mangle*, 187
- S*, *see* Species
- $S(x)$, *see* Survivors
- Sea squirrels, population fluctuations, 44-47
- Search time, 150
- P*, *see* Survival probability
- Paradox of enrichment, 154, 156
- Paramecium caudatum*, doubling time, 8
- Parasitism
- defined, 127
- in predator-prey cycles, 162-166
- Passive sampling model, 185-187
- Per capita growth rate, $[(1/N)(dN/dt)]$, 33-34
- Perennial plants, 59
- Period, of population cycle, 35-36
- Periodic variation, in carrying capacity, 40-42
- Persistence, metapopulation dynamics and, 90-93
- Peru, anchovy fisheries, 47-49
- Plusminus colchicus torquatus*, 19-20
- Pheasants, 19-20
- Plantago erecta*, 101
- Polycarpic reproduction, 59
- Population(s)
- age structure, 65-67
- closed, 3, 9, 90
- cyclic, 35-37; *see also* Cycles
- defined, 2
- migration and, 90
- persistence of, 90-93
- source vs. sink, 103
- see also* Metapopulation; Predator population; Victim population
- Population density
- ground squirrel study, 80-82
- growth rate and, 28-29
- r - K selection and, 76
- see also* Carrying capacity
- Population growth
- age-structured, 56-65
- discrete, 11-13
- exponential model, 5-11
- factors affecting, 2-6
- logistic model, 28-32
- of predator population, 141-142
- of prey (victim) population, 140-141, 147
- rate of, *see* Growth rate; Instantaneous rate of increase
- stage- or size-structured, 77-79
- time lags and, 4
- Population management, reproductive value and, 75
- see also* Optimal yield
- Population sites, fraction colonized (f), 93-101
- equilibrium value, 95
- internal colonization model, 97-98, 100-101
- island-mainland model, 96
- rescue effect, 98-101
- Population size (N), 2-6
- cycles in, *see* Cycles
- doubling time, 6, 8

- Semelparous reproduction, 59
 Senescence, 75
 Serpentine soil, 101–102
 σ (sigma), *see* Variance
 Simberloff, D., 187–188
 Sink population, 103
 Size-structured population growth, 77–79
 Snowshoe hare, population cycles, 161–162
 Source population, 103
 Sparrows, population fluctuations, 43–45
 Species, number of (S)
 equilibrium, 178–179
 species–area relationship, 172–174
 Species–area relationship, 172–174
 passive sampling model and, 185–187
see also MacArthur–Wilson model
 Species density, 179
 Species introductions, 19–20
Spermophilus arnatus, 80–82
 Stability, of population cycles, 159–160
 Stable age distribution, 68–76, 79
 Stable equilibrium, 31
 competitive coexistence in, 122–123, 125
 Stable limit cycles, 35–39
 Stage-structured population growth, 77–79
 teasel study, 82–85
 State-space graph
 defined, 116–117
 of gerbil competition, 133, 135
 of Lotka–Volterra competition model,
 118–123
 of Lotka–Volterra predation model,
 143–145, 148, 153, 154–160
 of marine worm competition, 131
 Static life table, 72
 Stationary age distribution 70
 Stochasticity, *see* Demographic stochasticity;
 Environmental stochasticity; Random
 variation
 Sunspots, population cycles and, 161, 162
 Surrey (England), immigration/extinction
 study, 188–190
 Survival probability
 to age x [$l_x(x)$], 58, 60–62
 within age class i (P_i), 66, 68
 Survivors, number of [$S(x)$], 58, 59–60
 Survivorship schedule [$l(x)$], 58, 59–61
 Leslie matrix and, 68
 life history strategies and, 75
 life-table calculations and, 62
 types of (curves), 61–62
T. see Turnover rate
 T phage, doubling time, 8
 Target effect, 184–185
 τ (tau), *see* Time lag
 Teasel, 82–85
 Territoriality, population fluctuation and,
 44–45
 Time lag (τ)
 exponential model and, 4, 10
 logistic model and, 34–39
 metapopulation model and, 95
 Tradeoffs, in life history strategies, 75–76
 Transition matrix, 77–79
 teasel study, 82–85
see also Leslie matrix
 Trees
 density-dependent responses, 34
 doubling time, 8
 size-structured model, 78, 79
Tribolium, intraguild predation, 127
Tribolium castaneum, doubling time, 8
Trichostrogylus tenuis, 162–166
 Turnover rate (T), MacArthur–Wilson model,
 178–179
 Uinta ground squirrels, 80–82
 Unstable equilibrium, competitive exclusion
 and, 123–124, 125
Ursus arctos horribilis, 20–22
 $r(x)$, *see* Reproductive value
V. see Victim population
 Variance (σ)
 in carrying capacity, 40–42
 stochasticity and, 14–16, 18
 Verhulst, P.-F., 30
 Vertical life table, 72
 Victim population
 carrying capacity, 148–149
 cycles of, 145–147
 equilibrium, 142–143
 growth of, 140–141, 147
 refuge effect, 156–157
 Virus, doubling time, 8
 Volterra, V., 113, 140
 Warblers, 90
 West Indies, number of bird species, 172–175
 White-winged crossbill, 175
 Wilson, E. O., 175, 187
 Worms
 competition studies, 128–132
 nematode, 162–166
 Yellowstone National Park, grizzly bear pop-
 ulation, 20–22
 Zapata wren, 175